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(54) Title: HUMAN CALCIUM CHANNEL COMPOSITIONS AND METHODS USING THEM

(57) Abstract

Isolated DNA encoding each of human calcium chanel α_1 -, α_2 -, β - and γ -subunits, including subunits that arise as splice variants of primary transcripts, is provided. In particular DNA clones encoding each of the α_{1A-1} , α_{1A-2} , α_{1E-1} , α_{1C-2} , α_{1E-3} , β_{3-1} , β_{2C} , β_{2D} , β_{2E} and β_4 subunits of human calcium channels are provided. Cells and vectors containing the DNA, subunit specific antibodies and nucleic acid probes and methods for identifying compounds that modulate the activity of human calcium channels are also provided.

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HUMAN CALCIUM CHANNEL COMPOSITIONS AND METHODS USING THEM

TECHNICAL FIELD

The present invention relates to molecular biology and pharmacology. More particularly, the invention relates to calcium channel compositions and methods of making and using the same.

BACKGROUND OF THE INVENTION

Calcium channels are membrane-spanning, multi-subunit proteins that allow controlled entry of Ca²⁺ ions into cells from the extracellular fluid. Cells throughout the animal kingdom, and at least some bacterial, fungal and plant cells, possess one or more types of calcium channel.

The most common type of calcium channel is voltage dependent. "Opening" of a voltage-dependent channel to allow an influx of Ca²⁺ ions into the cells requires a depolarization to a certain level of the potential difference between the inside of the cell bearing the channel and the extracellular medium bathing the cell. The rate of influx of Ca²⁺ into the cell depends on this potential difference. All "excitable" cells in animals, such as neurons of the central nervous system (CNS), peripheral nerve cells and muscle cells, including those of skeletal muscles, cardiac muscles, and venous and arterial smooth muscles, have voltage-dependent calcium channels.

Multiple types of calcium channels have been identified in mammalian cells from various tissues, including skeletal muscle, cardiac muscle, lung, smooth muscle and brain, [see, e.g., Bean, B.P. (1989) Ann. Rev. Physiol. 51:367-384 and Hess, P. (1990) Ann. Rev. Neurosci. 56:337]. The different types of calcium channels have been broadly categorized into four classes, L-, T-, N-, and P-type, distinguished by current kinetics, holding potential sensitivity and sensitivity to calcium channel agonists and antagonists.

Calcium channels are multisubunit proteins that contain two large subunits, designated α_1 and α_2 , which have molecular weights between about 130 and about 200 kilodaltons ("kD"),

and one to three different smaller subunits of less than about 60 kD in molecular weight. At least one of the larger subunits and possibly some of the smaller subunits are glycosylated. Some of the subunits are capable of being phosphorylated. The α_i subunit has a molecular weight of about to about 170 kD when analyzed by dodecylsulfate (SDS)-polyacrylamide gel electrophoresis (PAGE) after isolation from mammalian muscle tissue and has specific binding sites for various 1,4-dihydropyridines (DHPs) phenylalkylamines. Under non-reducing conditions (in the presence of N-ethylmaleimide), the α_2 subunit migrates in SDS-PAGE as a band corresponding to a molecular weight of about 160-190 kD. Upon reduction, a large fragment and smaller fragments are released. The β subunit of the rabbit skeletal muscle calcium channel is a phosphorylated protein that has a molecular weight of 52-65 kD as determined by SDS-PAGE analysis. This subunit is insensitive to reducing conditions. The γ subunit of the calcium channel, which is not observed in all purified preparations, appears to be a glycoprotein with an apparent molecular weight of 30-33 kD, as determined by SDS-PAGE analysis.

In order to study calcium channel structure and function, large amounts of pure channel protein are needed. Because of the complex nature of these multisubunit proteins, the varying concentrations of calcium channels in tissue sources of the protein, the presence of mixed populations of calcium channels in tissues, difficulties in obtaining tissues of interest, and the modifications of the native protein that can occur during the isolation procedure, it is extremely difficult to obtain large amounts of highly purified, completely intact calcium channel protein.

Characterization of a particular type of calcium channel by analysis of whole cells is severely restricted by the presence of mixed populations of different types of calcium channels in the majority of cells. Single-channel recording methods that are used to examine individual calcium channels

do not reveal any information regarding the molecular structure or biochemical composition of the channel. Furthermore, in performing this type of analysis, the channel is isolated from other cellular constituents that might be important for natural functions and pharmacological interactions.

Characterization of the gene or genes encoding calcium channels provides another means of characterization of different types of calcium channels. The amino acid sequence determined from a complete nucleotide sequence of the coding region of a gene encoding a calcium channel protein represents the primary structure of the protein. Furthermore, secondary structure of the calcium channel protein and the relationship of the protein to the membrane may be predicted based on analysis of the primary structure. For instance, hydropathy plots of the α_1 subunit protein of the rabbit skeletal muscle calcium channel indicate that it contains four internal repeats, each containing six putative transmembrane regions [Tanabe, T. et al. (1987) Nature 328:313].

Because calcium channels are present in various tissues and have a central role in regulating intracellular calcium ion concentrations, they are implicated in a number of vital processes in animals, including neurotransmitter release, muscle contraction, pacemaker activity, and secretion of These processes appear to be hormones and other substances. involved in numerous human disorders, such as CNS Calcium channels, thus, are also cardiovascular diseases. implicated in numerous disorders. A number of compounds useful for treating various cardiovascular diseases are thought to exert their including humans, animals, effects by modulating functions of voltagebeneficial dependent calcium channels present in cardiac and/or vascular Many of these compounds bind to calcium smooth muscle. channels and block, or reduce the rate of, influx of Ca2+ into the cells in response to depolarization of the cell membrane.

The results of studies of recombinant expression of rabbit calcium channel α_1 subunit-encoding cDNA clones and transcripts of the cDNA clones indicate that the α_1 subunit forms the pore through which calcium enters cells. The relevance of the barium currents generated in these recombinant cells to the actual current generated by calcium channels containing as one component the respective α_1 subunits in vivo is unclear. In order to completely and accurately characterize and evaluate different calcium channel types, however, it is essential to examine the functional properties of recombinant channels containing all of the subunits as found in vivo.

In order to conduct this examination and to fully understand calcium channel structure and function, it is critical to identify and characterize as many calcium channel subunits as possible. Also in order to prepare recombinant cells for use in identifying compounds that interact with calcium channels, it is necessary to be able to produce cells that express uniform populations of calcium channels containing defined subunits.

An understanding of the pharmacology of compounds that interact with calcium channels in other organ systems, such as the CNS, may aid in the rational design of compounds that specifically interact with subtypes of human calcium channels to have desired therapeutic effects, such as in the treatment of neurodegenerative and cardiovascular disorders. understanding to and the ability rationally design therapeutically effective compounds, however, have hampered by an inability to independently determine the types human calcium channels and the molecular nature of individual subtypes, particularly in the CNS, and by the unavailability of pure preparations of specific channel subtypes to use for evaluation of the specificity of calcium channel-effecting compounds. Thus, identification of DNA encoding human calcium channel subunits and the use of such DNA for expression of calcium channel subunits and functional

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calcium channels would aid in screening and designing therapeutically effective compounds.

Therefore, it is an object herein, to provide DNA encoding specific calcium channel subunits and to provide eukaryotic cells bearing recombinant tissue-specific or subtype- specific calcium channels. It is also an object to provide assays for identification of potentially therapeutic compounds that act as calcium channel antagonists and agonists.

SUMMARY OF THE INVENTION

Isolated and purified nucleic acid fragments that encode human calcium channel subunits are provided. DNA encoding α_1 subunits of a human calcium channel, and RNA, encoding such subunits, made upon transcription of such DNA are provided. In particular, DNA fragments encoding α_1 subunits of voltage-dependent human calcium channels (VDCCs) type A, type B (also referred to as VDCC IV), type C (also referred to as VDCC II) type D (also referred to as VDCC III) and type E are provided.

DNA encoding α_{1A} , α_{1B} , α_{1C} , α_{1D} and α_{1E} subunits is provided. DNA encoding an α_{1D} subunit that includes the amino acids substantially as set forth as residues 10-2161 of SEQ ID No. 1 is provided. DNA encoding an α_{1D} subunit that includes substantially the amino acids set forth as amino acids 1-34 in SEQ ID No. 2 in place of amino acids 373-406 of SEQ ID No. 1 is also provided. DNA encoding an α_{1C} subunit that includes the amino acids substantially as set forth in SEQ ID No. 3 or SEQ ID No. 6 and DNA encoding an α_{1B} subunit that includes an amino acid sequence substantially as set forth in SEQ ID No. 7 or in SEQ ID No. 8 is also provided.

DNA encoding $\alpha_{\mathtt{lA}}$ subunits is also provided. Such DNA includes DNA encoding an α_{la} subunit that has substantially the same sequence of amino acids as encoded by the DNA set forth in SEQ ID No. 22 or No. 23 or other splice variants of α_{1A} that include all or part of the sequence set forth in SEQ ID No. 22 The sequence set forth in SEQ ID NO. 22 is a splice variant designated $\alpha_{\text{lA-1}}$; and the sequence set forth in SEQ ID NO. 23 is a splice variant designated α_{1A-2} . DNA encoding α_{1A} subunits also include DNA encoding subunits that can be isolated using all or a portion of the DNA having SEQ ID NO. 21, 22 or 23 or DNA obtained from the phage lysate of an E. coli host containing DNA encoding an α_{iA} subunit that has been deposited in the American Type Culture Collection, 12301 Parklawn Drive, Rockville, Maryland 20852 U.S.A. Accession No. 75293 in accord with the Budapest Treaty.

DNA in such phage includes a DNA fragment having the sequence set forth in SEQ ID No. 21. This fragment selectively hybridizes under conditions of high stringency to DNA encoding α_{1A} but not to DNA encoding α_{1B} and, thus, can be used to isolate DNA that encodes α_{1A} subunits.

DNA encoding $\alpha_{\rm IE}$ subunits of a human calcium channel is also provided. This DNA includes DNA that encodes an $\alpha_{\rm IE}$ splice variant designated $\alpha_{\rm IE-1}$ encoded by the DNA set forth in SEQ ID No. 24, and a variant designated $\alpha_{\rm IE-3}$ encoded by SEQ ID No. 25. This DNA also includes other splice variants thereof that encodes sequences of amino acids encoded by all or a portion of the sequences of nucleotides set forth in SEQ ID Nos. 24 and 25 and DNA that hybridizes under conditions of high stringency to the DNA of SEQ ID. No. 24 or 25 and that encodes an $\alpha_{\rm IE}$ splice variant.

DNA encoding α_2 subunits of a human calcium channel, and RNA encoding such subunits, made upon transcription of such a DNA are provided. DNA encoding splice variants of the α_2 subunit, including tissue-specific splice variants, are also provided. In particular, DNA encoding the α_{2a} - α_{2e} subunit subtypes is provided. In particularly preferred embodiments, the DNA encoding the α_2 subunit that is produced by alternative processing of a primary transcript that includes DNA encoding the amino acids set forth in SEQ ID 11 and the DNA of SEQ ID No. 13 inserted between nucleotides 1624 and 1625 of SEQ ID No. 11 is provided. The DNA and amino acid sequences of α_{2a} - α_{2e} are set forth in SEQ. ID Nos. 11 and 29-32, respectively.

Isolated and purified DNA fragments encoding human calcium channel β subunits, including DNA encoding β_1 , β_2 , β_3 and β_4 subunits, and splice variants of the β subunits are provided. RNA encoding β subunits, made upon transcription of the DNA is also provided.

DNA encoding a β_1 subunit that is produced by alternative processing of a primary transcript that includes DNA encoding the amino acids set forth in SEQ ID No. 9, but including the

DNA set forth in SEQ ID No. 12 inserted in place of nucleotides 615-781 of SEQ ID No. 9 is also provided. DNA encoding β_1 subunits that are encoded by transcripts that have the sequence set forth in SEQ ID No. 9 including the DNA set forth in SEQ ID No. 12 inserted in place of nucleotides 615-781 of SEQ ID No. 9, but that lack one or more of the following sequences of nucleotides: nucleotides 14-34 of SEQ ID No. 12, nucleotides 13-34 of SEQ ID No. 12, nucleotides 35-55 of SEQ ID No 12, nucleotides 56-190 of SEQ ID No. 12 and nucleotides 191-271 of SEQ ID No. 12 are also provided. In particular, β_1 subunit splice variants β_{1-1} - β_{1-5} (see, SEQ ID Nos. 9, 10 and 33-35) described below, are provided.

 B_2 subunit splice variants β_{2c} - β_{2e} , that include all or a portion of SEQ ID Nos. 26, 29 and 30 are provided; β_3 subunit splice variants, including β_3 subunit splice variants that have the sequences set forth in SEQ ID Nos 19 and 20, and DNA encoding the β_4 subunit that includes DNA having the sequence set forth in SEQ ID No. 27 and the amino acid sequence set forth in SEQ ID No. 28 are provided.

Also Escherichia coli (E. coli) host cells harboring plasmids containing DNA encoding β_3 have been deposited in accord with the Budapest Treaty under Accession No. 69048 at the American Type Culture Collection. The deposited clone encompasses nucleotides 122-457 in SEQ ID No. 19 and 107-443 in SEQ ID No. 20.

DNA encoding β subunits that are produced by alternative processing of a primary transcript encoding a β subunit, including a transcript that includes DNA encoding the amino acids set forth in SEQ ID No. 9 or including a primary transcript that encodes β_3 as deposited under ATCC Accession No. 69048, but lacking and including alternative exons are provided or may be constructed from the DNA provided herein.

DNA encoding γ subunits of human calcium channels is also provided. RNA, encoding γ subunits, made upon transcription of the DNA are also provided. In particular, DNA containing

the sequence of nucleotides set forth in SEQ ID No. 14 is provided.

Full-length DNA clones and corresponding RNA transcripts, encoding α_1 , including splice variants of α_{1A} , α_{1D} , α_{1B} , α_{1C} , and α_{1E} , α_2 and β subunits, including β_{1-1} - β_{1-5} , β_{2C} , β_{2D} , β_{2E} , β_{3-1} and β_4 of human calcium channels are provided. Also provided are DNA clones encoding a substantial portions of the certain α_{1C} subtype subunits and γ subunits of voltage-dependent human calcium channels for the preparation of full-length DNA clones encoding the corresponding full-length subunits. Full-length clones may be readily obtained using the disclosed DNA as a probe as described herein.

The the α_{1A} subunit, α_{1C} subunit, α_{1E} subunit and splice variants thereof, the β_{2D} , β_{2C} and β_{2E} subunits and β_4 subunits and nucleic acids encoding these subunits are of particular interest herein.

Eukaryotic cells containing heterologous DNA encoding one or more calcium channel subunits, particularly human calcium channel subunits, or containing RNA transcripts of DNA clones encoding one or more of the subunits are provided. A single α_1 subunit can form a channel. The requisite combination of subunits for formation of active channels in selected cells, however, can be determined empirically using the methods herein. For example, if a selected α_1 subtype or variant does not form an active channel in a selected cell line, an additional subunit or subunits can be added until an active channel is formed.

In preferred embodiments, the cells contain DNA or RNA encoding a human α_1 subunit, preferably at least an α_{1D} , α_{1B} , α_{1A} or α_{1E} subunit. In more preferred embodiments, the cells contain DNA or RNA encoding additional heterologous subunits, including at least one β , α_2 or γ subunit. In such embodiments, eukaryotic cells stably or transiently transfected with any combination of one, two, three or four of the subunit-encoding DNA clones, such as DNA encoding any of α_1 , α_1 + β , α_1 + β + α_2 , are provided.

The eukaryotic cells provided herein contain heterologous DNA that encodes an α_1 subunit or heterologous encodes an $lpha_1$ subunit and heterologous DNA that encodes a etasubunit. At least one subunit is selected α_{1A-1} , α_{1A-2} , α_{1c-2} , $\alpha_{\text{1E-1}},~\alpha_{\text{1E-3}},~\beta_{\text{2C}},~\beta_{\text{2D}},~\beta_{\text{2F}},~\text{a}~\beta_{\text{3-1}},~\beta_{\text{3-2}}$ subunit or a β_4 subunit. In preferred embodiments, the cells express such heterologous calcium channel subunits and include one or more of the subunits in membrane-spanning heterologous calcium channels. In more preferred embodiments, the eukaryotic cells express functional, heterologous calcium channels that are capable of gating the passage of calcium channel-selective ions and/or binding compounds that, at physiological concentrations, modulate the activity of the heterologous calcium channel. In certain embodiments, the heterologous calcium channels include at least one heterologous calcium channel subunit. preferred embodiments, the calcium channels that are expressed surface of the eukaryotic cells are substantially or entirely of subunits encoded heterologous DNA or RNA. In preferred embodiments, the heterologous calcium channels of such cells are distinguishable from any endogenous calcium channels of the Such cells provide a means to obtain homogeneous host cell. populations of calcium channels. Typically, the cells contain the selected calcium channel as the only heterologous ion channel expressed by the cell.

In certain embodiments the recombinant eukaryotic cells that contain the heterologous DNA encoding the calcium channel subunits are produced by transfection with DNA encoding one or more of the subunits or are injected with RNA transcripts of DNA encoding one or more of the calcium channel subunits. The DNA may be introduced as a linear DNA fragment or may be included in an expression vector for stable or transient expression of the subunit-encoding DNA. Vectors containing DNA encoding human calcium channel subunits are also provided.

The eukaryotic cells that express heterologous calcium channels may be used in assays for calcium channel function or, in the case of cells transformed with fewer subunit-encoding nucleic acids than necessary to constitute a functional recombinant human calcium channel, such cells may be used to assess the effects of additional subunits on calcium channel activity. The additional subunits can be provided by subsequently transfecting such a cell with one or more DNA clones or RNA transcripts encoding human calcium channel subunits.

The recombinant eukaryotic cells that express membrane spanning heterologous calcium channels may be used in methods for identifying compounds that modulate calcium channel activity. In particular, the cells are used in assays that identify agonists and antagonists of calcium channel activity in humans and/or assessing the contribution of the various calcium channel subunits to the transport and regulation of transport of calcium ions. Because the cells constitute homogeneous populations of calcium channels, they provide a means to identify agonists or antagonists of calcium channel activity that are specific for each such population.

The assays that use the eukaryotic cells for identifying compounds that modulate calcium channel activity are also provided. In practicing these assays the eukaryotic cell that expresses a heterologous calcium channel, containing at least on subunit encoded by the DNA provided herein, is in a solution containing a test compound and a calcium channel selective ion, the cell membrane is depolarized, and current flowing into the cell is detected. If the test compound is one that modulates calcium channel activity, the current that is detected is different from that produced by depolarizing the same or a substantially identical cell in the presence of the same calcium channel-selective ion but in the absence of In preferred embodiments, prior to the the compound. depolarization step, the cell is maintained at a holding potential which substantially inactivates calcium channels which are endogenous to the cell. Also in preferred embodiments, the cells are mammalian cells, most preferably HEK cells, or amphibian occytes.

Nucleic acid probes, typically labeled for detection, containing at least about 14, preferably 16, or, if desired, 20 or 30 or more, contiguous nucleotides of α_{1D} , α_{1C} , α_{1B} , α_{1A} and α_{1E} , α_{2} , β , including β_{1} , β_{2} , β_{3} and β_{4} splice variants and γ subunit-encoding DNA are provided. Methods using the probes for the isolation and cloning of calcium channel subunit-encoding DNA, including splice variants within tissues and inter-tissue variants are also provided.

Purified human calcium channel subunits and purified human calcium channels are provided. The subunits and channels can be isolated from a eukaryotic cell transfected with DNA that encodes the subunit.

In another embodiment, immunoglobulins or antibodies obtained from the serum of an animal immunized with a substantially pure preparation of a human calcium channel, human calcium channel subunit or epitope-containing fragment a human calcium subunit are provided. Monoclonal antibodies produced using a human calcium channel, human calcium channel subunit or epitope-containing fragment thereof as an immunogen are also provided. E. coli fusion proteins including a fragment of a human calcium channel subunit may also be used as immunogen. Such fusion proteins may contain a bacterial protein or portion thereof, such as the E. coli TrpE protein, fused to a calcium channel subunit peptide. immunoglobulins that are produced using the calcium channel subunits or purified calcium channels as immunogens have, among other properties, the ability to specifically and preferentially bind to and/or cause the immunoprecipitation of a human calcium channel or a subunit thereof which may be present in a biological sample or a solution derived from such a biological sample. Such antibodies may also be used to selectively isolate cells that express calcium channels that contain the subunit for which the antibodies are specific.

Methods for modulating the activity of ion channels by contacting the calcium channels with an effective amount of the above-described antibodies are also provided.

A diagnostic method for determining the presence of Lambert Eaton Syndrome (LES) in a human based on immunological reactivity of LES immunoglobulin G (IgG) with a human calcium channel subunit or a eukaryotic cell which expresses a recombinant human calcium channel or a subunit thereof is also provided. In particular, an immunoassay method for diagnosing Lambert-Eaton Syndrome in a person by combining serum or an IgG fraction from the person (test serum) with calcium channel proteins, including the α and β subunits, and ascertaining whether antibodies in the test serum react with one or more of the subunits, or a recombinant cell which expresses one or more of the subunits to a greater extent than antibodies in control serum, obtained from a person or group of persons known to be free of the Syndrome, is provided. immunoassay procedure known in the art for antibodies against a given antigen in serum can be employed in the method.

DETAILED DESCRIPTION OF THE INVENTION Definitions:

Unless defined otherwise, all technical and scientific terms used herein have the same meaning as is commonly understood by one of skill in the art to which this invention belongs. All patents and publications referred to herein are incorporated by reference herein.

Reference to each of the calcium channel subunits includes the subunits that are specifically disclosed herein and human calcium channel subunits encoded by DNA that can be isolated by using the DNA disclosed as probes and screening an appropriate human cDNA or genomic library under at least low stringency. Such DNA also includes DNA that encodes proteins that have about 40% homology to any of the subunits proteins described herein or DNA that hybridizes under conditions of at least low stringency to the DNA provided herein and the

protein encoded by such DNA exhibits additional identifying characteristics, such as function or molecular weight.

It is understood that subunits that are encoded by transcripts that represent splice variants of the disclosed subunits or other such subunits may exhibit less than 40% overall homology to any single subunit, but will include regions of such homology to one or more such subunits. It is also understood that 40% homology refers to proteins that share approximately 40% of their amino acids in common or that share somewhat less, but include conservative amino acid substitutions, whereby the activity of the protein is not substantially altered.

As used herein, the α_1 subunits types, encoded by different genes, are designated as type α_{1A} , α_{1B} , α_{1C} , α_{1D} and α_{1E} . These types have also been referred to as VDCC IV for α_{1B} , VDCC II for α_{1C} and VDCC III for α_{1D} . Subunit subtypes, which are splice variants, are referred to, for example as α_{1B-1} , α_{1B} . α_{1C-1} etc.

Thus, as used herein, DNA encoding the α_1 subunit refers to DNA that hybridizes to the DNA provided herein under conditions of at least low stringency or encodes a subunit that has at least about 40% homology to protein encoded by DNA disclosed herein that encodes an α_1 subunit of a human calcium. An α_1 subunit may be identified by its ability to form a calcium channel. Typically, α_1 subunits have molecular masses greater than at least about 120 kD. Also, hydropathy plots of deduced α_1 subunit amino acid sequences indicate that the α_1 subunits contain four internal repeats, each containing six putative transmembrane domains.

The activity of a calcium channel may be assessed in vitro by methods known to those of skill in the art, including the electrophysiological and other methods described herein. Typically, α_1 subunits include regions to which one or more modulators of calcium channel activity, such as a 1,4-DHP or ω -CgTx, interact directly or indirectly. Types of α_1 subunits may be distinguished by any method known to those of skill in

the art, including on the basis of binding specificity. example, it has been found herein that α_{1B} subunits participate in the formation channels that have previously been referred to as N-type channels, $\alpha_{\text{\tiny 1D}}$ subunits participate in the formation of channels that had previously been referred to as L-type channels, and α_{1A} subunits appear to participate in the formation of channels that exhibit characteristics typical of channels that had previously been designated P-type channels. Thus, for example, the activity of channels that contain the α_{1B} subunit are insensitive to 1,4-DHPs; whereas the activity of channels that contain the α_{1D} subunit are modulated or altered by a 1,4-DHP. It is presently preferable to refer to calcium channels based on pharmacological characteristics and current kinetics and to avoid historical designations. Types and subtypes of α_1 subunits may be characterized on the basis of the effects of such modulators on the subunit or a channel containing the subunit as well as differences in currents and current kinetics produced by calcium channels containing the subunit.

As used herein, an α_2 subunit is encoded by DNA that hybridizes to the DNA provided herein under conditions of low stringency or encodes a protein that has at least about 40% homology with that disclosed herein. Such DNA encodes a protein that typically has a molecular mass greater than about 120 kD, but does not form a calcium channel in the absence of an α_1 subunit, and may alter the activity of a calcium channel that contains an α_1 subunit. Subtypes of the α_2 subunit that arise as splice variants are designated by lower case letter, such as α_{2a} , . . . α_{2e} . In addition, the α_2 subunit and the large fragment produced when the protein is subjected to reducing conditions appear to be glycosylated with at least N-linked sugars and do not specifically bind to the 1,4-DHPs and phenylalkylamines that specifically bind to the α , subunit. The smaller fragment, the C-terminal fragment, is referred to as the δ subunit and includes amino acids from about 946 (SEQ ID No. 11) through about the C-terminus. This

fragment may dissociate from the remaining portion of α_2 when the α_2 subunit is exposed to reducing conditions.

As used herein, a β subunit is encoded by DNA that hybridizes to the DNA provided herein under conditions of low stringency or encodes a protein that has at least about 40% homology with that disclosed herein and is a protein that typically has a molecular mass lower than the α subunits and on the order of about 50-80 kD, does not form a detectable calcium channel in the absence of an α_1 subunit, but may alter the activity of a calcium channel that contains an α_1 subunit or that contains an α_1 and α_2 subunit.

Types of the β subunit that are encoded by different genes are designated with subscripts, such as β_1 , β_2 , β_3 and β_4 . Subtypes of β subunits that arise as splice variants of a particular type are designated with a numerical subscript referring to the type and to the variant. Such subtypes include, but are not limited to the β_1 splice variants, including β_{1-1} - β_{1-5} and β_2 variants, including β_{2C} - β_{2E} .

As used herein, a γ subunit is a subunit encoded by DNA disclosed herein as encoding the γ subunit and may be isolated and identified using the DNA disclosed herein as a probe by hybridization or other such method known to those of skill in the art, whereby full-length clones encoding a γ subunit may be isolated or constructed. A γ subunit will be encoded by DNA that hybridizes to the DNA provided herein under conditions of low stringency or exhibits sufficient sequence homology to encode a protein that has at least about 40% homology with the γ subunit described herein.

Thus, one of skill in the art, in light of the disclosure herein, can identify DNA encoding α_1 , α_2 , β , δ and γ calcium channel subunits, including types encoded by different genes and subtypes that represent splice variants. For example, DNA probes based on the DNA disclosed herein may be used to screen an appropriate library, including a genomic or cDNA library, for hybridization to the probe and obtain DNA in one or more clones that includes an open reading fragment that

encodes an entire protein. Subsequent to screening an appropriate library with the DNA disclosed herein, the isolated DNA can be examined for the presence of an open reading frame from which the sequence of the encoded protein may be deduced. Determination of the molecular weight and comparison with the sequences herein should reveal the identity of the subunit as an α_1 , α_2 etc. subunit. Functional assays may, if necessary, be used to determine whether the subunit is an α_1 , α_2 subunit or β subunit.

For example, DNA encoding an α_{1A} subunit may be isolated by screening an appropriate library with DNA, encoding all or a portion of the human α_{1A} subunit. Such DNA includes the DNA in the phage deposited under ATCC Accession No. 75293 that encodes a portion of an α_1 subunit. DNA encoding an α_{1A} subunit may obtained from an appropriate library by screening with an oligonucleotide having all or a portion of the sequence set forth in SEQ ID No. 21, 22 and/or 23 or with the DNA in the deposited phage. Alternatively, such DNA may have a sequence that encodes an α_{1A} subunit that is encoded by SEQ ID No. 22 or 23.

Similarly, DNA encoding β_3 may be isolated by screening a human cDNA library with DNA probes prepared from the plasmid $\beta1.42$ deposited under ATCC Accession No. 69048 or obtained from an appropriate library using probes having sequences prepared according to the sequences set forth in SEQ ID Nos. 19 and/or 20. Also, DNA encoding β_4 may be isolated by screening a human cDNA library with DNA probes prepared according to DNA set forth in SEQ ID No. 27, which sets forth the DNA sequence of a clone encoding a β_4 subunit. The amino acid sequence is set forth in SEQ ID No. 28. Any method known to those of skill in the art for isolation and identification of DNA and preparation of full-length genomic or cDNA clones, including methods exemplified herein, may be used. DNA encoding

The subunit encoded by isolated DNA may be identified by comparison with the DNA and amino acid sequences of the

subunits provided herein. Splice variants share extensive regions of homology, but include non-homologous regions, subunits encoded by different genes share a uniform distribution of non-homologous sequences.

As used herein, a splice variant refers to a variant produced by differential processing of a primary transcript of genomic DNA that results in more than one type of mRNA. Splice variants may occur within a single tissue type or among tissues (tissue-specific variants). Thus, cDNA clones that encode calcium channel subunit subtypes that have regions of identical amino acids and regions of different amino acid sequences are referred to herein as "splice variants".

As used herein, a "calcium channel-selective ion" is an ion that is capable of flowing through, or being blocked from flowing through, a calcium channel which spans a cellular membrane under conditions which would substantially similarly permit or block the flow of Ca²⁺. Ba²⁺ is an example of an ion which is a calcium channel-selective ion.

As used herein, a compound that modulates calcium channel activity is one that affects the ability of the calcium channel to pass calcium channel-selective ions or affects other detectable calcium channel features, such as current kinetics. Such compounds include calcium channel antagonists and agonists and compounds that exert their effect on the activity of the calcium channel directly or indirectly.

As used herein, a "substantially pure" subunit or protein is a subunit or protein that is sufficiently free of other polypeptide contaminants to appear homogeneous by SDS-PAGE or to be unambiguously sequenced.

As used herein, selectively hybridize means that a DNA fragment hybridizes to a second fragment with sufficient specificity to permit the second fragment to be identified or isolated from among a plurality of fragments. In general, selective hybridization occurs at conditions of high stringency.

As used herein, heterologous or foreign DNA and RNA are used interchangeably and refer to DNA or RNA that does not occur naturally as part of the genome in which it is present or which is found in a location or locations in the genome that differ from that in which it occurs in nature. It is DNA or RNA that is not endogenous to the cell and has been artificially introduced into the cell. Examples heterologous DNA include, but are not limited to, DNA that encodes a calcium channel subunit and DNA that encodes RNA or proteins that mediate or alter expression of endogenous DNA by affecting transcription, translation, or other regulatable The cell that expresses processes. biochemical heterologous DNA, such as DNA encoding a calcium channel subunit, may contain DNA encoding the same or different calcium channel subunits. The heterologous DNA need not be expressed and may be introduced in a manner such that it is integrated into the host cell genome or is maintained episomally.

As used herein, operative linkage of heterologous DNA to regulatory and effector sequences of nucleotides, such as promoters, enhancers, transcriptional and translational stop sites, and other signal sequences, refers to the functional relationship between such DNA and such sequences of nucleotides. For example, operative linkage of heterologous DNA to a promoter refers to the physical and functional relationship between the DNA and the promoter such that the transcription of such DNA is initiated from the promoter by an RNA polymerase that specifically recognizes, binds to and transcribes the DNA in reading frame.

As used herein, isolated, substantially pure DNA refers to DNA fragments purified according to standard techniques employed by those skilled in the art [see, e.g., Maniatis et al. (1982) Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY].

As used herein, expression refers to the process by which nucleic acid is transcribed into mRNA and translated into

peptides, polypeptides, or proteins. If the nucleic acid is derived from genomic DNA, expression may, if an appropriate eukaryotic host cell or organism is selected, include splicing of the mRNA.

As used herein, vector or plasmid refers to discrete elements that are used to introduce heterologous DNA into cells for either expression of the heterologous DNA or for replication of the cloned heterologous DNA. Selection and use of such vectors and plasmids are well within the level of skill of the art.

As used herein, expression vector includes vectors capable of expressing DNA fragments that are in operative linkage with regulatory sequences, such as promoter regions, that are capable of effecting expression of such DNA fragments. Thus, an expression vector refers to a recombinant DNA or RNA construct, such as a plasmid, a phage, recombinant virus or other vector that, upon introduction into an appropriate host cell, results in expression of the cloned DNA. Appropriate expression vectors are well known to those of skill in the art and include those that are replicable in eukaryotic cells and/or prokaryotic cells and those that remain episomal or may integrate into the host cell genome.

As used herein, a promoter region refers to the portion of DNA of a gene that controls transcription of DNA to which it is operatively linked. The promoter region includes specific sequences of DNA that are sufficient for polymerase recognition, binding and transcription initiation. This portion of the promoter region is referred to as the promoter. In addition, the promoter region includes sequences that modulate this recognition, binding and transcription initiation activity of the RNA polymerase. These sequences may be cis acting or may be responsive to trans acting factors. Promoters, depending upon the nature of the regulation, may be constitutive or regulated.

As used herein, a recombinant eukaryotic cell is a eukaryotic cell that contains heterologous DNA or RNA.

As used herein, a recombinant or heterologous calcium channel refers to a calcium channel that contains one or more subunits that are encoded by heterologous DNA that has been introduced into and expressed in a eukaryotic cells that expresses the recombinant calcium channel. A recombinant calcium channel may also include subunits that are produced by DNA endogenous to the cell. In certain embodiments, the recombinant or heterologous calcium channel may contain only subunits that are encoded by heterologous DNA.

As used herein, "functional" with respect to a recombinant or heterologous calcium channel means that the channel is able to provide for and regulate entry of calcium channel-selective ions, including, but not limited to, Ca²⁺ or Ba²⁺, in response to a stimulus and/or bind ligands with affinity for the channel. Preferably such calcium channel activity is distinguishable, such as electrophysiological, pharmacological and other means known to those of skill in the art, from any endogenous calcium channel activity that in the host cell.

As used herein, a peptide having an amino acid sequence substantially as set forth in a particular SEQ ID No. includes peptides that have the same function but may include minor variations in sequence, such as conservative amino acid changes or minor deletions or insertions that do not alter the activity of the peptide. The activity of a calcium channel receptor subunit peptide refers to its ability to form functional calcium channels with other such subunits.

As used herein, a physiological concentration of a compound is that which is necessary and sufficient for a biological process to occur. For example, a physiological concentration of a calcium channel-selective ion is a concentration of the calcium channel-selective ion necessary and sufficient to provide an inward current when the channels open.

As used herein, activity of a calcium channel refers to the movement of a calcium channel-selective ion through a

calcium channel. Such activity may be measured by any method known to those of skill in the art, including, but not limited to, measurement of the amount of current which flows through the recombinant channel in response to a stimulus.

As used herein, a "functional assay" refers to an assay that identifies functional calcium channels. A functional assay, thus, is an assay to assess function.

As understood by those skilled in the art, assay methods for identifying compounds, such as antagonists and agonists, that modulate calcium channel activity, generally requires comparison to a control. One type of a "control" cell or "control" culture is a cell or culture that is treated substantially the same as the cell or culture exposed to the test compound except that the control culture is not exposed to the test compound. Another type of a "control" cell or "control" culture may be a cell or a culture of cells which are identical to the transfected cells except the cells employed for the control culture do not express functional calcium channels. In this situation, the response of test cell to the test compound is compared to the response (or lack of response) of the calcium channel-negative cell to the test compound, when cells or cultures of each type of cell are exposed to substantially the same reaction conditions in the presence of the compound being assayed. For example, methods that use patch clamp electrophysiological procedures, the same cell can be tested in the presence and absence of the test compound, by changing the external solution bathing the cell as known in the art.

It is also understood that each of the subunits disclosed herein may be modified by making conservative amino acid substitutions and the resulting modified subunits are contemplated herein. Suitable conservative substitutions of amino acids are known to those of skill in this art and may be made generally without altering the biological activity of the resulting molecule. Those of skill in this art recognize that, in general, single amino acid substitutions in non-

essential regions of a polypeptide do not substantially alter biological activity (see, <u>e.g.</u>, Watson <u>et al.</u> Molecular Biology of the Gene, 4th Edition, 1987, The Bejacmin/Cummings Pub. co., p.224). Such substitutions are preferably, although not exclusively, made in accordance with those set forth in TABLE 1 as follows:

	MABLE 1
Original residue Ala (A)	Conservative substitution Gly; Ser
Arg (R)	Lys
Asn (N)	Gln; His
Cys (C)	Ser
Gln (Q)	Asn
Glu (E)	Asp
Gly (G)	Ala; Pro
His (H)	Asn; Gin
lle (1)	Leu; Val
Leu (L)	ile; Val
Lys (K)	Arg; Gin; Glu
Met (M)	Leu; Tyr; lle
Phe (F)	Met; Leu; Tyr
Ser (S)	Thr
Thr (T)	Ser
Trp (W)	Tyr
Tyr (Y)	Trp; Phe
Val (V)	lle; Leu

Other substitutions are also permissible and may be determined empirically or in accord with known conservative substitutions. Any such modification of the polypeptide may be effected by any means known to those of skill in this art. Mutation may be effected by any method known to those of skill in the art, including site-specific or site-directed mutagenesis of DNA encoding the protein and the use of DNA amplification methods using primers to introduce and amplify alterations in the DNA template.

Identification and isolation of DNA encoding human calcium channel subunits

Methods for identifying and isolating DNA encoding α_1 , α_2 , β and γ subunits of human calcium channels are provided.

Identification and isolation of such DNA may be accomplished by hybridizing, under appropriate conditions, at least low stringency whereby DNA that encodes the desired

subunit is isolated, restriction enzyme-digested human DNA with a labeled probe having at least 14, preferably 16 or more nucleotides and derived from any contiguous portion of DNA having a sequence of nucleotides set forth herein by sequence identification number. Once a hybridizing fragment identified in the hybridization reaction, it can be cloned employing standard cloning techniques known to those of skill Full-length clones may be identified by the in the art. presence of a complete open reading frame and the identity of the encoded protein verified by sequence comparison with the subunits provided herein and by functional assays to assess calcium channel- forming ability or other function. method can be used to identify genomic DNA encoding the subunit or cDNA encoding splice variants of human calcium channel subunits generated by alternative splicing of the primary transcript of genomic subunit DNA. For instance, DNA, cDNA or genomic DNA, encoding a calcium channel subunit may be identified by hybridization to a DNA probe and characterized by methods known to those of skill in the art, such as restriction mapping and DNA sequencing, and compared to the DNA provided herein in order to identify heterogeneity or divergence in the sequences of the DNA. Such sequence differences may indicate that the transcripts from which the cDNA was produced result from alternative splicing of a primary transcript, if the non-homologous and homologous regions are clustered, or from a different gene if the nonhomologous regions are distributed throughout the cloned DNA.

Any suitable method for isolating genes using the DNA provided herein may be used. For example, oligonucleotides corresponding to regions of sequence differences have been used to isolate, by hybridization, DNA encoding the full-length splice variant and can be used to isolate genomic clones. A probe, based on a nucleotide sequence disclosed herein, which encodes at least a portion of a subunit of a human calcium channel, such as a tissue-specific exon, may be used as a probe to clone related DNA, to clone a full-length

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cDNA clone or genomic clone encoding the human calcium channel subunit.

Labeled, including, but not limited to, radioactively or enzymatically labeled, RNA or single-stranded DNA of at least 14 substantially contiguous bases, preferably 16 or more, generally at least 30 contiguous bases of a nucleic acid which encodes at least a portion of a human calcium channel subunit, the sequence of which nucleic acid corresponds to a segment of a nucleic acid sequence disclosed herein by reference to a SEQ ID No. are provided. Such nucleic acid segments may be used as probes in the methods provided herein for cloning DNA encoding calcium channel subunits. See, generally, Sambrook et al. (1989) Molecular Cloning: A Laboratory Manual, 2nd Edition, Cold Spring Harbor Laboratory Press.

In addition, nucleic acid amplification techniques, which are well known in the art, can be used to locate splice of calcium channel subunits by employing variants oligonucleotides based on DNA sequences surrounding the divergent sequence primers for amplifying human RNA or genomic Size and sequence determinations of the amplification products can reveal splice variants. Furthermore, isolation of human genomic DNA sequences by hybridization can yield DNA containing multiple exons, separated by introns, correspond to different splice variants of transcripts encoding human calcium channel subunits.

DNA encoding types and subtypes of each of the α_1 , α_2 , β and γ subunit of voltage-dependent human calcium channels has been cloned herein by nucleic acid amplication of cDNA from selected tissues or by screening human cDNA libraries prepared from isolated poly A+ mRNA from cell lines or tissue of human origin having such calcium channels. Among the sources of such cells or tissue for obtaining mRNA are human brain tissue or a human cell line of neural origin, such as a neuroblastoma cell line, human skeletal muscle or smooth muscle cells, and the like. Methods of preparing cDNA libraries are well known in the art [see generally Ausubel et al. (1987) Current

Protocols in Molecular Biology, Wiley-Interscience, New York; and Davis et al. (1986) Basic Methods in Molecular Biology, Elsevier Science Publishing Co., New York].

Preferred regions from which to construct probes include 5' and/or 3' coding sequences, sequences predicted to encode transmembrane domains, sequences predicted cytoplasmic loops, signal sequences, ligand-binding sites, and other functionally significant sequences (see Table, below). Either the full-length subunit-encoding DNA or fragments thereof can be used as probes, preferably labeled with suitable label means for ready detection. When fragments are used as probes, preferably the DNA sequences will be typically from the carboxyl-end-encoding portion of the DNA, and most preferably will include predicted transmembrane domainencoding portions based on hydropathy analysis of the deduced amino acid sequence [see, e.g., Kyte and Doolittle [(1982) J. Mol. Biol. 167:105].

Riboprobes that specific for human calcium channel subunit types or subtypes have been prepared. These probes are useful for identifying expression of particular subunits in selected tissues and cells. The regions from which the probes were prepared were identified by comparing the DNA and amino acid sequences of all known α or β subunit subtypes. Regions of least homology, preferably human-derived sequences, and generally about 250 to about 600 nucleotides were selected. Numerous riboprobes for α and β subunits have been prepared; some of these are listed in the following Table.

TABLE 2 SUMMARY OF RNA PROBES

SUBUNIT SPECIFICITY	NUCLEOTIDE POSITION	PROBE NAME	PROBE TYPE	ORIENTA- TION
αlA generic	3357-3840	pGEM7Zα1A*	riboprobe	n/a
	761-790	SE700	oligo	antisense
	3440-3464	SE718	oligo	antisense
	3542-3565	SE724	oligo	sense
α1B generic	3091-3463	pGEM7Zα1B _{eyt}	riboprobe	n/a
	6635-6858	pGEM7ZalBcooh	riboprobe	n/a
αlB-l specific	6490-6676	pCRII α1B-1/187	riboprobe	n/a
αlE generic	3114-3462	pGEM7Zα1E	riboprobe	n/a
α2b	1321-1603	pCRII@2b	riboprobe	n/a
β generic(?)	212-236	SE300	oligo_	antisense
β1 generic	1267-1291	SE301	oligo	antisense
β1-2 specific	1333-1362	SE17	oligo	antisense
	1682-1706	SE23	oligo	sense
	2742-2766	SE43	oligo_	antisense
	27-56	SE208	oligo	antisense
	340-364	SE274	oligo	antisense
	340-364	SE275	oligo	sense
β3 specific	1309-1509		riboprobe	n/a
β4 specific	1228-1560		riboprobe	n/a

* The pGEM series are available from Promega, Madison WI; see also, U.S. Patent No. 4,766,072.

The above-noted nucleotide regions are also useful in selecting regions of the protein for preparation of subunit-specific antibodies, discussed below.

The DNA clones and fragments thereof provided herein thus can be used to isolate genomic clones encoding each subunit and to isolate any splice variants by hybridization screening of libraries prepared from different human tissues. Nucleic acid amplification techniques, which are well known in the art, can also be used to locate DNA encoding splice variants

of human calcium channel subunits. This is accomplished by employing oligonucleotides based on DNA sequences surrounding divergent sequence(s) as primers for amplifying human RNA or genomic DNA. Size and sequence determinations of the amplification products can reveal the existence of splice variants. Furthermore, isolation of human genomic DNA sequences by hybridization can yield DNA containing multiple exons, separated by introns, that correspond to different splice variants of transcripts encoding human calcium channel subunits.

Once DNA encoding a calcium channel subunit is isolated, ribonuclease (RNase) protection assays can be employed to determine which tissues express mRNA encoding a particular calcium channel subunit or variant. These assays provide a sensitive means for detecting and quantitating an RNA species in a complex mixture of total cellular RNA. The subunit DNA is labeled and hybridized with cellular RNA. If complementary mRNA is present in the cellular RNA, a DNA-RNA hybrid results. The RNA sample is then treated with RNase, which degrades single-stranded RNA. Any RNA-DNA hybrids are protected from RNase degradation and can be visualized by gel electrophoresis and autoradiography. In situ hybridization techniques can also be used to determine which tissues express mRNA encoding a particular calcium channel subunit. The labeled subunit DNAs are hybridized to different tissue slices to visualize subunit mRNA expression.

With respect to each of the respective subunits (α_1 , α_2 , β or γ) of human calcium channels, once the DNA encoding the channel subunit was identified by a nucleic acid screening method, the isolated clone was used for further screening to identify overlapping clones. Some of the cloned DNA fragments can and have been subcloned into an appropriate vector such as pIBI24/25 (IBI, New Haven, CT), M13mp18/19, pGEM4, pGEM3, pGEM7Z, pSP72 and other such vectors known to those of skill in this art, and characterized by DNA sequencing and restriction enzyme mapping. A sequential series of

overlapping clones may thus be generated for each of the subunits until a full-length clone can be prepared by methods, known to those of skill in the art, that identification of translation initiation (start) translation termination (stop) codons. For expression of the cloned DNA, the 5' noncoding region and other transcriptional and translational control regions of such a clone may be replaced with an efficient ribosome binding site and other regulatory regions as known in the art. Other modifications of the 5' end, known to those of skill in the art, that may be optimize translation and/or transcription required to efficiency may also be effected, if deemed necessary.

Examples II-VIIII, below, describe in detail the cloning of each of the various subunits of a human calcium channel as well as subtypes and splice variants, including tissue-specific variants thereof. In the few instances in which partial sequences of a subunit are disclosed, it is well within the skill of the art, in view of the teaching herein, to obtain the corresponding full-length clones and sequence thereof encoding the subunit, subtype or splice variant thereof using the methods described above and exemplified below.

Identification and isolation of DNA encoding $lpha_1$ subunits

A number of voltage-dependent calcium channel α_1 subunit genes, which are expressed in the human CNS and in other tissues, have been identified and have been designated as α_{1A} , α_{1B} (or VDCC IV), α_{1C} (or VDCC II), α_{1D} (or VDCC III) and α_{1E} . DNA, isolated from a human neural cDNA library, that encodes each of the subunit types has been isolated. DNA encoding subtypes of each of the types, which arise as splice variants are also provided. Subtypes are herein designated, for example, as α_{1B-1} , α_{1B-2} .

The α_1 subunits types A B, C, D and E of voltage-dependent calcium channels, and subtypes thereof, differ with respect to sensitivity to known classes of calcium channel

agonists and antagonists, such as DHPs, phenylalkylamines, omega conotoxin (ω -CgTx), the funnel web spider toxin ω -Aga-IV, and pyrazonoylguanidines. These subunit types also appear to differ in the holding potential and in the kinetics of currents produced upon depolarization of cell membranes containing calcium channels that include different types of α_1 subunits.

DNA that encodes an α_1 subunit that binds to at least one compound selected from among dihydropyridines, phenylalkylamines, ω -CgTx, components of funnel web spider toxin, and pyrazonoylguanidines is provided. For example, the α_{1B} subunit provided herein appears to specifically interact with ω -CgTx in N-type channels, and the α_{1D} subunit provided herein specifically interacts with DHPs in L-type channels.

Identification and isolation of DNA encoding the $\alpha_{\rm 1D}$ human calcium channel subunit

The $\alpha_{\rm 1D}$ subunit cDNA has been isolated using fragments of the rabbit skeletal muscle calcium channel $\alpha_{\rm 1}$ subunit cDNA as a probe to screen a cDNA library of a human neuroblastoma cell line, IMR32, to obtain clone $\alpha 1.36$. This clone was used as a probe to screen additional IMR32 cell cDNA libraries to obtain overlapping clones, which were then employed for screening until a sufficient series of clones to span the length of the nucleotide sequence encoding the human $\alpha_{\rm 1D}$ subunit was obtained. Full-length clones encoding $\alpha_{\rm 1D}$ were constructed by ligating portions of partial $\alpha_{\rm 1D}$ clones as described in Example II. SEQ ID No. 1 shows the 7,635 nucleotide sequence of the cDNA encoding the $\alpha_{\rm 1D}$ subunit. There is a 6,483 nucleotide sequence reading frame which encodes a sequence of 2,161 amino acids (as set forth in SEQ ID No. 1).

SEQ ID No. 2 provides the sequence of an alternative exon encoding the IS6 transmembrane domain [see Tanabe, T., et al. (1987) Nature 328:313-318 for a description of transmembrane domain terminology] of the α_{1D} subunit.

SEQ ID No. 1 also shows the 2,161 amino acid sequence deduced from the human neuronal calcium channel α_{1D} subunit

DNA. Based on the amino acid sequence, the α_{1D} protein has a calculated Mr of 245,163. The α_{1D} subunit of the calcium channel contains four putative internal repeated sequence regions. Four internally repeated regions represent 24 putative transmembrane segments, and the amino- and carboxyl-termini extend intracellularly.

The α_{1D} subunit has been shown to mediate DHP-sensitive, high-voltage-activated, long-lasting calcium channel activity. This calcium channel activity was detected when oöcytes were co-injected with RNA transcripts encoding an α_{1D} and β_{1-2} or α_{1D} , α_{2b} and β_{1-2} subunits. This activity was distinguished from Ba²⁺ currents detected when oöcytes were injected with RNA transcripts encoding the β_{1-2} ± α_{2b} subunits. These currents pharmacologically and biophysically resembled Ca²⁺ currents reported for uninjected oöcytes.

Identification and isolation of DNA encoding the $\alpha_{1\lambda}$ human calcium channel subunit

Biological material containing DNA encoding a portion of the α_{1A} subunit had been deposited in the American Type Culture Collection, 12301 Parklawn Drive, Rockville, Maryland 20852 U.S.A. under the terms of the Budapest Treaty on the International Recognition of Deposits of Microorganisms for Purposes of Patent Procedure and the Regulations promulgated under this Treaty. Samples of the deposited material are and will be available to industrial property offices and other persons legally entitled to receive them under the terms of the Treaty and Regulations and otherwise in compliance with the patent laws and regulations of the United States of America and all other nations or international organizations in which this application, or an application claiming priority of this application, is filed or in which any patent granted on any such application is granted.

A portion of an α_{1A} subunit is encoded by an approximately 3 kb insert in λ gt10 phage designated α 1.254 in E. coli host strain NM514. A phage lysate of this material has been deposited as at the American Type Culture Collection under

ATCC Accession No. 75293, as described above. DNA encoding α_{1A} may also be identified by screening with a probe prepared from DNA that has SEQ ID No. 21:

5' CTCAGTACCATCTCTGATACCAGCCCCA 3'.

 α_{1A} splice variants have been obtained. The sequences of two α_{1A} splice variants, α_{1a-1} and α_{1a-2} are set forth in SEQ. ID Nos. 22 and 23. Other splice variants may be obtained by screening a human library as described above or using all or a portion of the sequences set forth in SEQ ID Nos. 22 and 23.

Identification and isolation of DNA encoding the $\alpha_{\rm 1B}$ human calcium channel subunit

DNA encoding the α_{1B} subunit was isolated by screening a human basal ganglia cDNA library with fragments of the rabbit skeletal muscle calcium channel α_1 subunit-encoding cDNA. A portion of one of the positive clones was used to screen an IMR32 cell cDNA library. Clones that hybridized to the basal ganglia DNA probe were used to further screen an IMR32 cell cDNA library to identify overlapping clones that in turn were used to screen a human hippocampus cDNA library. In this way, a sufficient series of clones to span nearly the entire length of the nucleotide sequence encoding the human α_{1B} subunit was obtained. Nucleic acid amplification of specific regions of the IMR32 cell α_{1B} mRNA yielded additional segments of the α_{1B} coding sequence.

A full-length α_{1B} DNA clone was constructed by ligating portions of the partial cDNA clones as described in Example II.C. SEQ ID Nos. 7 and 8 show the nucleotide sequences of DNA clones encoding the α_{1B} subunit as well as the deduced amino acid sequences. The α_{1B} subunit encoded by SEQ ID No. 7 is referred to as the α_{1B-1} subunit to distinguish it from another α_{1B} subunit, α_{1B-2} , encoded by the nucleotide sequence shown as SEQ ID No. 8, which is derived from alternative splicing of the α_{1B} subunit transcript.

Nucleic acid amplification of IMR32 cell mRNA using oligonucleotide primers designed according to nucleotide

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sequences within the α_{1B-1} -encoding DNA has identified variants of the α_{1B} transcript that appear to be splice variants because they contain divergent coding sequences.

Identification and isolation of DNA encoding the $\alpha_{\rm lc}$ human calcium channel subunit

clones α_{1c} -specific DNA were isolated. Numerous Characterization of the sequence revealed the α_{1c} coding sequence, the α_{1c} initiation of translation sequence, and an alternatively spliced region of α_{1c} . Alternatively spliced variants of the α_{10} subunit have been identified. 3 sets forth DNA encoding a substantial protion of an α_{1c} subunit. The DNA sequences set forth in SEQ ID No. 4 and No. 5 encode two possible amino terminal ends of the α_{1c} protein. SEQ ID No. 6 encodes an alternative exon for the IV S3 transmembrane domain. The sequences of portions of two α_{1c} splice variants, designated α_{1c-1} and α_{1c-2} , are set forth in SEQ ID NOs. 3 and 36, respectively.

The isolation and identification of DNA clones encoding portions of the $\alpha_{\rm ic}$ subunit is described in detail in Example II.

Identification and isolation of DNA encoding the α_{12} human calcium channel subunit

DNA encoding α_{1E} human calcium channel subunits have been isolated from an oligo dT-primed human hippocampus library. The resulting clones, which are splice variants, were designated α_{1E-1} and α_{1E-3} . The subunit designated α_{1E-1} has the amino acid sequence set forth in SEQ ID No. 24, and a subunit designated α_{1E-3} has the amino acid sequence set forth in SEQ ID No. 25. These splice variants differ by virtue of a 57 base pair insert between nucleotides 2405 and 2406 of SEQ. ID No. 24.

The α_{1E} subunits provided herein appear to participate in the formation of calcium channels that have properties of high-voltage activated calcium channels and low-voltage activated channels. These channels are rapidly inactivating

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compared to other high voltage-activated calcium channels. In addition these channels exhibit pharmacological profiles that are similar to voltage-activated channels, but are also sensitive to DHPs and ω -Aga-IVA, which block certain high voltage activated channels. Additional details regarding the electrophysiology and pharmacology of channels containing α_{IE} subunits is provided in Example VII. F.

Identification and isolation of DNA encoding encoding additional α_1 human calcium channel subunit types and subtypes

DNA encoding additional α_1 subunits can be isolated and identified using the DNA provided herein as described for the α_{1A} , α_{1B} , α_{1C} , α_{1D} and α_{1E} subunits or using other methods known to those of skill in the art. In particular, the DNA provided herein may be used to screen appropriate libraries to isolate related DNA. Full-length clones can be constructed using methods, such as those described herein, and the resulting subunits characterized by comparison of their sequences and electrophysiological and pharmacological properties with the subunits exemplified herein.

Identification and isolation of DNA encoding β human calcium channel subunits DNA encoding β_1

To isolate DNA encoding the β_1 subunit, a human hippocampus cDNA library was screened by hybridization to a DNA fragment encoding a rabbit skeletal muscle calcium channel β subunit. A hybridizing clone was selected and was in turn used to isolate overlapping clones until the overlapping clones encompassing DNA encoding the entire the human calcium channel β subunit were isolated and sequenced.

Five alternatively spliced forms of the human calcium channel β_1 subunit have been identified and DNA encoding a number of forms have been isolated. These forms are designated β_{1-1} , expressed in skeletal muscle, β_{1-2} , expressed in the CNS, β_{1-3} , also expressed in the in the CNS, β_{1-4} , expressed in aorta tissue and HEK 293 cells, and β_{1-5} ,

expressed in HEK 293 cells. Full-length DNA clones encoding the β_{1-2} and β_{1-3} subunits have been constructed. The subunits β_{1-1} , β_{1-2} , β_{1-4} and β_{1-5} have been identified by nucleic acid amplification analysis as alternatively spliced forms of the β subunit. Sequences of the β_1 splice variants are set forth in SEQ ID Nos. 9, 10 and 33-35.

DNA encoding β_2

DNA encoding the β_2 splice variants has been obtained. These splice variants include eta_{2c} - eta_{2e} . Splice variants eta_{2c} - eta_{2e} include all of sequence set forth in SEQ ID No. 26, except for the portion at the 5' end (up to nucleotide 182), which differs among splice variants. The sequence set forth in SEQ Additional splice variants may be ID No. 26 encodes β_{2D} . herein described the methods using oligonucleotides including all or portions of the DNA set forth in SEQ ID. No. 26 or may be prepared or obtained as described in the Examples. The sequences of variants β_{2C} and β_{2E} are set forth in SEQ ID Nos. 37 and 38, respectively.

DNA encoding β_3

DNA encoding the β_3 subunit and any splice variants thereof may be isolated by screening a library, as described above for the β_1 subunit, using DNA probes prepared according to SEQ ID Nos. 19, 20 or using all or a portion of the deposited β_3 clone plasmid $\beta1.42$ (ATCC Accession No. 69048).

The $E.\ coli$ host containing plasmid $\beta1.42$ that includes DNA encoding a β_3 subunit has been deposited as ATCC Accession No. 69048 in the American Type Culture Collection, 12301 Parklawn Drive, Rockville, Maryland 20852 U.S.A. under the terms of the Budapest Treaty on the International Recognition of Deposits of Microorganisms for Purposes of Patent Procedure and the Regulations promulgated under this Treaty. Samples of the deposited material are and will be available to industrial property offices and other persons legally entitled to receive them under the terms of the Treaty and Regulations and otherwise in compliance with the patent laws and regulations

of the United States of America and all other nations or international organizations in which this application, or an application claiming priority of this application, is filed or in which any patent granted on any such application is granted.

The β_3 encoding plasmid is designated $\beta_1.42$. The plasmid contains a 2.5 kb EcoRI fragment encoding β_3 inserted into vector pGem $^{\circ}7zF(+)$ and has been deposited in $E.\ coli$ host strain DH5 α . The sequences of β_3 splice variants, designated β_{3-1} and β_{3-2} are set forth in SEQ ID Nos. 19 and 20, respectively.

Identification and isolation of DNA encoding the $\alpha 2$ human calcium channel subunit

DNA encoding a human neuronal calcium channel α_2 subunit was isolated in a manner substantially similar to that used for isolating DNA encoding an α_1 subunit, except that a human genomic DNA library was probed under low and high stringency conditions with a fragment of DNA encoding the rabbit skeletal muscle calcium channel α_2 subunit. The fragment included nucleotides having a sequence corresponding to the nucleotide sequence between nucleotides 43 and 272 inclusive of rabbit back skeletal muscle calcium channel α_2 subunit cDNA as disclosed in PCT International Patent Application Publication No. WO 89/09834, which corresponds to U.S. Application Serial No. 07/620,520 (now allowed U.S. Application Serial No. 07/914,231), which is a continuation-in-part of United States Serial No. 176,899, filed April 4, 1988.

Example IV describes the isolation of DNA clones encoding α_2 subunits of a human calcium channel from a human DNA library using genomic DNA and cDNA clones, identified by hybridization to the genomic DNA, as probes.

SEQ ID Nos. 11 and 29-32 show the sequence of DNA encoding α_2 subunits. As described in Example V, nucleic acid amplification analysis of RNA from human skeletal muscle, brain tissue and aorta using oligonucleotide primers specific for a region of the human neuronal α_2 subunit cDNA that

diverges from the rabbit skeletal muscle calcium channel α_2 subunit cDNA identified splice variants of the human calcium channel α_2 subunit transcript.

Identification and isolation of DNA encoding $\boldsymbol{\gamma}$ human calcium channel subunits

DNA encoding a portion of a human neuronal calcium channel γ subunit has been isolated as described in detail in Example VI. SEQ ID No. 14 shows the nucleotide sequence at the 3'-end of this DNA which includes a reading frame encoding a sequence of 43 amino acid residues. Since the portion that has been obtained is homologous to the rabbit clone, described in allowed co-owned U.S. Application Serial No. 07/482,384, the remainder of the clone can be obtained using routine methods.

Antibodies

Antibodies, monoclonal or polyclonal, specific for calcium channel subunit subtypes or for calcium channel types can be prepared employing standard techniques, known to those of skill in the art, using the subunit proteins or portions thereof as antigens. Anti-peptide and anti-fusion protein antibodies can be used [see, for example, Bahouth et al. (1991) Trends Pharmacol. Sci. 12:338-343; Current Protocols in Molecular Biology (Ausubel et al., eds.) John Wiley and Sons, Factors to consider in selecting portions New York (1984)]. of the calcium channel subunits for use as immunogens (as either a synthetic peptide or a recombinantly produced bacterial fusion protein) include antigenicity accessibility (i.e., extracellular and cytoplasmic domains), uniqueness to the particular subunit, and other factors known to those of skill in this art.

The availability of subunit-specific antibodies makes application the technique of the possible the distribution and immunohistochemistry monitor to expression density of various subunits (e.g., in normal vs Such antibodies could also be diseased brain tissue). employed in diagnostic, such as LES diagnosis, and therapeutic applications, such as using antibodies that modulate activities of calcium channels.

The antibodies can be administered to a subject employing standard methods, such as, for example, by intraperitoneal, intramuscular, intravenous, or subcutaneous injection, implant or transdermal modes of administration, and the like. One of skill in the art can empirically determine dose forms, treatment regiments, etc., depending on the mode of administration employed.

Subunit-specific monoclonal antibodies and polyclonal antisera have been prepared. The regions from which the antigens were identified by comparing the DNA and amino acid sequences of all known α or β subunit subtypes. Regions of least homology, preferably human-derived sequences selected. The selected regions or fusion proteins containing the selected regions are used as immunogens. Hydrophobicity analyses of residues in selected protein regions and fusion proteins are also performed; regions of high hydrophobicity Also, and more importantly, when preparing are avoided. fusion proteins in bacterial hosts, rare codons are avoided. In particular, inclusion of 3 or more successive rare codons a selected host is avoided. Numerous antibodies, polyclonal and monoclonal, specific for α or β subunits types or subtypes have been prepared; some of these are listed in the following Table. Exemplary antibodies and peptide antigens used to prepare the antibodies are set forth in the following Table:

TABLE 3

SPECIFICITY	AMINO ACID NUMBER	ANTIGEN NAME	ANTIBODY TYPE
αl generic	112-140	peptide 1A#1	polyclonal
αl generic	1420-1447	peptide 1A#2	polyclonal
αlA generic	1048-1208	αlA#2(b)GST fusion	polyclonal
		· ·	monoclonal
αlB generic	983-1106	α1B#2(b) GST fusion	polyclonal
			monoclonal

α1B-1	2164-2339	αlB-1#3 GST fusion	polyclonal
α1B-2	2164-2237	α1B-2#4 GST fusion	polyclonal
αlE generic	985-1004 (α1E-3)	α1E#2(a) GST fusion	polyclonal

* GST gene fusion system is available from Pharmacia; see also, Smith et al. (1988) Gene 67:31. The system provides pGEX plasmids that are designed for inducible, high-level expression of genes or gene fragments as fusions with Schistosoma japonicum GST. Upon expression in a bacterial host, the resulting fusion proteins are purified from bacterial lysates by affinity chromatography.

The GST fusion proteins are each specific for the cytoplasmic loop region IIS6-IIS1, which is a region of low subtype homology for all subtypes, including α_{1c} and α_{1D} , for which similar fusions and antisera can be prepared.

Preparation of recombinant eukaryotic cells containing DNA encoding heterologous calcium channel subunits

DNA encoding one or more of the calcium channel subunits or a portion of a calcium channel subunit may be introduced into a host cell for expression or replication of the DNA. Such DNA may be introduced using methods described in the following examples or using other procedures well known to those skilled in the art. Incorporation of cloned DNA into a suitable expression vector, transfection of eukaryotic cells with a plasmid vector or a combination of plasmid vectors, each encoding one or more distinct genes or with linear DNA, and selection of transfected cells are also well known in the art [see, e.g., Sambrook et al. (1989) Molecular Cloning: A Cold Spring Edition, Laboratory Manual, Second Cloned full-length DNA encoding any of Laboratory Press]. the subunits of a human calcium channel may be introduced into a plasmid vector for expression in a eukaryotic cell. DNA may be genomic DNA or cDNA. Host cells may be transfected with one or a combination of the plasmids, each of which encodes at least one calcium channel subunit. Alternatively, host cells may be transfected with linear DNA using methods well known to those of skill in the art.

While the DNA provided herein may be expressed in any eukaryotic cell, including yeast cells such as P. pastoris

[see, e.g., Cregg et al. (1987) Bio/Technology 5:479], mammalian expression systems for expression of the DNA encoding the human calcium channel subunits provided herein are preferred.

The heterologous DNA may be introduced by any method known to those of skill in the art, such as transfection with a vector encoding the heterologous DNA. Particularly preferred vectors for transfection of mammalian cells are the pSV2dhfr expression vectors, which contain the SV40 early promoter, mouse dhfr gene, SV40 polyadenylation and splice sites and sequences necessary for maintaining the vector in bacteria, cytomegalovirus (CMV) promoter-based vectors such as pCDNA1, or pcDNA-amp and MMTV promoter-based vectors. DNA encoding the human calcium channel subunits has been inserted in the vector pCDNA1 at a position immediately following the CMV promoter. The vector pCDNA1 is presently preferred.

Stably or transiently transfected mammalian cells may be prepared by methods known in the art by transfecting cells with an expression vector having a selectable marker gene such as the gene for thymidine kinase, dihydrofolate reductase, neomycin resistance or the like, and, for transient transfection, growing the transfected cells under conditions selective for cells expressing the marker gene. Functional voltage-dependent calcium channels have been produced in HEK 293 cells transfected with a derivative of the vector pCDNA1 that contains DNA encoding a human calcium channel subunit.

The heterologous DNA may be maintained in the cell as an episomal element or may be integrated into chromosomal DNA of the cell. The resulting recombinant cells may then be cultured or subcultured (or passaged, in the case of mammalian cells) from such a culture or a subculture thereof. Methods for transfection, injection and culturing recombinant cells are known to the skilled artisan. Eukaryotic cells in which DNA or RNA may be introduced, include any cells that are transfectable by such DNA or RNA or into which such DNA may be injected. Virtually any eukaryotic cell can serve as a

vehicle for heterologous DNA. Preferred cells are those that can also express the DNA and RNA and most preferred cells are those that can form recombinant or heterologous calcium channels that include one or more subunits encoded by the heterologous DNA. Such cells may be identified empirically or selected from among those known to be readily transfected or Preferred cells for introducing DNA include those that can be transiently or stably transfected and include, but are not limited to, cells of mammalian origin, such as COS cells, mouse L cells, CHO cells, human embryonic kidney cells, African green monkey cells and other such cells known to those of skill in the art, amphibian cells, such as Xenopus laevis occytes, or those of yeast such as Saccharomyces cerevisiae or Pichia pastoris. Preferred cells for expressing injected RNA transcripts or cDNA include Xenopus laevis oöcytes. that are preferred for transfection of DNA are those that can be readily and efficiently transfected. Such cells are known to those of skill in the art or may be empirically identified. Preferred cells include DG44 cells and HEK 293 cells, particularly HEK 293 cells that can be frozen in liquid nitrogen and then thawed and regrown. Such HEK 293 cells are described, for example in U.S. Patent No. 5,024,939 to Gorman [see, also Stillman et al. (1985) Mol. Cell. Biol. 5:2051-2060].

The cells may be used as vehicles for replicating heterologous DNA introduced therein or for expressing the heterologous DNA introduced therein. In certain embodiments, the cells are used as vehicles for expressing the heterologous DNA as a means to produce substantially pure human calcium channel subunits or heterologous calcium channels. Host cells containing the heterologous DNA may be cultured under conditions whereby the calcium channels are expressed. The calcium channel subunits may be purified using protein purification methods known to those of skill in the art. For example, antibodies, such as those provided herein, that specifically bind to one or more of the subunits may be used

for affinity purification of the subunit or calcium channels containing the subunits.

Substantially pure subunits of a human calcium channel $\alpha_{\scriptscriptstyle 1}$ subunits of a human calcium channel, $\alpha_{\scriptscriptstyle 2}$ subunits of a human calcium channel, β subunits of a human calcium channel and γ subunits of а human calcium channel provided. Substantially pure isolated calcium channels that contain at least one of the human calcium channel subunits are also Substantially pure calcium channels that contain a mixture of one or more subunits encoded by the host cell and one or more subunits encoded by heterologous DNA or RNA that has been introduced into the cell are also provided. Substantially pure subtype- or tissue-type specific calcium channels are also provided.

In other embodiments, eukaryotic cells that contain heterologous DNA encoding at least one of an α_1 subunit of a human calcium channel , an α_2 subunit of a human calcium channel, a β subunit of a human calcium channel and a γ subunit of a human calcium channel are provided. In accordance with one preferred embodiment, the heterologous DNA is expressed in the eukaryotic cell and preferably encodes a human calcium channel α_1 subunit.

Expression of heterologous calcium channels: electrophysiology and pharmacology

Electrophysiological methods for measuring calcium channel activity are known to those of skill in the art and are exemplified herein. Any such methods may be used in order to detect the formation of functional calcium channels and to characterize the kinetics and other characteristics of the resulting currents. Pharmacological studies may be combined with the electrophysiological measurements in order to further characterize the calcium channels.

With respect to measurement of the activity of functional heterologous calcium channels, preferably, endogenous ion channel activity and, if desired, heterologous channel activity of channels that do not contain the desired subunits,

of a host cell can be inhibited to a significant extent by chemical, pharmacological and electrophysiological means, including the use of differential holding potential, to increase the S/N ratio of the measured heterologous calcium channel activity.

Thus, various combinations of subunits encoded by the DNA provided herein are introduced into eukaryotic cells. The resulting cells can be examined to ascertain whether functional channels are expressed and to determine the properties of the channels. In particularly preferred aspects, the eukaryotic cell which contains the heterologous DNA expresses it and forms a recombinant functional calcium channel activity. In more preferred aspects, the recombinant calcium channel activity is readily detectable because it is a type that is absent from the untransfected host cell or is of a magnitude and/or pharmacological properties or exhibits biophysical properties not exhibited in the untransfected cell.

The eukaryotic cells can be transfected with various combinations of the subunit subtypes provided herein. The resulting cells will provide a uniform population of calcium channels for study of calcium channel activity and for use in the drug screening assays provided herein. Experiments that have been performed have demonstrated the inadequacy of prior classification schemes.

Preferred among transfected cells is a recombinant eukaryotic cell with a functional heterologous calcium channel. The recombinant cell can be produced by introduction of and expression of heterologous DNA or RNA transcripts encoding an α_1 subunit of a human calcium channel, more preferably also expressing, a heterologous DNA encoding a β subunit of a human calcium channel and/or heterologous DNA encoding an α_2 subunit of a human calcium channel. Especially preferred is the expression in such a recombinant cell of each of the α_1 , β and α_2 subunits encoded by such heterologous DNA or RNA transcripts, and optionally expression of heterologous

DNA or an RNA transcript encoding a γ subunit of a human calcium channel. The functional calcium channels preferably include at least an α_1 subunit and a β subunit of a human calcium channel. Eukaryotic cells expressing these two subunits and also cells expressing additional subunits, have been prepared by transfection of DNA and by injection of RNA transcripts. Such cells have exhibited voltage-dependent calcium channel activity attributable to calcium channels that contain one or more of the heterologous human calcium channel subunits. For example, eukaryotic cells expressing heterologous calcium channels containing an α_2 subunit in addition to the α_1 subunit and a β subunit have been shown to exhibit increased calcium selective ion flow across the cellular membrane in response to depolarization, indicating that the $\alpha_{\rm 2}$ subunit may potentiate calcium channel function. Cells that have been co-transfected with increasing ratios of $\alpha_{\scriptscriptstyle 2}$ to $\alpha_{\scriptscriptstyle 1}$ and the activity of the resulting calcium channels has been measured. The results indicate that α_2 increasing the amount of α_2 -encoding DNA relative to the other transfected subunits increases calcium channel activity.

Eukaryotic cells which express heterologous calcium channels containing at least a human $lpha_1$ subunit, a human etasubunit and a human α_2 subunit are preferred. Eukaryotic cells transformed with a composition containing cDNA or an RNA transcript that encodes an α_1 subunit alone or in combination with a β and/or an α_2 subunit may be used to produce cells that express functional calcium channels. Since recombinant cells expressing human calcium channels containing all of the human subunits encoded by the heterologous cDNA or RNA are especially preferred, it is desirable to inject or transfect such host cells with a sufficient concentration of the subunit-encoding nucleic acids to form calcium channels that contain the human subunits encoded by heterologous DNA or RNA. The precise amounts and ratios of DNA or RNA encoding the subunits may be empirically determined and optimized for a

particular combination of subunits, cells and assay conditions.

In particular, mammalian cells have been transiently and stably transected with DNA encoding one or more human calcium Such cells express heterologous calcium channel subunits. channels that exhibit pharmacological and electrophysiological properties that can be ascribed to human calcium channels. Such cells, however, represent homogeneous populations and the electrophysiological data and pharmacological insights into human calcium channel activity heretofore For example, HEK cells that have been unattainable. transiently transfected with DNA encoding the α_{1E-1} , α_{2b} , and β_{1-3} The resulting cells transiently express these subunits. subunits, which form calcium channels that have properties that appear to be a pharmacologically distinct class of voltage-activated calcium channels distinct from those of L-, N-, T- and P-type channels. The observed α_{1E} currents were insensitive to drugs and toxins previously used to define other classes of voltage-activated calcium channels.

HEK cells that have been transfiently transfected with DNA encoding α_{1B-1} , α_{2b} , and β_{1-2} express heterologous calcium channels that exhibt sensitivity to ω -conotoxin and currents typical of N-type channels. It has been found that alteration of the molar raios of α_{1B-1} , α_{2b} and β_{1-2} introduced into the cells into to achieve equivalent mRNA levels significantly increased the number of receptors per cell, the current density, and affected the K_d for ω -conotoxin.

The electrophyiological properties of these channels produced from α_{1B-1} , α_{2b} , and $\beta_{1\cdot 2}$ was compared with those of channels produced by transiently transfecting HEK cells with DNA encoding α_{1B-1} , α_{2b} and $\beta_{1\cdot 3}$. The channels exhibited similar voltage dependence of activation, substantially identical voltage dependence, similar kinetics of activation and tail currents that could be fit by a single exponential. The voltage dependence of the kinetics of inactivation was significantly different at all voltages examined.

In certain embodiments, the eukaryotic cell with a heterologous calcium channel is produced by introducing into the cell a first composition, which contains at least one RNA transcript that is translated in the cell into a subunit of a human calcium channel. In preferred embodiments, the subunits that are translated include an $lpha_i$ subunit of a human calcium channel. More preferably, the composition that is introduced contains an RNA transcript which encodes an α_1 subunit of a human calcium channel and also contains (1) an RNA transcript which encodes a β subunit of a human calcium channel and/or (2) an RNA transcript which encodes an α_2 subunit of a human calcium channel. Especially preferred is the introduction of RNA encoding an $lpha_{\scriptscriptstyle 1}$, a eta and an $lpha_{\scriptscriptstyle 2}$ human calcium channel subunit, and, optionally, a γ subunit of a human calcium Methods for in vitro transcription of a cloned channel. DNA and injection of the resulting RNA into eukaryotic cells are well known in the art. Transcripts of any of the fulllength DNA encoding any of the subunits of a human calcium channel may be injected alone or in combination with other transcripts into eukaryotic cells for expression in the cells. Amphibian oöcytes are particularly preferred for expression of in vitro transcripts of the human calcium channel subunit cDNA clones provided herein. Amphibian oocytes that express functional heterologous calcium channels have been produced by this method.

Assays and Clinical uses of the cells and calcium channels Assays

Assays for identifying compounds that modulate calcium channel activity

Among the uses for eukaryotic cells which recombinantly express one or more subunits are assays for determining whether a test compound has calcium channel agonist or antagonist activity. These eukaryotic cells may also be used to select from among known calcium channel agonists and antagonists those exhibiting a particular calcium channel

subtype specificity and to thereby select compounds that have potential as disease- or tissue-specific therapeutic agents.

In vitro methods for identifying compounds, such as calcium channel agonist and antagonists, that modulate the activity of calcium channels using eukaryotic cells that express heterologous human calcium channels are provided.

In particular, the assays use eukaryotic cells that express heterologous human calcium channel subunits encoded by heterologous DNA provided herein, for screening potential calcium channel agonists and antagonists which are specific for human calcium channels and particularly for screening for compounds that are specific for particular human calcium channel subtypes. Such assays may be used in conjunction with methods of rational drug design to select among agonists and antagonists, which differ slightly in structure, those particularly useful for modulating the activity of human calcium channels, and to design or select compounds that exhibit subtype- or tissue- specific calcium assays should antagonist and agonist activities. These accurately predict the relative therapeutic efficacy of a compound for the treatment of certain disorders in humans. In addition, since subtype-and tissue-specific calcium channel subunits are provided, cells with tissue- specific or subtypespecific recombinant calcium channels may be prepared and used in assays for identification of human calcium channel tissueor subtype-specific drugs.

Desirably, the host cell for the expression of calcium channel subunits does not produce endogenous calcium channel subunits of the type or in an amount that substantially interferes with the detection of heterologous calcium channel subunits in ligand binding assays or detection of heterologous calcium channel function, such as generation of calcium current, in functional assays. Also, the host cells preferably should not produce endogenous calcium channels which detectably interact with compounds having, at physiological concentrations (generally nanomolar or picomolar

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concentrations), affinity for calcium channels that contain one or all of the human calcium channel subunits provided herein.

With respect to ligand binding assays for identifying a compound which has affinity for calcium channels, cells are employed which express, preferably, at least a heterologous α_1 subunit. Transfected eukaryotic cells which express at least an α_1 subunit may be used to determine the ability of a test compound to specifically bind to heterologous calcium channels by, for example, evaluating the ability of the test compound to inhibit the interaction of a labeled compound known to specifically interact with calcium channels. Such ligand binding assays may be performed on intact transfected cells or membranes prepared therefrom.

The capacity of a test compound to bind to or otherwise interact with membranes that contain heterologous calcium channels or subunits thereof may be determined by using any appropriate method, such as competitive binding analysis, such as Scatchard plots, in which the binding capacity of such membranes is determined in the presence and absence of one or more concentrations of a compound having known affinity for the calcium channel. Where necessary, the results may be compared to a control experiment designed in accordance with methods known to those of skill in the art. For example, as a negative control, the results may be compared to those of assays of an identically treated membrane preparation from host cells which have not been transfected with one or more subunit-encoding nucleic acids.

The assays involve contacting the cell membrane of a recombinant eukaryotic cell which expresses at least one subunit of a human calcium channel, preferably at least an α_1 subunit of a human calcium channel, with a test compound and measuring the ability of the test compound to specifically bind to the membrane or alter or modulate the activity of a heterologous calcium channel on the membrane.

In preferred embodiments, the assay uses a recombinant cell that has a calcium channel containing an α_1 subunit of a human calcium channel in combination with a β subunit of a human calcium channel and/or an α_2 subunit of a human calcium channel. Recombinant cells expressing heterologous calcium channels containing each of the α_1 , β and α_2 human subunits, and, optionally, a γ subunit of a human calcium channel are especially preferred for use in such assays.

the assays for identifying In certain embodiments, compounds that modulate calcium channel activity are practiced by measuring the calcium channel activity of a eukaryotic cell having a heterologous, functional calcium channel when such cell is exposed to a solution containing the test compound and a calcium channel-selective ion and comparing the measured calcium channel activity to the calcium channel activity of the same cell or a substantially identical control cell in a solution not containing the test compound. The cell is maintained in a solution having a concentration of calcium channel-selective ions sufficient to provide an inward current when the channels open. Rcombinant cells expressing calcium channels that include each of the α_1 , β and α_2 human subunits, and, optionally, a γ subunit of a human calcium channel, are especially preferred for use in such assays. Methods for practicing such assays are known to those of skill in the art. For example, for similar methods applied with Xenopus laevis oöcytes and acetylcholine receptors, see, Mishina et al. [(1985) Nature 313:364] and, with such oöcytes and sodium channels [see, Noda et al. (1986) Nature 322:826-828]. For similar studies which have been carried out with the acetylcholine receptor, see, e.g., Claudio et al. [(1987) Science 238:1688-1694].

Functional recombinant or heterologous calcium channels may be identified by any method known to those of skill in the art. For example, electrophysiological procedures for measuring the current across an ion-selective membrane of a cell, which are well known, may be used. The amount and

duration of the flow of calcium-selective ions through heterologous calcium channels of a recombinant cell containing DNA encoding one or more of the subunits provided herein has been measured using electrophysiological recordings using a two electrode and the whole-cell patch clamp techniques. order to improve the sensitivity of the assays, known methods can be used to eliminate or reduce non-calcium currents and calcium currents resulting from endogenous calcium channels, when measuring calcium currents through recombinant channels. For example, the DHP Bay K 8644 specifically enhances L-type calcium channel function by increasing the duration of the open state of the channels [see, e.g., Hess, J.B., et al. (1984) Nature 311:538-544]. Prolonged opening of the channels results in calcium currents of increased magnitude duration. Tail currents can be observed upon repolarization of the cell membrane after activation of ion channels by a depolarizing voltage command. The opened channels require a finite time to close or "deactivate" upon repolarization, and the current that flows through the channels during this period is referred to as a tail current. Because Bay K 8644 prolongs opening events in calcium channels, it tends to prolong these tail currents and make them more pronounced.

In practicing these assays, stably or transiently transfected cells or injected cells that express voltage-dependent human calcium channels containing one or more of the subunits of a human calcium channel desirably may be used in assays to identify agents, such as calcium channel agonists and antagonists, that modulate calcium channel activity. Functionally testing the activity of test compounds, including compounds having unknown activity, for calcium channel agonist or antagonist activity to determine if the test compound potentiates, inhibits or otherwise alters the flow of calcium ions or other ions through a human calcium channel can be accomplished by (a) maintaining a eukaryotic cell which is transfected or injected to express a heterologous functional calcium channel capable of regulating the flow of calcium

channel-selective ions into the cell in a medium containing calcium channel-selective ions (i) in the presence of and (ii) in the absence of a test compound; (b) maintaining the cell under conditions such that the heterologous calcium channels are substantially closed and endogenous calcium channels of the cell are substantially inhibited (c) depolarizing the membrane of the cell maintained in step (b) to an extent and for an amount of time sufficient to cause (preferably, substantially only) the heterologous calcium channels to become permeable to the calcium channel-selective ions; and (d) comparing the amount and duration of current flow into the cell in the presence of the test compound to that of the current flow into the cell, or a substantially similar cell, in the absence of the test compound.

The assays thus use cells, provided herein, that express functional calcium channels and measure heterologous functionally, such as electrophysiologically, the ability of a test compound to potentiate, antagonize or otherwise modulate the magnitude and duration of the flow of calcium channel-selective ions, such as Ca** or Ba**, through the heterologous functional channel. The amount of current which flows through the recombinant calcium channels of a cell may be determined directly, such as electrophysiologically, or by which independent reaction occurs monitoring an intracellularly and which is directly influenced in a calcium (or other) ion dependent manner. Any method for assessing the activity of a calcium channel may be used in conjunction with the cells and assays provided herein. in one embodiment of the method for testing a compound for its ability to modulate calcium channel activity, the amount of current is measured by its modulation of a reaction which is sensitive to calcium channel-selective ions and uses a eukaryotic cell which expresses a heterologous calcium channel contains a transcriptional control operatively linked for expression to a structural gene that encodes an indicator protein. The transcriptional control

element used for transcription of the indicator gene is responsive in the cell to a calcium channel-selective ion, such as Ca²⁺ and Ba⁺. The details of such transcriptional based assays are described in commonly owned PCT International Patent Application No. PCT/US91/5625, filed August 7, 1991, which claims priority to copending commonly owned allowed U.S. Application Serial No. 07/563,751, filed August 7, 1990; see also, commonly owned published PCT International Patent Application PCT US92/11090, which corresponds to co-pending U.S. Applications Serial Nos. 08/229,150 and 08/244,985.

Assays for diagnosis of LES

LES is an autoimmune disease characterized by an insufficient release of acetylcholine from motor nerve terminals which normally are responsive to nerve impulses. Immunoglobulins (IgG) from LES patients block individual voltage-dependent calcium channels and thus inhibit calcium channel activity [Kim and Neher, Science 239:405-408 (1988)]. A diagnostic assay for Lambert Eaton Syndrome (LES) is provided herein. The diagnostic assay for LES relies on the immunological reactivity of LES IgG with the human calcium channels or particular subunits alone or in combination or expressed on the surface of recombinant cells. For example, such an assay may be based on immunoprecipitation of LES IgG by the human calcium channel subunits and cells that express such subunits provided herein.

Clinical applications

In relation to therapeutic treatment of various disease states, the availability of DNA encoding human calcium channel subunits permits identification of any alterations in such genes (e.g., mutations) which may correlate with the occurrence of certain disease states. In addition, the creation of animal models of such disease states becomes possible, by specifically introducing such mutations into synthetic DNA fragments can then be introduced into laboratory animals or in vitro assay systems to determine the effects thereof.

Also, genetic screening can be carried out using the nucleotide sequences as probes. Thus, nucleic acid samples from subjects having pathological conditions suspected of involving alteration/modification of any one or more of the calcium channel subunits can be screened with appropriate probes to determine if any abnormalities exist with respect to any of the endogenous calcium channels. Similarly, subjects having a family history of disease states related to calcium channel dysfunction can be screened to determine if they are also predisposed to such disease states.

EXAMPLES

The following examples are included for illustrative purposes only and are not intended to limit the scope of the invention.

EXAMPLE I: PREPARATION OF LIBRARIES USED FOR ISOLATION OF DNA ENCODING HUMAN NEURONAL VOLTAGE-DEPENDENT CALCIUM CHANNEL SUBUNITS

A. RNA Isolation

1. IMR32 cells

IMR32 cells were obtained from the American Type Culture Collection (ATCC Accession No. CCL127, Rockville, MD) in DMEM. 10% fetal bovine serum, grown penicillin/streptomycin (GIBCO, Grand Island, NY) plus 1.0 mM dibutyryl cAMP (dbcAMP) for ten days. Total RNA was isolated from the cells according to the procedure described by H.C. Birnboim [(1988) Nucleic Acids Research 16:1487-1497]. Poly(A*) RNA was selected according to standard procedures [see, e.g., Sambrook et al. (1989) Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory Press; pg. 7.26-7.29].

2. Human thalamus tissue

Human thalamus tissue (2.34 g), obtained from the National Neurological Research Bank, Los Angeles, CA, that had been stored frozen at -70°C was pulverized using a mortar and pestle in the presence of liquid nitrogen and the cells were lysed in 12 ml of lysis buffer (5 M guanidinium isothiocyanate, 50 mM TRIS, pH 7.4, 10 mM EDTA, 5% β -

mercaptoethanol). Lysis buffer was added to the lysate to yield a final volume of 17 ml. N-laurylsarcosine and CsCl were added to the mixture to yield final concentrations of 4% and 0.01 g/ml, respectively, in a final volume of 18 ml.

The sample was centrifuged at 9,000 rpm in a Sorvall SS34 rotor for 10 min at room temperature to remove the insoluble material as a pellet. The supernatant was divided into two equal portions and each was layered onto a 2-ml cushion of a solution of 5.7 M CsCl, 0.1 M EDTA contained in separate centrifuge tubes to yield approximately 9 ml per tube. The samples were centrifuged in an SW41 rotor at 37,000 rpm for 24 h at 20°C.

After centrifugation, each RNA pellet was resuspended in 3 ml ETS (10 mM TRIS, pH 7.4, 10 mM EDTA, 0.2% SDS) and combined into a single tube. The RNA was precipitated with 0.25 M NaCl and two volumes of 95% ethanol.

The precipitate was collected by centrifugation and resuspended in 4 ml PK buffer (0.05 M TRIS, pH 8.4, 0.14 M NaCl, 0.01 M EDTA, 1% SDS). Proteinase K was added to the sample to a final concentration of 200 μ g/ml. The sample was incubated at 22°C for 1 h, followed by extraction with an equal volume of phenol:chloroform:isoamylalcohol (50:48:2) two times, followed by one extraction with an equal volume of chloroform: isoamylalcohol (24:1). The RNA was precipitated with ethanol and NaCl. The precipitate was resuspended in 400 μ l of ETS buffer. The yield of total RNA was approximately 1.0 mg. Poly A* RNA (30 μ g) was isolated from the total RNA according to standard methods as stated in Example I.A.1.

B. Library Construction

Double-stranded cDNA was synthesized according to standard methods [see, e.g., Sambrook et al. (1989) IN: Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory Press, Chapter 8]. Each library was prepared in substantially the same manner except for differences in: 1) the oligonucleotide used to prime the first strand cDNA synthesis, 2) the adapters that were attached to the double-

stranded cDNA, 3) the method used to remove the free or unused adapters, and 4) the size of the fractionated cDNA ligated into the λ phage vector.

1. IMR32 cDNA library #1

Single-stranded cDNA was synthesized using IMR32 poly(A*) RNA (Example I.A.1.) as a template and was primed using oligo (dT) $_{12-18}$ (Collaborative Research Inc., Bedford, MA): The single-stranded cDNA was converted to double-stranded cDNA and the yield was approximately $2\mu g$. EcoI adapters:

- 5'-AATTCGGTACGTACACTCGAGC-3' = 22-mer (SEQ ID No.15)
- 3'- GCCATGCATGTGAGCTCG-5' = 18-mer (SEQ ID No.16) also containing SnaBI and XhoI restriction sites were then added to the double-stranded cDNA according to the following procedure.

a. Phosphorylation of 18-mer

The 18-mer was phosphorylated using standard methods [see, e.g., Sambrook et al. (1989) IN: Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory Press, Chapter 8] by combining in a 10 μ l total volume the 18-mer (225 pmoles) with [\$^{32}P] γ -ATP (7000 Ci/mmole; 1.0 μ l) and kinase (2 U) and incubating at 37° C for 15 minutes. After incubation, 1 μ l 10 mM ATP and an additional 2 U of kinase were added and incubated at 37°C for 15 minutes. Kinase was then inactivated by boiling for 10 minutes.

b. Hybridization of 22-mer

The 22-mer was hybridized to the phosphorylated 18-mer by addition of 225 pmoles of the 22-mer (plus water to bring volume to 15 μ l), and incubation at 65°C for 5 minutes. The reaction was then allowed to slow cool to room temperature.

The adapters were thus present at a concentration of 15 pmoles/ μ l, and were ready for cDNA-adapter ligation.

Ligation of adapters to cDNA

After the EcoRI, SnaBI, XhoI adapters were ligated to the double-stranded cDNA using a standard protocol [see, e.g., Sambrook et al. (1989) IN: Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory Press, Chapter 8], the

ligase was inactivated by heating the mixture to 72°C for 15 minutes. The following reagents were added to the cDNA ligation reaction and heated at 37°C for 30 minutes: cDNA ligation reaction (20 μ l), water (24 μ l), 10x kinase buffer (3 μ l), 10 mM ATP (1 μ l) and kinase (2 μ l of 2 U/ μ l). The reaction was stopped by the addition of 2 μ l 0.5M EDTA, followed by one phenol/chloroform extraction and one chloroform extraction.

d. Size Selection and Packaging of cDNA

The double-stranded cDNA with the EcoRI, SnaBI, XhoI adapters ligated was purified away from the free or unligated adapters using a 5 ml Sepharose CL-4B column (Sigma, St. 100 μ l fractions were collected and those Louis, MO). containing the CDNA, determined by monitoring radioactivity, were pooled, ethanol precipitated, resuspended in TE buffer and loaded onto a 1% agarose gel. electrophoresis, the gel was stained with ethidium bromide and the 1 to 3 kb fraction was cut from the gel. embedded in the agarose was eluted using the "Geneluter Electroelution System" (Invitrogen, San Diego, CA). eluted cDNA was collected by ethanol precipitation resuspended in TE buffer at 0.10 pmol/ μ l. The cDNA was ligated to 1 μ g of EcoRI digested, dephosphorylated λ qt11 in a 5 μ l reaction volume at a 2- to 4- fold molar excess ratio of cDNA over the \(\lambda\gt11\) vector. The ligated \(\lambda\gt11\) containing the cDNA insert was packaged into λ phage virions in vitro using the Gigapack (Stratagene, La Jolla, CA) kit. packaged phage were plated on an E. coli Y1088 bacterial lawn in preparation for screening.

2. IMR32 cDNA library #2

This library was prepared as described (Example I.B.1.) with the exception that 3 to 9 kb cDNA fragments were ligated into the λ gtll phage vector rather than the 1 to 3 kb fragments.

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3. IMR32 cDNA library #3

IMR32 cell poly(A*) RNA (Example I.A.1.) was used as a template to synthesize single-stranded cDNA. The primers for the first strand cDNA synthesis were random primers (hexadeoxy-nucleotides [pd(N) $_6$] Cat #5020-1, Clontech, Palo Alto, CA). The double-stranded cDNA was synthesized, EcoRI, SnaBI, XhoI adapters were added to the cDNA, the unligated adapters were removed, and the double-stranded cDNA with the ligated adapters was fractionated on an agarose gel, as described in Example I.B.1. The cDNA fraction greater than 1.8 kb was eluted from the agarose, ligated into $\lambda gtl1$, packaged, and plated into a bacterial lawn of Y1088 (as described in Example I.B.1.).

4. IMR32 cDNA library #4

IMR32 cell poly(A*) RNA (Example I.A.1.) was used as a template to synthesize single-stranded cDNA. The primers for the first strand cDNA synthesis were oligonucleotides: 89-365a specific for the $\alpha_{\rm 1D}$ (VDCC III) type $\alpha_{\rm 1}$ -subunit (see Example II.A.) coding sequence (the complementary sequence of nt 2927 to 2956, SEQ ID No. 1), 89-495 specific for the $\alpha_{\rm 1C}$ (VDCC II) type $\alpha_{\rm 1}$ -subunit (see Example II.B.) coding sequence (the complementary sequence of nt 852 to 873, SEQ ID No. 3), and 90-12 specific for the $\alpha_{\rm 1C}$ -subunit coding sequence (the complementary sequence of nt 2496 to 2520, SEQ ID No. 3). The cDNA library was then constructed as described (Example I.B.3), except that the cDNA size-fraction greater than 1.5 kb was eluted from the agarose rather than the greater than 1.8 kb fraction.

5. IMR32 cDNA library #5

The cDNA library was constructed as described (Example I.B.3.) with the exception that the size-fraction greater than 1.2 kb was eluted from the agarose rather than the greater than 1.8 kb fraction.

6. Human thalamus cDNA library #6

Human thalamus poly (A^+) RNA (Example I.A.2.) was used as a template to synthesize single-stranded cDNA. Oligo (dT) was

used to prime the first strand synthesis (Example I.B.1.). The double-stranded cDNA was synthesized (Example I.B.1.) and EcoRI, KpnI, NcoI adapters of the following sequence:

- 5' CCATGGTACCTTCGTTGACG 3'= 20-mer (SEQ ID NO. 17)
- 3' GGTACCATGGAAGCAACTGCTTAA 5'= 24-mer (SEQ ID NO. 18) were ligated to the double-stranded cDNA as described (Example I.B.1.) with the 20-mer replacing the 18-mer and the 24-mer replacing the 22-mer. The unligated adapters were removed by passing the cDNA-adapter mixture through a 1 ml Bio Gel A-50 (Bio-Rad Laboratories, Richmond, CA) column. Fractions (30 μ l) were collected and 1 μ l of each fraction in the first peak of radioactivity was electrophoresed on a 1% agarose gel. After electrophoresis, the gel was dried on a vacuum gel drier and exposed to x-ray film. The fractions containing cDNA fragments greater than 600 bp were pooled, ethanol precipitated, and ligated into Agt11 (Example I.B.1.). construction of the cDNA library was completed as described (Example I.B.1.).

C. Hybridization and Washing Conditions

Hybridization of radiolabelled nucleic to immobilized DNA for the purpose of screening cDNA libraries, DNA Southern transfers, or northern transfers was routinely performed in standard hybridization conditions [hybridization: 50% deionized formamide, 200 μ g/ml sonicated sperm DNA (Cat #223646, Boehringer Biochemicals, Indianapolis, IN), 5 x SSPE, 5 x Denhardt's, 42° C.; wash: 0.2 x SSPE, 0.1% SDS, 65° C]. The recipes for SSPE and Denhardt's and the preparation of deionized formamide are described, for example, in Sambrook et al. (1989) Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory Press, Chapter 8). In some hybridizations, lower stringency conditions were used in that 10% deionized formamide replaced deionized formamide described for the standard hybridization conditions.

The washing conditions for removing the non-specific probe from the filters was either high, medium, or low stringency as described below:

- high stringency: 0.1 x SSPE, 0.1% SDS, 65°C
- medium stringency: 0.2 x SSPE, 0.1% SDS, 50°C
- low stringency: 1.0 x SSPE, 0.1% SDS, 50°C.

It is understood that equivalent stringencies may be achieved using alternative buffers, salts and temperatures.

ISOLATION OF DNA ENCODING THE HUMAN NEURONAL EXAMPLE II: CALCIUM CHANNEL α_1 SUBUNIT

Isolation of DNA encoding the $lpha_{ ext{ip}}$ subunit

Reference list of partial α_{lD} cDNA clones

Numerous $\alpha_{\text{1D}}\text{-specific cDNA}$ clones were isolated in order to characterize the complete $\alpha_{\text{\tiny 1D}}$ coding sequence plus portions of the 5' and 3' untranslated sequences. SEQ ID No. 1 shows the complete α_{iD} DNA coding sequence, plus 510 nucleotides of $\alpha_{\mbox{\tiny 1D}}$ 5' untranslated sequence ending in the guanidine nucleotide adjacent to the adenine nucleotide of the proposed initiation of translation as well as 642 nucleotides of 3' untranslated sequence. Also shown in SEQ ID No. 1 is the deduced amino acid sequence. A list of partial cDNA clones used to characterize the α_{1D} sequence and the nucleotide position of each clone relative to the full-length α_{1D} cDNA sequence, which is set forth in SEQ ID No. 1, is shown below. The isolation and characterization of these clones are described below (Example II.A.2.).

IMR32	7 744	
IMK32	1.144	nt 1 to 510 of SEQ ID No. 1
		5' untranslated sequence,
		nt 511 to 2431, SEQ ID No. 1
IMR32*	1.136	nt 1627 to 2988, SEQ ID No. 1
		nt 1 to 104 of SEQ ID No. 2
		additional exon,
IMR32@	1.80	nt 2083 to 6468, SEQ ID No. 1
IMR32#	1.36	nt 2857 to 4281, SEQ ID No. 1
IMR32	1.163	nt 5200 to 7635, SEQ ID No. 1

- * 5' of nt 1627, IMR32 1.136 encodes an intron and an additional exon described in Example II.A.2.d.
 - @ IMR32 1.80 contains two deletions, nt 2984 to 3131 and nt 5303 to 5349 (SEQ ID No. 1). The 148 nt deletion (nt 2984 to 3131) was corrected by performing a polymerase chain reaction described in Example II.A.3.b.
 - # IMR32 1.36 contains a 132 nt deletion (nt 3081 to 3212).
 - 2. Isolation and characterization of individual clones listed in Example II.A.1.

a. IMR32 1.36

Two million recombinants of the IMR32 cDNA library #1 (Example I.B.1.) were screened in duplicate at a density of approximately 200,000 plaques per 150 mm plate using a mixture of radiolabelled fragments of the coding region of the rabbit skeletal muscle calcium channel α_1 cDNA [for the sequence of the rabbit skeletal muscle calcium channel α_1 subunit cDNA, see, Tanabe et al. (1987). Nature 328:313-

318]:	Fragment	Nucleotides
	KpnI-EcoRI	-78 to 1006
	EcoRI-XhoI	1006 to 2653
	ApaI-ApaI	3093 to 4182
	Balli-Sacl	4487 to 5310

The hybridization was performed using low stringency hybridization conditions (Example I.C.) and the filters were washed under low stringency (Example I.C.). Only one α_{1D} -specific recombinant (IMR32 1.36) of the 2 x 10 6 screened was identified. IMR32 1.36 was plaque purified by standard methods (J. Sambrook et al. (1989) Molecular Cloning, A Laboratory Manual, Cold Spring Harbor Laboratory Press, Chapter 8) subcloned into pGEM3 (Promega, Madison, WI) and characterized by DNA sequencing.

b. IMR32 1.80

Approximately 1 x 10^6 recombinants of the IMR32 cDNA library #2 (Example I.B.2.) were screened in duplicate at a

density of approximately 100,000 plaques per 150 mm plate using the IMR32 1.36 cDNA fragment (Example II.A.1) as a probe. Standard hybridization conditions were used, and the filters were washed under high stringency (Example I.C.). Three positive plaques were identified one of which was IMR32 1.80. IMR32 1.80 was plaque purified by standard methods, restriction mapped, subcloned, and characterized by DNA sequencing.

c. IMR32 1.144

Approximately 1 x 10^6 recombinants of the IMR32 cDNA library #3 (Example I.B.3) were screened with the EcoRI-PvuII fragment (nt 2083 to 2518, SEQ ID No. 1) of IMR32 1.80. hybridization was performed using standard hybridization conditions (Example I.C.) and the filters were washed under high stringency (Example I.C.). Three positive plaques were identified one of which was IMR32 1.144. plaque purified, restriction mapped, and the cDNA insert was subcloned into pGEM7Z (Promega, Madison, WI) and characterized by DNA sequencing. This characterization revealed that IMR32 1.144 has a series of ATG codons encoding seven possible initiating methionines (nt 511 to 531, SEQ ID No. 1). Nucleic acid amplification analysis, and DNA sequencing of cloned nucleic acid amplification analysis products encoding these seven ATG codons confirmed that this sequence is present in the α_{1D} transcript expressed in dbcAMP-induced IMR32 cells.

d. IMR32 1.136

Approximately 1 x 10⁶ recombinants of the IMR32 cDNA library #4 (Example I.B.4) were screened with the EcoRI-PvuII fragment (nt 2083 to 2518, SEQ ID No. 1) of IMR32 1.80 (Example II.A.1.). The hybridization was performed using standard hybridization conditions (Example I.C.) and the filters were washed under high stringency (Example I.C.). Six positive plaques were identified one of which was IMR32 1.136. IMR32 1.136 was plaque purified, restriction mapped, and the cDNA insert was subcloned into a standard plasmid vector, pSP72 (Promega, Madison, WI.), and characterized by DNA

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sequencing. This characterization revealed that IMR32 1.136 encodes an incompletely spliced $\alpha_{\rm 1D}$ transcript. The clone contains nucleotides 1627 to 2988 of SEQ ID No. 1 preceded by an approximate 640 bp intron. This intron is then preceded by a 104 nt exon (SEQ ID No. 2) which is an alternative exon encoding the IS6 transmembrane domain [see, e.g., Tanabe et al. (1987) Nature 328:313-318 for a description of the IS1 to IVS6 transmembrane terminology] of the $\alpha_{\rm 1D}$ subunit and can replace nt 1627 to 1730, SEQ ID No. 1, to produce a completely spliced $\alpha_{\rm 1D}$ transcript.

e. IMR32 1.163

Approximately 1 x 10 6 recombinants of the IMR32 cDNA library #3 (Example I.B.3.) were screened with the NcoI-XhoI fragment of IMR32 1.80 (Example II.A.1.) containing nt 5811 to 6468 (SEQ ID No. 1). The hybridization was performed using standard hybridization conditions (Example I.C.) and the filters were washed under high stringency (Example I.C.). Three positive plaques were identified one of which was IMR32 1.163. IMR32 1.163 was plaque purified, restriction mapped, and the cDNA insert was subcloned into a standard plasmid vector, pSP72 (Promega, Madison, WI.), and characterized by DNA sequencing. This characterization revealed that IMR32 1.163 contains the α_{1D} termination codon, nt 6994 to 6996 (SEQ ID No. 1).

3. Construction of a full-length α_{1D} cDNA [pVDCCIII(A)]

 $\alpha_{\rm 1D}$ cDNA clones IMR32 1.144, IMR32 1.136, IMR32 1.80, and IMR32 1.163 (Example II.A.2.) overlap and include the entire $\alpha_{\rm 1D}$ coding sequence, nt 511 to 6993 (SEQ ID No. 1), with the exception of a 148 bp deletion, nt 2984 to 3131 (SEQ ID No. 1). Portions of these partial cDNA clones were ligated to generate a full-length $\alpha_{\rm 1D}$ cDNA in a eukaryotic expression vector. The resulting vector was called pVDCCIII(A). The construction of pVDCCIII(A) was performed in four steps described in detail below: (1) the construction of pVDCCIII/5' using portions of IMR32 1.144, IMR32 1.136, and

IMR32 1.80, (2) the construction of pVDCCIII/5'.3 that corrects the 148 nt deletion in the IMR32 1.80 portion of pVDCCIII/5', (3) the construction of pVDCCIII/3'.1 using portions of IMR32 1.80 and IMR32 1.163, and (4) the ligation of a portion of the pVDCCIII/5'.3 insert, the insert of pVDCCIII/3'.1, and pcDNA1 (Invitrogen, San Diego, CA) to form pVDCCIII(A). The vector pcDNA1 is a eukaryotic expression vector containing a cytomegalovirus (CMV) promoter which is a constitutive promoter recognized by mammalian host cell RNA polymerase II.

Each of the DNA fragments used in preparing the full-length construct was purified by electrophoresis through an agarose gel onto DE81 filter paper (Whatman, Clifton, NJ) and elution from the filter paper using 1.0 M NaCl, 10 mM TRIS, pH 8.0, 1 mM EDTA. The ligations typically were performed in a 10 μ l reaction volume with an equal molar ratio of insert fragment and a two-fold molar excess of the total insert relative to the vector. The amount of DNA used was normally about 50 ng to 100 ng.

a. pVDCCIII/5'

To construct pVDCCIII/5', IMR32 1.144 (Example II.A.2.c.) was digested with XhoI and EcoRI and the fragment containing the vector (pGEM7Z), α_{1D} nt 1 to 510 (SEQ ID No. 1), and α_{1D} nt (SEQ ID No. 1) was isolated by 1732 The EcoRI-ApaI fragment of IMR32 1.136 electrophoresis. (Example II.A.2.d.) nucleotides 1733 to 2671 (SEQ ID No. 1) was isolated, and the Apal-HindIII fragment of IMR32 1.80 (Example II.A.2.b.), nucleotides 2672 to 4492 (SEQ ID No. 1) The three DNA clones were ligated to form was isolated. pVDCCIII/5' containing nt 1 to 510 (5' untranslated sequence; SEQ ID No. 1) and nt 511 to 4492 (SEQ ID No. 1).

b. pvDccIII/5'.3

Comparison of the IMR32 1.36 and IMR32 1.80 DNA sequences revealed that these two cDNA clones differ through the α_{1D} coding sequence, nucleotides 2984 to 3212. nucleic acid amplification analysis of IMR32 1.80 and dbcAMP-induced

(1.0 mM, 10 days) IMR32 cytoplasmic RNA (isolated according to Ausubel, F.M. et al. (Eds) (1988) Current Protocols in Molecular Biology, John Wiley and Sons, New York) revealed that IMR32 1.80 had a 148 nt deletion, nt 2984 to 3131 (SEO ID No. 1), and that IMR32 1.36 had a 132 nt deletion, nt 3081 to 3212. To perform the nucleic acid amplification analysis, the amplification reaction was primed with α_{n} -specific oligonucleotides 112 (nt 2548 to 2572, SEQ ID No. 1) and 311 (the complementary sequence of nt 3928 to 3957, SEQ ID No. 1). These products were then reamplified using α_{n} -specific oligonucleotides 310 (nt 2583 to 2608 SEQ ID No. 1) and 312 (the complementary sequence of nt 3883 to 3909). reamplified product, which contains AccI and BglII restriction sites, was digested with AccI and BglII and the AccI-BglII fragment, nt 2765 to 3890 (SEQ ID No. 1) was cloned into AccIdigested pVDCCIII/5' to replace AccI-BglII the pVDCCIII/5' fragment that had the deletion. This new construct was named pVDCCIII/5'.3. DNA sequence determination of pVDCCIII/5'.3 through the amplified region confirmed the 148 nt deletion in IMR32 1.80.

c. pVDCCIII/3'.1

To construct pVDCCIII/3'.1, the cDNA insert of IMR32 1.163 (Example II.A.2.e.) was subcloned into pBluescript II (Stratagene, La Jolla, CA) as an XhoI fragment. sites on the cDNA fragment were furnished by the adapters used to construct the cDNA library (Example I.B.3.). was oriented such that the translational orientation of the insert of IMR32 1.163 was opposite to that of the lacZ gene in the plasmid, as confirmed by analysis restriction enzyme digests of the resulting plasmid. This was done to preclude the possibility of expression of α_{n} sequences in DH5 α cells transformed with this plasmid due to fusion with the lacz gene. This plasmid was then digested with HindIII and BglII and the HindIII - BglII fragment (the HindIII site comes from the vector and the BglII site is at nt 6220, SEQ ID No. 1) was eliminated, thus deleting nt 5200 to 6220 (SEQ ID

No. 1) of the IMR32 1.163 clone and removing this sequence from the remainder of the plasmid which contained the 3' BglII - XhoI fragment, nt 6221 to 7635 (SEQ ID No. 1). pVDCCIII/3'.1 was then made by splicing together the HindIII-PvuII fragment from IMR32 1.80 (nucleotides 4493-5296, SEQ ID No. 1), the PvuII - BglII fragment of IMR32 1.163 (nucleotides 5294 to 6220, SEQ ID No. 1) and the HindIII-BglII-digested pBluescript plasmid containing the 3' BglII/XhoI IMR32 1.163 fragment (nt 6221 to 7635, SEQ ID No. 1).

d. pVDCCIII(A): the full-length α_{1D} construct

To construct pVDCCIII(A), the DraI-HindIII fragment (5' untranslated sequence nt 330 to 510, SEQ ID No. 1 and coding sequence nt 511 to 4492, SEQ ID No. 1) of pVDCCIII/5'.3 (Example II.A.3.b.) was isolated; the HindIII-XhoI fragment of pVDCCIII/3'.1 (containing nt 4493 to 7635, SEQ ID No. 1, plus the XhoI site of the adapter) (Example II.A.3.c.) isolated; and the plasmid vector, pcDNA1, was digested with EcoRV and XhoI and isolated on an agarose gel. The three DNA fragments were ligated and MC1061-P3 (Invitrogen, San Diego, was transformed. Isolated clones were analyzed by restriction mapping and DNA sequencing and pVDCCIII(A) was identified which had the fragments correctly ligated together: DraI-HindIII, HindIII-XhoI, XhoI-EcoRV with the blunt-end DraI and EcoRV site ligating together to form the circular plasmid.

The amino-terminus of the α_{1D} subunit is encoded by the seven consecutive 5' methionine codons (nt 511 to 531, SEQ ID No. 1). This 5' portion plus nt 532 to 537, encoding two lysine residues, were deleted from pVDCCIII(A)

and replaced with an efficient ribosomal binding site (5'-ACCACC-3') to form pVDCCIII.RBS(A). Expression experiments in which transcripts of this construct were injected into Xenopus laevis occytes did not result in an enhancement in the recombinant voltage-dependent calcium channel expression level relative to the level of expression in occytes injected with transcripts of pVDCCIII(A).

B. Isolation of DNA encoding the α_{1c} subunit

1. Reference List of Partial α_{ic} cDNA clones

Numerous α_{1c} -specific cDNA clones were isolated in order to characterize the α_{ic} coding sequence, the α_{ic} initiation of translation, and an alternatively spliced region of α_{ic} . ID No. 3 sets forth one α_{1c} coding sequence (α_{1c-1}) and deduced amino acid sequence; SEQ ID No. 36 sets forth another splice variant designated $\alpha_{\text{1c-2}}$. SEQ ID No. 4 and No. 5 encode two possible amino terminal ends of an α_{1c} splice variant. No. 6 encodes an alternative exon for the IV S3 transmembrane domain. Other α_{1c} variants can be constructed by selecting the alternative amino terminal ends in place of the ends in SEQ ID No. 3 or 36 and/or inserting the alternative exon (SEQ ID No. 6) in the appropriate location, such as in SEQ ID NO. 3 in In addition, place of nucleotides 3904-3987. nucleotide sequence (nucleotides 1391-1465 in SEQ ID No. 3) can be deleted or inserted to produce an alternative α_{ic} splice variant.

Shown below is a list of clones used to characterize the α_{1c} sequence and the nucleotide position of each clone relative to the characterized α_{1c} sequence (SEQ ID No. 3). The isolation and characterization of these cDNA clones are described below (Example II.B.2).

IMR32	1.66	nt 1 to 916, SEQ ID No. 3
		nt 1 to 132, SEQ ID No. 4
IMR32	1.157	nt 1 to 873, SEQ ID No. 3
		nt 1 to 89, SEQ ID No. 5
IMR32	1.67	nt 50 to 1717, SEQ ID No. 3
*IMR32	1.86	nt 1366 to 2583, SEQ ID No. 3
^e 1.16G		nt 758 to 867, SEQ ID No. 3
IMR32	1.37	nt 2804 to 5904, SEQ ID No. 3
CNS	1.30	nt 2199 to 3903, SEQ ID No. 3
		nt 1 to 84 of alternative exon,
		SEQ ID No. 6
IMR32	1.38	nt 2448 to 4702, SEQ ID No. 3
		nt 1 to 84 of alternative exon,

SEQ ID No. 6

- * IMR32 1.86 has a 73 nt deletion compared to the rabbit cardiac muscle calcium channel α_1 subunit cDNA sequence.
 - $^{\circ}$ 1.16G is an α_{1c} genomic clone.
 - 2. Isolation and characterization of clones described in Example II.B.1.

a. CNS 1.30

Approximately 1 x 10⁶ recombinants of the human thalamus cDNA library No. 6 (Example I.B.6.) were screened with fragments of the rabbit skeletal muscle calcium channel α_1 cDNA described in Example II.A.2.a. The hybridization was performed using standard hybridization conditions (Example I.C.) and the filters were washed under low stringency (Example I.C.). Six positive plaques were identified, one of which was CNS 1.30. CNS 1.30 was plaque purified, restriction mapped, subcloned, and characterized by DNA sequencing. CNS 1.30 encodes α_{1c} -specific sequence nt 2199 to 3903 (SEQ ID No. 3) followed by nt 1 to 84 of one of two identified alternative α_{1c} exons (SEQ ID No. 6). 3' of SEQ ID No. 6, CNS 1.30 contains an intron and, thus, CNS 1.30 encodes a partially spliced α_{1c} transcript.

b. 1.16G

Approximately 1 x 10^6 recombinants of a λ EMBL3-based human genomic DNA library (Cat # HL1006d Clontech Corp., Palo Alto, CA) were screened using a rabbit skeletal muscle cDNA fragment (nt -78 to 1006, Example II.A.2.a.). The hybridization was performed using standard hybridization conditions (Example I.C.) and the filters were washed under low stringency (Example I.C.). Fourteen positive plaques were identified, one of which was 1.16G. Clone 1.16G was plaque purified, restriction mapped, subcloned, and portions were characterized by DNA sequencing. DNA sequencing revealed that 1.16G encodes α_{1c} -specific sequence as described in Example II.B.1.

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c. IMR32 1.66 and IMR32 1.67

Approximately 1 x 106 recombinants of IMR32 cDNA library #5 (Example I.B.5.) were screened with a 151 bp KpnI-SacI fragment of 1.16G (Example II.B.2.b.) encoding α_{1c} sequence (nt 758 to 867, SEQ ID No. 3). The hybridization was performed using standard hybridization conditions (Example I.C.). filters were then washed in 0.5 x SSPE at 65°C. positive plaques, IMR32 1.66 and IMR32 1.67 were identified. The hybridizing plaques were purified, restriction mapped, subcloned, and characterized by DNA sequencing. Two of these cDNA clones, IMR32 1.66 and 1.67, encode α_{1c} subunits as described (Example II.B.1.). In addition, IMR32 1.66 encodes a partially spliced $\alpha_{\rm ic}$ transcript marked by a GT splice donor dinucleotide beginning at the nucleotide 3' of nt 916 (SEQ ID The intron sequence within 1.66 is 101 nt long. No. 3). IMR32 1.66 encodes the α_{1c} initiation of translation, nt 1 to 3 (SEQ ID No. 3) and 132 nt of 5' untranslated sequence (SEQ ID No. 4) precede the start codon in IMR32 1.66.

d. IMR32 1.37 and IMR32 1.38

Approximately 2 x 10⁶ recombinants of IMR32 cDNA library #1 (Example I.B.1.) were screened with the CNS 1.30 cDNA fragment (Example II.B.2.a.). The hybridization was performed using low stringency hybridization conditions (Example I.C.) and the filters were washed under low stringency (Example I.C.). Four positive plaques were identified, plaque purified, restriction mapped, subcloned, and characterized by DNA sequencing. Two of the clones, IMR32 1.37 and IMR32 1.38 encode α_{1c} -specific sequences as described in Example II.B.1.

DNA sequence comparison of IMR32 1.37 and IMR32 1.38 revealed that the $\alpha_{\rm ic}$ transcript includes two exons that encode the IVS3 transmembrane domain. IMR32 1.37 has a single exon, nt 3904 to 3987 (SEQ ID No. 3) and IMR32 1.38 appears to be anomalously spliced to contain both exons juxtaposed, nt 3904 to 3987 (SEQ ID No. 3) followed by nt 1 to 84 (SEQ ID No. 6). The alternative splice of the $\alpha_{\rm ic}$ transcript to contain either of the two exons encoding the IVS3 region was confirmed by

comparing the CNS 1.30 sequence to the IMR32 1.37 sequence. CNS 1.30 contains nt 1 to 84 (SEQ ID No. 6) preceded by the identical sequence contained in IMR32 1.37 for nt 2199 to 3903 (SEQ ID No. 3). As described in Example II.B.2.a., an intron follows nt 1 to 84 (SEQ ID No. 6). Two alternative exons have been spliced adjacent to nt 3903 (SEQ ID No. 3) represented by CNS 1.30 and IMR32 1.37.

e. IMR32 1.86

IMR32 cDNA library #1 (Example I.B.1.) was screened in duplicate using oligonucleotide probes 90-9 (nt 1462 to 1491, SEO ID No. 3) and 90-12 (nt 2496 to 2520, SEQ ID No. 3). These oligonucleotide probes were chosen in order to isolate a clone that encodes the α_{1c} subunit between the 3' end of IMR32 1.67 (nt 1717, SEQ ID No. 3) and the 5' end of CNS 1.30 (nt 2199, SEQ ID No. 3). The hybridization conditions were standard hybridization conditions (Example I.C.) with the exception that the 50% deionized formamide was reduced to 20%. The filters were washed under low stringency (Example I.C.). Three positive plaques were identified one of which was IMR32 IMR32 1.86 was plaque purified, subcloned, characterized by restriction mapping and DNA sequencing. IMR32 1.86 encodes α_{ic} sequences as described in Example Characterization by DNA sequencing revealed that II.B.1. IMR32 1.86 contains a 73 nt deletion compared to the DNA encoding rabbit cardiac muscle calcium channel α_1 subunit [Mikami et al. (1989) Nature 340:230], nt 2191 to 2263. These missing nucleotides correspond to nt 2176-2248 of SEQ ID No. Because the 5'-end of CNS 1.30 overlaps the 3'-end of IMR32 1.86, some of these missing nucleotides, i.e., nt 2205-2248 of SEQ ID No. 3, are accounted for by CNS 1.30. remaining missing nucleotides of the 73 nucleotide deletion in IMR32 1.86 (i.e., nt 2176-2204 SEQ ID No. 3) were determined by nucleic acid amplification analysis of dbcAMP-induced IMR32 cell RNA. The 73 nt deletion is a frame-shift mutation and, thus, needs to be corrected. The exact human sequence through this region, (which has been determined by the DNA sequence of

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CNS 1.30 and nucleic acid amplification analysis of IMR32 cell RNA) can be inserted into IMR32 1.86 by standard methods, e.g., replacement of a restriction fragment or site-directed mutagenesis.

f. IMR32 1.157

One million recombinants of IMR32 cDNA library #4 (Example I.B.4.) were screened with an XhoI-EcoRI fragment of IMR32 1.67 encoding α_{1C} nt 50 to 774 (SEQ ID No. 3). hybridization was performed using standard hybridization conditions (Example I.C.). The filters were washed under high One of the positive plaques stringency (Example I.C.). This plaque was purified, the identified was IMR32 1.157. insert was restriction mapped and subcloned to a standard plasmid vector pGEM7Z (Promega, Madison, WI). The DNA was characterized by sequencing. IMR32 1.157 appears to encodes an alternative 5' portion of the α_{1c} sequence beginning with nt 1 to 89 (SEQ ID No. 5) and followed by nt 1 to 873 (SEQ ID Analysis of the 1.66 and 1.157 5' sequence is No. 3). described below (Example II.B.3.).

3. Characterization of the α_{1c} initiation of translation site

Portions of the sequences of IMR32 1.157 (nt 57 to 89, SEQ ID No. 5; nt 1 to 67, SEQ ID No. 3), IMR32 1.66 (nt 100 to 132, SEQ ID No. 4; nt 1 to 67, SEQ ID No. 3), were compared to the rabbit lung CaCB-receptor cDNA sequence, nt -33 to 67 [Biel et al. (1990) FEBS Lett. 269:409]. The human sequences are possible alternative 5' ends of the $\alpha_{\rm lc}$ transcript encoding the region of initiation of translation. IMR32 1.66 closely matches the CaCB receptor cDNA sequence and diverges from the CaCB receptor cDNA sequence in the 5' direction beginning at nt 122 (SEQ ID No. 4). The start codon identified in the CaCB receptor cDNA sequence is the same start codon used to describe the $\alpha_{\rm lc}$ coding sequence, nt 1 to 3 (SEQ ID No. 3).

The sequences of α_{1c} splice variants, designated α_{1c-1} and α_{1c-2} are set forth in SEQ ID NOs. 3 and 36.

C. Isolation of partial cDNA clones encoding the α_{1B} subunit and construction of a full-length clone

A human basal ganglia cDNA library was screened with the rabbit skeletal muscle α_1 subunit cDNA fragments (see Example II.A.2.a for description of fragments) under low stringency conditions. One of the hybridizing clones was used to screen an IMR32 cell cDNA library to obtain additional partial α_{1B} cDNA clones, which were in turn used to further screen an IMR32 cell cDNA library for additional partial cDNA clones. One of the partial IMR32 α_{1B} clones was used to screen a human hippocampus library to obtain a partial α_{1B} clone encoding the 3' end of the α_{1B} coding sequence. The sequence of some of the regions of the partial cDNA clones was compared to the sequence of products of nucleic acid amplification analysis of IMR32 cell RNA to determine the accuracy of the cDNA sequences.

Nucleic acid amplification analysis analysis of IMR32 cell RNA and genomic DNA using oligonucleotide primers corresponding to sequences located 5' and 3' of the STOP codon of the DNA encoding the $\alpha_{\text{\tiny 1B}}$ subunit revealed an alternatively spliced $\alpha_{\text{\tiny IB}}$ -encoding mRNA in IMR32 cells. This second mRNA product is the result of differential splicing of the α_{1B} subunit transcript to include another exon that is not present in the mRNA corresponding to the other 3' α_{1B} cDNA sequence that was initially isolated. To distinguish these splice variants of the α_{1B} subunit, the subunit encoded by a DNA sequence corresponding to the form containing the additional exon is referred to as $lpha_{ ext{BB-1}}$ (SEQ ID No. 7), whereas the subunit encoded by a DNA sequence corresponding to the form lacking the additional exon is referred to as $\alpha_{\text{1B-2}}$ (SEQ ID No. 8). sequence of α_{1B-1} diverges from that of α_{1B-2} beginning at nt 6633 (SEQ ID No. 7). Following the sequence of the additional exon in $\alpha_{\rm 1B-1}$ (nt 6633-6819; SEQ ID No. 7), the $\alpha_{\rm 1B-1}$ and $\alpha_{\rm 1B-2}$ sequences are identical (i.e., nt 6820-7362 in SEQ ID No. 7 and nt 6633-7175 in SEQ ID No. 8). SEQ ID No. 7 and No. 8 set forth 143 nt of 5' untranslated sequence (nt 1-143) as well as

202 nt of 3' untranslated sequence (nt 7161-7362, SEQ ID No. 7) of the DNA encoding $\alpha_{\rm 1B-1}$ and 321 nt of 3' untranslated sequence (nt 6855-7175, SEQ ID No. 8) of the DNA encoding $\alpha_{\rm 1B-2}$.

Nucleic acid amplification analysis analysis of the IS6 region of the α_{IB} transcript revealed what appear to be additional splice variants based on multiple fragment sizes seen on an ethidium bromide-stained agarose gel containing the products of the amplification reaction.

A full-length $\alpha_{\text{1B-1}}$ cDNA clone designated pcDNA- $\alpha_{\text{1B-1}}$ was prepared in an eight-step process as follows.

- STEP 1: The SacI restriction site of pGEM3 (Promega, Madison, WI) was destroyed by digestion at the SacI site, producing blunt ends by treatment with T4 DNA polymerase, and religation. The new vector was designated pGEMASac.
- STEP 2: Fragment 1 (HindIII/KpnI; nt 2337 to 4303 of SEQ ID No. 7) was ligated into HindIII/KpnI digested pGEM3ΔSac to produce pα1.177HK.
- STEP 3: Fragment 1 has a 2 nucleotide deletion (nt 3852 and 3853 of SEQ ID No. 7). The deletion was repaired by inserting an amplfied fragment (fragment 2) of IMR32 RNA into pα1.177HK. Thus, fragment 2 (NarI/KpnI; nt 3828 to 4303 of SEQ ID No. 7) was inserted into NarI/KpnI digested pα1.177HK replacing the NarI/KpnI portion of fragment 1 and producing pα1.177HK/PCR.
- STEP 4: Fragment 3 (KpnI/KpnI; nt 4303 to 5663 of SEQ ID No. 7) was ligated into KpnI digested pal.177HK/PCR to produce palB5'K.
- STEP 5: Fragment 4 (EcoRI/HindIII; EcoRI adaptor plus nt 1 to 2337 of SEQ ID No. 7) and fragment 5 (HindIII/XhoI fragment of pxlB5'K; nt 2337 to 5446 of SEQ ID No. 7) were ligated together into EcoRI/XhoI digested pcDNA1 (Invitrogen, San Diego, CA) to produce pxlB5'.

- STEP 6: Fragment 6 (EcoRI/EcoRI; EcoRI adapters on both ends plus nt 5749 to 7362 of SEQ ID No. 7) was ligated into EcoRI digested pBluescript II KS (Stratagene, La Jolla, CA) with the 5' end of the fragment proximal to the KpnI site in the polylinker to produce pα1.230.
- STEP 7: Fragment 7 (KpnI/XhoI; nt 4303 to 5446 of SEQ ID No. 7), and fragment 8 (XhoI/CspI; nt 5446 to 6259 of SEQ ID No. 7) were ligated into KpnI/CspI digested pα1.230 (removes nt 5749 to 6259 of SEQ ID No. 7 that was encoded in pα1.230 and maintains nt 6259 to 7362 of SEQ ID No. 7) to produce pα1B3'.
- STEP 8: Fragment 9 (SphI/XhoI; nt 4993 to 5446 of SEQ ID No. 7) and fragment 10 (XhoI/XbaI of pαlB3'; nt 5446 to 7319 of SEQ ID No. 7) were ligated into SphI/XbaI digested pαlB5' (removes nt 4993 to 5446 of SEQ ID No. 7 that were encoded in pαlB5' and maintains nt 1 to 4850 of SEQ ID No. 7) to produce pcDNAα_{1B-1}.

The resulting construct, pcDNA α_{1B-1} , contains, in pCDNA1, a full-length coding region encoding α_{1B-1} (nt 144-7362, SEQ ID No. 7), plus 5' untranslated sequence (nt 1-143, SEQ ID No. 7) and 3' untranslated sequence (nt 7161-7319, SEQ ID No. 7) under the transcriptional control of the CMV promoter.

D. Isolation of DNA encoding human calcium channel $\alpha_{1\lambda}$ subunits

1. Isolation of partial clones

DNA clones encoding portions of human calcium channel α_{1A} subunits were obtained by hybridization screening of human cerebellum cDNA libraries and nucleic acid amplification of human cerebellum RNA. Clones corresponding to the 3' end of the α_{1A} coding sequence were isolated by screening 1 x 10⁶ recombinants of a randomly primed cerebellum cDNA library (size-selected for inserts greater than 2.8 kb in length) under low stringency conditions (6X SSPE, 5X Denhart's solution, 0.2% SDS, 200 $\mu g/ml$ sonicated herring sperm DNA,

42°C) with oligonucleotide 704 containing nt 6190-6217 of the rat α_{1A} coding sequence [Starr et al. (1992) Proc. Natl. Acad. Sci. U.S.A. 88:5621-5625]. Washes were performed under low stringency conditions. Several clones that hybridized to the probe (clones $\alpha_1.251-\alpha_1.259$ and $\alpha_1.244$) were purified and characterized by restriction enzyme mapping and DNA sequence analysis. At least two of the clones, $\alpha_1.244$ and $\alpha_1.254$, contained a translation termination codon. Although clones $\alpha_1.244$ and $\alpha_1.254$ are different lengths, they both contain a sequence of nucleotides that corresponds to the extreme 3' end of the α_{1A} transcript, i.e., the two clones overlap. These two clones are identical in the region of overlap, except, clone $\alpha_1.244$ contains a sequence of 5 and a sequence of 12 nucleotides that are not present in $\alpha_1.254$.

To obtain additional α_{1A} -encoding clones, recombinants of a randomly primed cerebellum cDNA library (size-selected for inserts ranging from 1.0 to 2.8 kb in screened hybridization for oligonucleotides: oligonucleotide 701 (containing nucleotides 2288-2315 of the rat α_{1k} coding sequence), oligonucleotide 702 (containing nucleotides 3559-3585 of the rat α_{1h} coding sequence) and oligonucleotide 703 (containing nucleotides 4798-4827 of the rat α_{1A} coding sequence). Hybridization and washes were performed using the same conditions as used for the first screening with oligonucleotide 704, except that washes were conducted at 45°C . Twenty clones (clones $\alpha1.269$ α1.288) hybridized to the probe. Several clones were plaquepurified and characterized by restriction enzyme mapping and One clone, α 1.279, contained a DNA sequence analysis. sequence of about 170 nucleotides that is not present in other clones corresponding to the same region of the This region may be present in other sequence. None of the clones contained a translation variants. intiation codon.

To obtain clones corresponding to the 5' end of the human α_{lA} coding sequence, another cerebellum cDNA library was

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prepared using oligonucleotide 720 (containing nucleotides 2485-2510 of SEQ ID No. 22) to specifically prime first-strand The library (8 x 10⁵ recombinants) was cDNA synthesis. screened for hybridization to three oligonucleotides: oligonucleotide 701, oligonucleotide 726 (containing nucleotides 2333-2360 of the rat α_{1A} coding sequence) and oligonucleotide 700 (containing nucleotides 767-796 of the rat α_{1h} coding sequence) under low stringency hybridization and washing conditions. Approximately 50 plaques hybridized to Hybridizing clones $\alpha 1.381-\alpha 1.390$ were plaquethe probe. purified and characterized by restriction enzyme maping and DNA sequence analysis. At least one of the clones, α 1.381, contained a translation initiation codon.

Alignment of the sequences of the purified clones revealed that the sequences overlapped to comprise the entire $\alpha_{1\lambda}$ coding sequence. However, not all the overlapping sequences of partial clones contained convenient enzyme restriction sites for use in ligating partial clones to construct a full-length α_{1A} coding sequence. To obtain DNA fragments containing convenient restriction enzyme sites that could be used in constructing a full-length α_{1A} DNA, cDNA was synthesized from RNA isolated from human cerebellum tissue and subjected to nucleic acid amplification. The oligonucleotides used as primers corresponded to human α_{iA} coding sequence located 5' and 3' of selected restriction enzyme sites. Thus, in the first amplification reaction, oligonucleotides 753 (containing nucleotides 2368-2391 of SEQ ID No. 22) and 728 (containing nucleotides 3179-3202 of SEQ ID No. 22) were used as the primer pair. To provide a sufficient amount of the desired DNA fragment, the product of this amplification was reamplified using oligonucleotides 753 and 754 (containing nucleotides 3112-3135 of SEQ ID No. 22 as the primer pair. The resulting product was 768 bp in length. In the second amplification reaction, oligonucleotides 719 (containing nucleotides 4950-4975 of SEQ ID No. 22 and 752 (containing nucleotides 5647-5670 of SEQ ID No. 22) were used as the primer pair. To provide a sufficient amount of the desired second DNA fragment, the product of this amplification was reamplified using oligonucleotides 756 (containing nucleotides 5112-5135 of SEQ ID No. 22) and 752 as the primer pair. The resulting product was 559 bp in length.

2. Construction of full-Length α_{1a} coding sequences Portions of clone $\alpha 1.381$, the 768-bp nucleic acid amplification product, clone $\alpha 1.278$, the 559-bp nucleic acid amplification product, and clone $\alpha 1.244$ were ligated at convenient restriction sites to generate a full-length α_{1a} coding sequence referred to as α_{1a-1} .

Comparison of the results of sequence analysis of clones α 1.244 and α 1.254 indicated that the primary transcript of the α_{1A} subunit gene is alternatively spliced to yield at least two variant mRNAs encoding different forms of the α_{12} subunit. One form, α_{1A-1} , is encoded by the sequence shown in SEQ ID No. 22. The sequence encoding a second form, α_{1A-2} , differs from the α_{1A-1} 1-encoding sequence at the 3' end in that it lacks a 5-nt sequence found in clone $\alpha 1.244$ (nucleotides 7035-7039 of SEQ ID No. 22). This deletion shifts the reading frame and introduces a translation termination codon resulting in an α_{1A-2} coding sequence that encodes a shorter $\alpha_{\mathtt{l}\mathtt{A}}$ subunit than that encoded by the α_{1k-1} splice variant. Consequently, a portion of the 3' end of the α_{1A-1} coding sequence is actually 3' untranslated sequence in the α_{1A-2} DNA. The complete sequence of α_{1A-2} , which can be constructed by ligating portions of clone α1.381, the 768-bp nucleic acid amplification product, clone \$\alpha 1.278, the 559-bp nucleic acid amplification product and clone α 1.254, is set forth in SEQ ID No. 23.

E. Isolation of DNA Encoding the α_{1E} Subunit

DNA encoding α_{lE} subunits of the human calcium channel were isolated from human hippocampus libraries. The selected clones sequenced. DNA sequence analysis of DNA clones encoding the α_{lE} subunit indicated that at least two alternatively spliced forms of the same α_{lE} subunit primary transcript are expressed. One form has the sequence set forth

in SEQ ID No. 24 and was designated $\alpha_{\rm 1E-1}$ and the other was designated $\alpha_{\rm 1E-3}$, which has the sequence obtained by inserting a 57 base pair fragment between nucleotides 2405 and 2406 of SEQ ID No. 24. The resulting sequence is set forth in SEQ ID No. 25.

The subunit designated $\alpha_{\text{1E-1}}$ has a calculated molecular weight of 254,836 and the subunit designated $\alpha_{\text{1E-3}}$ has a calculated molecular weight of 257,348. $\alpha_{\text{1E-3}}$ has a 19 amino acid insertion (encoded by SEQ ID No. 25) relative to $\alpha_{\text{1E-1}}$ in the region that appears to be the cytoplasmic loop between transmembrane domains IIS6 and IIIS1.

EXAMPLE III: ISOLATION OF cDNA CLONES ENCODING THE HUMAN NEURONAL CALCIUM CHANNEL β_1 subunit

A. Isolation of partial cDNA clones encoding the β subunit and construction of a full-length clone encoding the β_1 subunit

A human hippocampus cDNA library was screened with the rabbit skeletal muscle calcium channel β_1 subunit cDNA fragment (nt 441 to 1379) [for isolation and sequence of the rabbit skeletal muscle calcium channel β_1 subunit cDNA, see U.S. Patent Application Serial NO. 482,384 or Ruth et al. (1989) Science 245:1115] using standard hybridization conditions (Example I.C.). A portion of one of the hybridizing clones was used to rescreen the hippocampus library to obtain additional cDNA clones. The cDNA inserts of hybridizing clones were characterized by restriction mapping and DNA sequencing and compared to the rabbit skeletal muscle calcium channel β_1 subunit cDNA sequence.

Portions of the partial β_1 subunit cDNA clones were ligated to generate a full-length clone encoding the entire β_1 subunit. SEQ ID No. 9 shows the β_1 subunit coding sequence (nt 1-1434) as well as a portion of the 3' untranslated sequence (nt 1435-1546). The deduced amino acid sequence is also provided in SEQ ID No. 9. In order to perform expression experiments, full-length β_1 subunit cDNA clones were constructed as follows.

Step 1: DNA fragment 1 (~800 bp of 5' untranslated sequence plus nt 1-277 of SEQ ID No. 9) was ligated to DNA fragment 2 (nt 277-1546 of SEQ ID No. 9 plus 448 bp of intron sequence) and cloned into pGEM7Z. The resulting plasmid, p β 1-1.18, contained a full-length β_1 subunit clone that included a 448-bp intron.

Step 2: To replace the 5' untranslated sequence of $p\beta1$ -1.18 with a ribosome binding site, a double-stranded adapter was synthesized that contains an EcoRI site, sequence encoding a ribosome binding site (5'-ACCACC-3') and nt 1-25 of SEQ ID No. 9. The adapter was ligated to SmaI-digested $p\beta1$ -1.18, and the products of the ligation reaction were digested with EcoRI.

Step 3: The EcoRI fragment from step 2 containing the EcoRI adapter, efficient ribosome binding site and nt 1-1546 of SEQ ID No. 9 plus intron sequence was cloned into a plasmid vector and designated p β 1-1.18RBS. The EcoRI fragment of p β 1-1.18RBS was subcloned into EcoRI-digested pcDNA1 with the initiation codon proximal to CMV promoter to form pHBCaCH β_{1a} RBS(A).

Step 4: To generate a full-length clone encoding the β_1 subunit lacking intron sequence, DNA fragment 3 (nt 69-1146 of SEQ ID No. 9 plus 448 bp of intron sequence followed by nt 1147-1546 of SEQ ID No. 9), was subjected to site-directed mutagenesis to delete the intron sequence, thereby yielding p β 1(-). The EcoRI-XhoI fragment of p β 1-1.18RBS (containing of the ribosome binding site and nt 1-277 of SEQ ID No. 9) was ligated to the XhoI-EcoRI fragment of p β 1(-) (containing of nt 277-1546 of SEQ ID No. 9) and cloned into pcDNA1 with the initiation of translation proximal to the CMV promoter. The resulting expression plasmid was designated pHBCaCH β_{1b} RBS(A).

B. Splice Variant β_{1-3}

DNA sequence analysis of the DNA clones encoding the β_1 subunit indicated that in the CNS at least two alternatively spliced forms of the same human β_1 subunit primary transcript are expressed. One form is represented by the sequence shown

in SEQ ID No. 9 and is referred to as β_{1-2} . The sequences of β_{1-2} and the alternative form, β_{1-3} , diverge at nt 1334 (SEQ ID No. 9). The complete β_{1-3} sequence (nt 1-1851), including 3' untranslated sequence (nt 1795-1851), is set forth in SEQ ID No. 10.

EXAMPLE IV: ISOLATION OF cDNA CLONES ENCODING THE HUMAN NEURONAL CALCIUM CHANNEL α_2 -subunit

A. Isolation of cDNA clones

The complete human neuronal α_2 coding sequence (nt 35-3310) plus a portion of the 5' untranslated sequence (nt 1 to 34) as well as a portion of the 3' untranslated sequence (nt 3311-3600) is set forth in SEO ID No. 11.

To isolate DNA encoding the human neuronal α, subunit, human α_2 genomic clones first were isolated by probing human genomic Southern blots using a rabbit skeletal muscle calcium channel α_2 subunit cDNA fragment [nt 43 to 272, Ellis et al. (1988) Science 240:1661]. Human genomic DNA was digested with EcoRI, electrophoresed, blotted, and probed with the rabbit skeletal muscle probe using standard hybridization conditions (Example I.C.) and low stringency washing conditions (Example I.C.). Two restriction fragments were identified, 3.5 kb and These EcoRI restriction fragments were cloned by 3.0 kb. preparing a Agt11 library containing human genomic EcoRI fragments ranging from 2.2 kb to 4.3 kb. The library was screened as described above using the rabbit α_2 probe, hybridizing clones were isolated and characterized by DNA sequencing. HGCaCHα2.20 contained the 3.5 kb fragment and $HGCaCH\alpha 2.9$ contained the 3.0 kb fragment.

Restriction mapping and DNA sequencing revealed that $HGCaCH\alpha 2.20$ contains an 82 bp exon (nt 130 to 211 of the human α_2 coding sequence, SEQ ID No. 11) on a 650 bp PstI-XbaI restriction fragment and that $HGCaCH\alpha 2.9$ contains 105 bp of an exon (nt 212 to 316 of the coding sequence, SEQ ID No. 11) on a 750 bp XbaI-BgIII restriction fragment. These restriction fragments were used to screen the human basal ganglia cDNA library (Example II.C.2.a.). $HBCaCH\alpha 2.1$ was isolated (nt 29

to 1163, SEQ ID No. 11) and used to screen a human brain stem cDNA library (ATCC Accession No. 37432) obtained from the American Type Culture Collection, 12301 Parklawn Drive, Rockville, MD. 20852. Two clones were isolated, HBCaCH α 2.5 (nt 1 to 1162, SEQ ID No. 11) and HBCaCH α 2.8 (nt 714 to 1562, SEQ ID No. 11, followed by 1600 nt of intervening sequence). A 2400 bp fragment of HBCaCH α 2.8 (beginning at nt 759 of SEQ ID No. 11 and ending at a SmaI site in the intron) was used to rescreen the brain stem library and to isolate HBCaCH α 2.11 (nt 879 to 3600, SEQ ID No. 11). Clones HBCaCH α 2.5 and HBCaCH α 2.11 overlap to encode an entire human brain α 2 protein.

B. Construction of pHBCaCHα2A

To construct pHBCaCHα2A containing DNA encoding a fulllength human calcium channel α_2 subunit, an (EcoRI)-PvuII fragment of HBCaCHα2.5 (nt 1 to 1061, SEQ ID No. 11, EcoRI adapter, PvuII partial digest) and a PvuII-PstI fragment of HBCaCHα2.11 (nt 1061 to 2424 SEQ ID No. 11; PvuII partial were ligated into EcoRI-PstI-digested (Stratagene, La Jolla, CA). Subsequently, an (EcoRI)-PstI fragment (nt 1 to 2424 SEQ ID No. 11) was isolated and ligated to a PstI-(EcoRI) fragment (nt 2424 to 3600 SEQ ID No. 11) of HBCaCHα2.11 in EcoRI-digested pIBI24 to produce DNA, HBCaCHα2, encoding a full-length human brain α_2 subunit. The 3600 bp EcoRI insert of HBCaCHα2 (nt 1 to 3600, SEQ ID No. 11) was subcloned into pcDNA1 (pHBCaCHα2A) with the methionine initiating codon proximal to the CMV promoter. The 3600 bp EcoRI insert of HBCaCHα2 was also subcloned into pSV2dHFR [Subramani et al. (1981). Mol. Cell. Biol. 1:854-864] which SV40 early promoter, the mouse reductase (dhfr) gene, SV40 polyadenylation and splice sites and sequences required for maintenance of the vector in bacteria.

EXAMPLE V. DIFFERENTIAL PROCESSING OF THE HUMAN eta_1 TRANSCRIPT AND THE HUMAN $lpha_2$ TRANSCRIPT

A. Differential processing of the β_1 transcript

Nucleic acid amplification analysis of the human β_1 transcript present in skeletal muscle, aorta, hippocampus and basal ganglia, and HEK 293 cells revealed differential processing of the region corresponding to nt 615-781 of SEQ ID No. 9 in each of the tissues. Four different sequences that result in five different processed β_1 transcripts through this region were identified. The β_1 transcripts from the different tissues contained different combinations of the four sequences, except for one of the β_1 transcripts expressed in HEK 293 cells (β_{1-5}) which lacked all four sequences.

None of the β_1 transcripts contained each of the four sequences; however, for ease of reference, all four sequences are set forth end-to-end as a single long sequence in SEQ ID No. 12. The four sequences that are differentially processed are sequence 1 (nt 14-34 in SEQ ID No. 12), sequence 2 (nt 35-55 in SEQ ID No. 12), sequence 3 (nt 56-190 in SEQ ID No. 12) and sequence 4 (nt 191-271 in SEQ ID No. 12). The forms of the β_1 transcript that have been identified include: form that lacks sequence 1 called $eta_{\scriptscriptstyle 1-1}$ (expressed in skeletal muscle), (2) a form that lacks sequences 2 and 3 called β_{1-2} (expressed in CNS), (3) a form that lacks sequences 1, 2 and 3 called $eta_{ ext{1-4}}$ (expressed in aorta and HEK cells) and (4) a form that lacks sequences 1-4 called $\beta_{\text{1-5}}$ (expressed in HEK cells). Additionally, the β_{1-4} and β_{1-5} contain a guanine nucleotide (nt 13 in SEQ ID No. 12) that is absent in the β_{1-1} and β_{1-2} forms. The sequences of eta_1 splice variants are set forth in SEQ ID Nos. 9, 10 and 33-35.

B. Differential processing of transcripts encoding the α_2 subunit.

The complete human neuronal α_2 coding sequence (nt 35-3307) plus a portion of the 5' untranslated sequence (nt 1 to 34) as well as a portion of the 3' untranslated sequence (nt 3308-3600) is set forth as SEQ ID No. 11.

Nucleic acid amplification analysis of the human α_2 transcript present in skeletal muscle, aorta, and CNS revealed differential processing of the region corresponding to nt 1595-1942 of SEQ ID No. 11 in each of the tissues.

The analysis indicated that the primary transcript of the genomic DNA that includes the nucleotides corresponding to nt 1595-1942 also includes an additional sequence (SEQ ID 5'CCTATTGGTGTAGGTATACCAACAATTAATTT AAGAAAAAGGAGACCCAATATCCAG 3') inserted between nt 1624 and 1625 of SEQ ID No. 11. Five alternatively spliced variant transcripts that differ in the presence or absence of one to three different portions of the region of the primary transcript that includes the region of nt 1595-1942 of SEQ ID No. 11 plus SEQ ID No. 13 inserted between nt 1624 and 1625 have been identified. The five α_2 -encoding transcripts from the different tissues include different combinations of the three sequences, except for one of the α_2 transcripts expressed in aorta which lacks all three sequences. the α_2 transcripts contained each of the three sequences. sequences of the three regions that are differentially processed are sequence 1 (SEQ ID No. 13), sequence 2 (5' AACCCCAAATCTCAG 3', which is nt 1625-1639 of SEQ ID No. 11), and sequence 3 (5' CAAAAAAGGGCAAAATGAAGG 3', which is nt 1908-1928 of SEQ ID No. 11). The five α_2 forms identified are (1) a form that lacks sequence 3 called α_{2a} (expressed in skeletal muscle), (2) a form that lacks sequence 1 called α_{2b} (expressed in CNS), (3) a form that lacks sequences 1 and 2 called α_{2c} expressed in aorta), (4) a form that sequences 1, 2 and 3 called α_{2d} (expressed in aorta) and (5) a form that lacks sequences 1 and 3 called $\alpha_{\rm 2e}$ (expressed in aorta).

The sequences of α_{2a} - α_{2e} are set forth in SEQ. ID Nos. 29 - 32, respectively.

EXAMPLE VI: ISOLATION OF DNA ENCODING A CALCIUM CHANNEL γ SUBUNIT FROM A HUMAN BRAIN CDNA LIBRARY

A. Isolation of DNA encoding the γ subunit

Approximately 1 x 10° recombinants from a \(\lambda gtll-based \) human hippocampus cDNA library (Clontech catalog #HL1088b, Palo Alto, CA) were screened by hybridization to a 484 bp sequence of the rabbit skeletal muscle calcium channel γ subunit cDNA (nucleotides 621-626 of the coding sequence plus 438 nucleotides of 3'-untranslated sequence) contained in vector γ J10 [Jay, S. et al. (1990). Science 248:490-492]. Hybridization was performed using moderate conditions (20% deionized formamide, 5x Denhardt's, 6 x SSPE, 0.2% SDS, 20 μ g/ml herring sperm DNA, 42°C) and the filters were washed under low stringency (see Example I.C.). A plaque that hybridized to this probe was purified and insert DNA was subcloned into pGEM7Z. This cDNA insert was designated $\gamma 1.4$.

B. Characterization of γ 1.4

 $\gamma 1.4$ was confirmed by DNA hybridization and characterized by DNA sequencing. The 1500 bp SstI fragment of $\gamma 1.4$ hybridized to the rabbit skeletal muscle calcium channel γ subunit cDNA $\gamma J10$ on a Southern blot. SEQ analysis of this fragment revealed that it contains of approximately 500 nt of human DNA sequence and ~1000 nt of $\lambda gtll$ sequence (included due to apparent destruction of one of the EcoRI cloning sites in $\lambda gtll$). The human DNA sequence contains of 129 nt of coding sequence followed immediately by a translational STOP codon and 3' untranslated sequence (SEQ ID No. 14).

To isolate the remaining 5' sequence of the human γ subunit cDNA, human CNS cDNA libraries and/or preparations of mRNA from human CNS tissues can first be assayed by nucleic acid amplification analysis methods using oligonucleotide primers based on the γ cDNA-specific sequence of $\gamma 1.4$. Additional human neuronal γ subunit-encoding DNA can be isolated from cDNA libraries that, based on the results of the nucleic acid amplification analysis assay, contain γ -specific

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Alternatively, cDNA libraries can amplifiable cDNA. constructed from mRNA preparations that, based on the results of the nucleic acid amplification analysis assays, contain γ specific amplifiable transcripts. Such libraries constructed by standard methods using oligo dT to prime firststrand cDNA synthesis from poly A+ RNA (see Example I.B.). Alternatively, first-strand cDNA can be specified by priming first-strand CDNA synthesis with а γ cDNA-specific oligonucleotide based on the human DNA sequence in $\gamma 1.4$. cDNA library can then be constructed based on this firststrand synthesis and screened with the γ -specific portion of $\gamma 1.4.$

EXAMPLE VII: ISOLATION OF cDNA CLONES ENCODING THE HUMAN NEURONAL Ca CHANNEL β_2 SUBUNIT

Isolation of DNA Encoding human calcium channel $oldsymbol{eta}_2$ subunits

Sequencing of clones isolated as described in Example III revealed a clone encoding a human neuronal calcium channel β_2 (designated see, SEQ ID No. $oldsymbol{eta_{2D}}$ oligonucleotide based on the 5' end of this clone was used to prime a human hippocampus cDNA library. The library was screened with this β_2 clone under conditions of low to medium stringency (final wash 0.5 X SSPE, 50° C). Several hybridizing clones were isolated and sequenced. Among these clones were those that encode β_{2C} , β_{2D} and β_{2E} . For example, the sequence of β_{2c} is set forth in SEQ ID NO. 37, and the sequeence of β_{2r} is set forth in SEQ ID No. 38.

A randomly primed hippocampus library was then screened using a combination of the clone encoding β_{2D} and a portion of the β_3 clone deposited under ATCC Accession No. 69048. Multiple hybridizing clones were isolated. Among these were clones designated β_{101} , β_{102} and β_{104} . β_{101} appears to encodes the 5' end of a splice variant of β_2 , designated β_{2E} . β_{102} and β_{104} encode portions of the 3' end of β_2 .

It appears that the β_2 splice variants include nucleotides 182-2294 of SEQ ID No. 26 and differ only between

the start codon and nucleotides that correspond to 212 of SEQ. ID No. 26.

EXAMPLE VIII: ISOLATION OF cDNA CLONES ENCODING HUMAN CALCIUM CHANNEL β_4 and β_5 SUBUNITS

A. Isolation of cDNA Clones Encoding a Human β_4 Subunit A clone containing a translation initiation codon and

approximately 60% of the eta_4 coding sequence was obtained from a human cerebellum cDNA library (see nucleotides 1-894 of Sequence ID No. 27). To obtain DNA encoding the remaining 3' portion of the β_4 coding sequence, a human cerebellum cDNA library was screened for hybridization a nucleic acid amplification product under high stringency hybridization and wash conditions. Hybridizing clones are purified and characterized by restriction enzyme mapping and DNA sequence analysis to identify those that contain sequence corresponding to the 3' end of the β_4 subunit coding sequence and a Selected clones are ligated to the clone termination codon. containing the 5' half of the β_4 coding sequence at convenient restriction sites to generate a full-length cDNA encoding a β_4 subunit. The sequence of a full-length β_4 clone is set forth in SEQ ID No. 27; the amino acid sequence is set forth in SEQ ID No. 28.

B. Isolation of cDNA Clones Encoding a Human $\beta 3$ Subunit

Sequencing of clones isolated as described in Example III also revealed a clone encoding a human neuronal calcium channel β_3 subunit. This clone has been deposited as plasmid $\beta_{1.42}$ (ATCC Accession No. 69048).

To isolate a full-length cDNA clone encoding a complete β_3 subunit, a human hippocampus cDNA library (Stratagene, La Jolla, CA) was screened for hybridization to a 5' EcoRI-PstI fragment of the cDNA encoding β_{1-2} using lower stringency hybridization conditions (20% deionized formamide, 200 μ g/ml sonicated herring sperm DNA, 5% SSPE, 5% Denhardt's solution, 42° C) and wash conditions. One of the hybridizing clones contained both translation initiation and termination codons

and encodes a complete β_3 subunit designated β_{3-1} (Sequence ID No. 19). In vitro transcripts of the cDNA were prepared and injected into Xenopus oocytes along with transcripts of the α_{1B-1} and α_{2b} cDNAs using methods similiar to those described in Example IX.D. Two-electrode voltage clamp recordings of the oocytes revealed significant voltage-dependent inward Ba²⁺ currents.

An additional β_3 subunit-encoding clone, designated β_{3-2} , was obtained by screening a human cerebellum cDNA library for hybridization to the nucleic acid amplification product referred to in Example VIII.A. under lower stringency (20% deionized formamide, 200 μ g/ml sonicated herring sperm DNA, 5X SSPE, 5X Denhardt's solution, 42° C) hybridization and wash conditions. The 5' ends of this clone (Sequence ID No. 20, β_3 . 2) and the first β_3 subunit, designated β_{3-1} , (Sequence ID No. 19) differ at their 5' ends and are splice variants of the β_3 gene.

EXAMPLE IX: RECOMBINANT EXPRESSION OF HUMAN NEURONAL CALCIUM CHANNEL SUBUNIT-ENCODING CDNA AND RNATRANSCRIPTS IN MAMMALIAN CELLS

A. Recombinant Expression of the Human Neuronal Calcium Channel α_2 subunit cDNA in DG44 Cells

1. Stable transfection of DG44 cells

DG44 cells [dhfr Chinese hamster ovary cells; see, e.g., Urlaub, G. et al. (1986) Som. Cell Molec. Genet. 12:555-566] obtained from Lawrence Chasin at Columbia University were stably transfected by CaPO, precipitation methods [Wigler et al. (1979) Proc. Natl. Acad. Sci. USA 76:1373-1376] with pSV2dhfr vector containing the human neuronal calcium channel for polycistronic (see Example IV) α_2 -subunit CDNA expression/selection in transfected cells. Transfectants were grown on 10% DMEM medium without hypoxanthine or thymidine in order to select cells that had incorporated the expression Twelve transfectant cell lines were established as indicated by their ability to survive on this medium.

2. Analysis of α_2 subunit cDNA expression in transfected DG44 cells

Total RNA was extracted according to the method of Birnboim [(1988) Nuc. Acids Res. 16:1487-1497] from four of the DG44 cell lines that had been stably transfected with pSV2dhfr containing the human neuronal calcium channel α_2 subunit cDNA. RNA (~15 μ g per lane) was separated on a 1% agarose formaldehyde gel, transferred to nitrocellulose and hybridized to the random-primed human neuronal calcium channel α_2 cDNA (hybridization: 50% formamide, 5 x SSPE, Denhardt's, 42° C.; wash :0.2 x SSPE, 0.1% SDS, 65° C.). Northern blot analysis of total RNA from four of the DG44 cell lines that had been stably transfected with pSV2dhfr containing the human neuronal calcium channel α_2 subunit cDNA revealed that one of the four cell lines contained hybridizing mRNA the size expected for the transcript of the $\alpha_{\scriptscriptstyle 2}$ subunit cDNA (5000 nt based on the size of the cDNA) when grown in the presence of 10 mM sodium butyrate for two days. nonspecifically induces transcription and is often used for inducing the SV40 early promoter [Gorman, C. and Howard, B. (1983) Nucleic Acids Res. 11:1631]. This cell line, $44\alpha_2-9$, also produced mRNA species smaller (several species) and larger (6800 nt) than the size expected for the transcript of the α_2 cDNA (5000 nt) that hybridized to the α_2 cDNA-based The 5000- and 6800-nt transcripts produced by this transfectant should contain the entire α_2 subunit coding sequence and therefore should yield a full-length α_2 subunit protein. A weakly hybridizing 8000-nucleotide transcript was present in untransfected and transfected DG44 Apparently, DG44 cells transcribe a calcium channel α , subunit or similar gene at low levels. The level of expression of this endogenous α_2 subunit transcript did not appear to be affected by exposing the cells to butyrate before isolation of RNA for northern analysis.

Total protein was extracted from three of the DG44 cell lines that had been stably transfected with pSV2dhfr

containing the human neuronal calcium channel α_2 subunit cDNA. Approximately 10^7 cells were sonicated in 300 μl of a solution containing 50 mM HEPES, 1 mM EDTA, 1 mM PMSF. An equal volume of 2x loading dye [Laemmli, U.K. (1970). Nature 227:680] was added to the samples and the protein was subjected to electrophoresis on an 8% polyacrylamide gel electrotransferred to nitrocellulose. The nitrocellulose was incubated with polyclonal guinea pig antisera (1:200 dilution) directed against the rabbit skeletal muscle calcium channel α_2 subunit (obtained from K. Campbell, University of Iowa) followed by incubation with [125I]-protein A. The blot was exposed to X-ray film at -70° C. Reduced samples of protein from the transfected cells as well as from untransfected DG44 cells contained immunoreactive protein of the size expected for the α_2 subunit of the human neuronal calcium channel (130-150 kDa). The level of this immunoreactive protein was higher in $44\alpha_2$ -9 cells that had been grown in the presence of 10 mM sodium butyrate than in $44\alpha_2$ -9 cells that were grown in the absence of sodium butyrate. These data correlate well with those obtained in northern analyses of total RNA from $44\alpha_2$ -9 and untransfected DG44 cells. Cell line $44\alpha_2$ -9 also produced a 110 kD immunoreactive protein that may be either a product of proteolytic degradation of the full-length α_2 subunit or a product of translation of one of the shorter (<5000 nt) mRNAs produced in this cell line that hybridized to the α_2 subunit cDNA probe.

B. Expression of DNA encoding human neuronal calcium channel α_1 , α_2 and β_1 subunits in HEK cells

Human embryonic kidney cells (HEK 293 cells) were transiently and stably transfected with human neuronal DNA encoding calcium channel subunits. Individual transfectants were analyzed electrophysiologically for the presence of voltage-activated barium currents and functional recombinant voltage-dependent calcium channels were.

Transfection of HEK 293 cells

Separate expression vectors containing DNA encoding human neuronal calcium channel α_{1D} , α_2 and β_1 subunits, plasmids pVDCCIII(A), pHBCaCH α_2 A, and pHBCaCH β_{1a} RBS(A), respectively, were constructed as described in Examples II.A.3, IV.B. and III.B.3., respectively. These three vectors were used to transiently co-transfect HEK 293 cells. For stable transfection of HEK 293 cells, vector pHBCaCH β_{1b} RBS(A) (Example III.B.3.) was used in place of pHBCaCH β_{1a} RBS(A) to introduce the DNA encoding the β_1 subunit into the cells along with pVDCCIII(A) and pHBCaCH α_2 A.

a. Transient transfection

Expression vectors pVDCCIII(A), pHBCaCHα,A pHBCaCH β_{1a} RBS (A) were used in two sets of transient transfections of HEK 293 cells (ATCC Accession No. CRL1573). In one transfection procedure, HEK 293 cells were transiently cotransfected with the α_1 subunit cDNA expression plasmid, the $lpha_2$ subunit cDNA expression plasmid, the eta_1 subunit cDNA expression plasmid plasmid and pCMVβgal Laboratories, Palo Alto, CA). Plasmid pCMVetagal contains the lacZ gene (encoding E. coli β -galactosidase) fused to the cytomegalovirus (CMV) promoter and was included in this transfection as a marker gene for monitoring the efficiency of transfection. In the other transfection procedure, HEK 293 cells were transiently co-transfected with the α_1 subunit cDNA expression plasmid pVDCCIII(A) and pCMV β gal. transfections, $2-4 \times 10^6$ HEK 293 cells in a 10-cm tissue culture plate were transiently co-transfected with 5 μg of each of the plasmids included in the experiment according to standard CaPO, precipitation transfection procedures (Wigler et al. (1979) Proc. Natl. Acad. Sci. USA 76:1373-1376). transfectants were analyzed for β -galactosidase expression by direct staining of the product of a reaction involving β galactosidase and the X-gal substrate [Jones, J.R. (1986) EMBO 5:3133-3142] and by measurement of β -galactosidase activity [Miller, J.H. (1972) Experiments in Molecular Genetics, pp.

352-355, Cold Spring Harbor Press]. To evaluate subunit cDNA expression in these transfectants, the cells were analyzed for subunit transcript production (northern analysis), subunit protein production (immunoblot analysis of cell lysates) and functional calcium channel expression (electrophysiological analysis).

b. Stable transfection

HEK 293 cells were transfected using the calcium phosphate transfection procedure [Current Protocols in Molecular Biology, Vol. 1, Wiley Inter-Science, Supplement 14, Unit 9.1.1-9.1.9 (1990)]. Ten-cm plates, each containing one-to-two million HEK 293 cells, were transfected with 1 ml of DNA/calcium phosphate precipitate containing 5 μ g pVDCCIII(A), 5 μ g pHBCaCH α_2 A, 5 μ g pHBCaCH β_{1b} RBS(A), 5 μ g pCMVBgal and 1 μ g pSV2neo (as a selectable marker). After 10-20 days of growth in media containing 500 μ g G418, colonies had formed and were isolated using cloning cylinders.

2. Analysis of HEK 293 cells transiently transfected with DNA encoding human neuronal calcium channel subunits

a. Analysis of β -galactosidase expression

Transient transfectants were assayed for β -galactosidase expression by β -galactosidase activity assays (Miller, J.H., (1972) Experiments in Molecular Genetics, pp. 352-355, Cold Spring Harbor Press) of cell lysates (prepared as described in Example VII.A.2) and staining of fixed cells (Jones, J.R. (1986) EMBO 5:3133-3142). The results of these assays indicated that approximately 30% of the HEK 293 cells had been transfected.

b. Northern analysis

PolyA+ RNA was isolated using the Invitrogen Fast Trak Kit (InVitrogen, San Diego, CA) from HEK 293 cells transiently transfected with DNA encoding each of the α_1 , α_2 and β_1 subunits and the lacZ gene or the α_1 subunit and the lacZ gene. The RNA was subjected to electrophoresis on an agarose gel and transferred to nitrocellulose. The nitrocellulose was then hybridized with one or more of the following radiolabeled

probes: the lacZ gene, human neuronal calcium channel $lpha_{ exttt{1D}}$ subunit-encoding cDNA, human neuronal calcium channel α_2 subunit-encoding cDNA or human neuronal calcium channel eta_1 subunit-encoding cDNA. Two transcripts that hybridized with the $\alpha_{\scriptscriptstyle 1}$ subunit-encoding cDNA were detected in HEK 293 cells transfected with the DNA encoding the $\alpha_{\scriptscriptstyle 1},~\alpha_{\scriptscriptstyle 2},~$ and $\beta_{\scriptscriptstyle 1}$ subunits and the lacZ gene as well as in HEK 293 cells transfected with the $lpha_1$ subunit cDNA and the lacZ gene. One mRNA species was the size expected for the transcript of the $lpha_1$ subunit cDNA (8000 nucleotides). The second RNA species was smaller (4000 nucleotides) than the size expected for this transcript. RNA of the size expected for the transcript of the lacZ gene was detected in cells transfected with the $\alpha_{\scriptscriptstyle 1}$, $\alpha_{\scriptscriptstyle 2}$ and $\beta_{\scriptscriptstyle 1}$ subunitencoding cDNA and the lacZ gene and in cells transfected with the $\alpha_{\scriptscriptstyle 1}$ subunit cDNA and the lacZ gene by hybridization to the lacZ gene sequence.

RNA from cells transfected with the α_1 , α_2 and β_1 subunit-encoding cDNA and the lacZ gene was also hybridized with the α_2 and β_1 subunit cDNA probes. Two mRNA species hybridized to the α_2 subunit cDNA probe. One species was the size expected for the transcript of the α_2 subunit cDNA (4000 nucleotides). The other species was larger (6000 nucleotides) than the expected size of this transcript. Multiple RNA species in the cells co-transfected with α_1 , α_2 and β_1 subunit-encoding cDNA and the lacZ gene hybridized to the β_1 subunit cDNA probe. Multiple β subunit transcripts of varying sizes were produced since the β subunit cDNA expression vector contains two potential polyA addition sites.

c. Electrophysiological analysis

Individual transiently transfected HEK 293 cells were assayed for the presence of voltage-dependent barium currents using the whole-cell variant of the patch clamp technique [Hamill et al. (1981). Pflugers Arch. 391:85-100]. HEK 293 cells transiently transfected with pCMV β gal only were assayed for barium currents as a negative control in these experiments. The cells were placed in a bathing solution that

contained barium ions to serve as the current carrier. Choline chloride, instead of NaCl or KCl, was used as the major salt component of the bath solution to eliminate currents through sodium and potassium channels. The bathing solution contained 1 $mM \ MgCl_2$ and was buffered at pH 7.3 with 10 mM HEPES (pH adjusted with sodium or tetraethylammonium Patch pipettes were filled with a solution hvdroxide). containing 135 mM CsCl, 1 mM MgCl₂, 10 mM glucose, 10 mM EGTA, and 10 mM HEPES (pH adjusted to 7.3 with tetraethylammonium hydroxide). Cesium and tetraethylammonium ions block most types of potassium channels. Pipettes were coated with Sylgard (Dow-Corning, Midland, MI) resistances of 1-4 megohm. Currents were measured through a 500 megohm headstage resistor with the Axopatch IC (Axon Instruments, Foster City, CA) amplifier, interfaced with a Labmaster (Scientific Solutions, Solon, OH) data acquisition board in an IBM-compatible PC. PClamp (Axon Instruments) was used to generate voltage commands and acquire data. Data were analyzed with pClamp or Quattro Professional (Borland International, Scotts Valley, CA) programs.

To apply drugs, "puffer" pipettes positioned within several micrometers of the cell under study were used to apply solutions by pressure application. The drugs used for pharmacological characterization were dissolved in a solution identical to the bathing solution. Samples of a 10 mM stock solution of Bay K 8644 (RBI, Natick, MA), which was prepared in DMSO, were diluted to a final concentration of 1 μ M in 15 mM Ba²⁺-containing bath solution before they were applied.

Twenty-one negative control HEK 293 cells (transiently transfected with the lacZ gene expression vector pCMV β gal only) were analyzed by the whole-cell variant of the patch clamp method for recording currents. Only one cell displayed a discernable inward barium current; this current was not affected by the presence of 1 μ M Bay K 8644. In addition, application of Bay K 8644 to four cells that did not display Ba²⁺ currents did not result in the appearance of any currents.

Two days after transient transfection of HEK 293 cells with α_1 , α_2 and β_1 subunit-encoding cDNA and the lacZ gene, individual transfectants were assayed for voltage-dependent The currents in nine transfectants were barium currents. recorded. Because the efficiency of transfection of one cell can vary from the efficiency of transfection of another cell, degree of expression of heterologous proteins individual transfectants varies and some cells do not incorporate or express the foreign DNA. Inward barium currents were detected in two of these nine transfectants. these assays, the holding potential of the membrane was -90 mV. The membrane was depolarized in a series of voltage steps to different test potentials and the current in the presence and absence of 1 μ M Bay K 8644 was recorded. The inward barium current was significantly enhanced in magnitude by the addition of Bay K 8644. The largest inward barium current (~160 pA) was recorded when the membrane was depolarized to 0 mV in the presence of 1 μM Bay K 8644. A comparison of the I-V curves, generated by plotting the largest current recorded after each depolarization versus the depolarization voltage, corresponding to recordings conducted in the absence and presence of Bay K 8644 illustrated the enhancement of the voltage-activated current in the presence of Bay K 8644.

Pronounced tail currents were detected in the tracings of currents generated in the presence of Bay K 8644 in HEK 293 cells transfected with α_1 , α_2 and β_1 subunit-encoding cDNA and the lacZ gene, indicating that the recombinant calcium channels responsible for the voltage-activated barium currents recorded in this transfected appear to be DHP-sensitive.

The second of the two transfected cells that displayed inward barium currents expressed a ~50 pA current when the membrane was depolarized from -90 mV. This current was nearly completely blocked by 200 μM cadmium, an established calcium channel blocker.

Ten cells that were transiently transfected with the DNA encoding the α_1 subunit and the lacZ gene were analyzed by

whole-cell patch clamp methods two days after transfection. One of these cells displayed a 30 pA inward barium current. This current amplified 2-fold in the presence of 1 μ M Bay K 8644. Furthermore, small tail currents were detected in the presence of Bay K 8644. These data indicate that expression of the human neuronal calcium channel α_{1D} subunit-encoding cDNA in HEK 293 yields a functional DHP-sensitive calcium channel.

3. Analysis of HEK 293 cells stably transfected with DNA encoding human neuronal calcium channel subunits

Individual stably transfected HEK 293 cells were assayed electrophysiologically for the presence of voltage-dependent barium currents as described for electrophysiological analysis of transiently transfected HEK 293 cells (see Example VII.B.2.c). In an effort to maximize calcium channel activity via cyclic-AMP-dependent kinase-mediated phosphorylation [Pelzer, et al. (1990) Rev. Physiol. Biochem. Pharmacol. 114:107-207], cAMP (Na salt, 250 μ M) was added to the pipet solution and forskolin (10 μ M) was added to the bath solution in some of the recordings. Qualitatively similar results were obtained whether these compounds were present or not.

Barium currents were recorded from stably transfected cells in the absence and presence of Bay K 8644 (1 μM). the cell was depolarized to -10 mV from a holding potential of -90 mV in the absence of Bay K 8644, a current of approximately 35pA with a rapidly deactivating tail current was recorded. During application of Bay K 8644, an identical depolarizing protocol elicited a current of approximately 75 pA, accompanied by an augmented and prolonged tail current. The peak magnitude of currents recorded from this same cell as a function of a series of depolarizing voltages were assessed. The responses in the presence of Bay K 8644 not only increased, but the entire current-voltage relation shifted about -10 mV. Thus, three typical hallmarks of Bay K 8644 action, namely increased current magnitude, prolonged tail currents, and negatively shifted activation voltage, were

observed, clearly indicating the expression of a DHP-sensitive calcium channel in these stably transfected cells. No such effects of Bay K 8644 were observed in untransfected HEK 293 cells, either with or without cAMP or forskolin.

C. Use of pCMV-based vectors and pcDNA1-based vectors for expression of DNA encoding human neuronal calcium channel subunits

1. Preparation of constructs

Additional expression vectors were constructed using pCMV. The full-length α_{1D} cDNA from pVDCCIII(A) (see Example II.A.3.d), the full-length α_2 cDNA, contained on a 3600 bp EcoRI fragment from $HBCaCH\alpha_2$ (see Example IV.B) and a fulllength $eta_{ exttt{1}}$ subunit cDNA from pHBCaCH $eta_{ exttt{1b}}$ RBS(A) (see Example III.B.3) were separately subcloned into plasmid pCMV β gal. Plasmid pCMVetagal was digested with NotI to remove the lacZThe remaining vector portion of the plasmid, referred to as pCMV, was blunt-ended at the NotI sites. The fulllength $lpha_2$ -encoding DNA and eta_1 -encoding DNA, contained on separate EcoRI fragments, were isolated, blunt-ended and separately ligated to the blunt-ended vector fragment of pCMV locating the cDNAs between the CMV promoter and SV40 polyadenylation sites in pCMV. To ligate the $\alpha_{\text{1D}}\text{-encoding cDNA}$ in the polylinkers pCMV, the restriction sites immediately 5' of the CMV promoter and immediately 3' of the polyadenylation site were removed from pCMV. polylinker was added at the NotI site. The polylinker had the following sequence of restriction enzyme recognition sites:

The α_{1D} -encoding DNA, isolated as a <code>BamHI/XhoI</code> fragment from pVDCCIII(A), was then ligated to <code>XbaII/SalI-digested</code> pCMV to place it between the CMV promoter and SV40 polyadenylation site.

Plasmid pCMV contains the CMV promoter as does pcDNA1, but differs from pcDNA1 in the location of splice donor/splice acceptor sites relative to the inserted subunit-encoding DNA. After inserting the subunit-encoding DNA into pCMV, the splice donor/splice acceptor sites are located 3' of the CMV promoter and 5' of the subunit-encoding DNA start codon. After inserting the subunit-encoding DNA into pcDNA1, the splice donor/splice acceptor sites are located 3' of the subunit cDNA stop codon.

2. Transfection of HEK 293 cells

HEK 293 cells were transiently co-transfected with the α_{1D} , α_2 and β_1 subunit-encoding DNA in pCMV or with the α_{1D} , α_2 and β subunit-encoding DNA in pcDNA1 (vectors pVDCCIII(A), pHBCaCH α_2 A and pHBCaCH β_{1b} RBS(A), respectively), as described in Example VII.B.1.a. Plasmid pCMV β gal was included in each transfection as a measure of transfection efficiency. The results of β -galactosidase assays of the transfectants (see Example VII.B.2.), indicated that HEK 293 cells were transfected equally efficiently with pCMV- and pcDNA1-based plasmids. The pcDNA1-based plasmids, however, are presently preferred for expression of calcium channel receptors.

D. Expression in Xenopus laevis oöcytes of RNA encoding human neuronal calcium channel subunits

Various combinations of the transcripts of DNA encoding the human neuronal α_{1D} , α_2 and β_1 subunits prepared in vitro were injected into Xenopus laevis oöcytes. Those injected with combinations that included a_{1D} exhibited voltage-activated barium currents.

1. Preparation of transcripts

Transcripts encoding the human neuronal calcium channel α_{1D} , α_2 and β_1 subunits were synthesized according to the instructions of the mCAP mRNA CAPPING KIT (Strategene, La Jolla, CA catalog #200350). Plasmids pVDCC III.RBS(A), containing pcDNA1 and the α_{1D} cDNA that begins with a ribosome binding site and the eighth ATG codon of the coding sequence (see Example III.A.3.d), plasmid pHBCaCH α_1 A containing pcDNA1 and an α_2 subunit cDNA (see Example IV), and plasmid pHBCaCH β_{1D} RBS(A) containing pcDNA1 and the β_1 DNA lacking intron sequence and containing a ribosome binding site (see Example III), were linearized by restriction digestion. The α_{1D} cDNA- and α_2 subunit-encoding plasmids were digested with XhoI, and the β_1 subunit- encoding plasmid was digested with EcoRV. The DNA insert was transcribed with T7 RNA polymerase.

2. Injection of occytes

Xenopus laevis oöcytes were isolated and defolliculated by collagenase treatment and maintained in 100 mM NaCl, 2 mM KCl, 1.8 mM CaCl₂, 1 mM MgCl₂, 5 mM HEPES, pH 7.6, 20 μ g/ml ampicillin and 25 μ g/ml streptomycin at 19-25°C for 2 to 5 days after injection and prior to recording. For each transcript that was injected into the oöcyte, 6 ng of the specific mRNA was injected per cell in a total volume of 50 nl.

3. Intracellular voltage recordings

Injected occytes were examined for voltage-dependent barium currents using two-electrode voltage clamp methods [Dascal, N. (1987) CRC Crit. Rev. Biochem. 22:317]. The pClamp (Axon Instruments) software package was used in conjunction with a Labmaster 125 kHz data acquisition interface to generate voltage commands and to acquire and analyze data. Quattro Professional was also used in this analysis. Current signals were digitized at 1-5 kHz, and filtered appropriately. The bath solution contained of the following: 40 mM BaCl₂, 36 mM tetraethylammonium chloride

(TEA-Cl), 2 mM KCl, 5 mM 4-aminopyridine, 0.15 mM niflumic acid, 5 mM HEPES, pH 7.6.

a. Electrophysiological analysis of occytes injected with transcripts encoding the human neuronal calcium channel α_1 , α_2 and β_1 -subunits

Uninjected oöcytes were examined by two-electrode voltage clamp methods and a very small (25 nA) endogenous inward Ba^{2+} current was detected in only one of seven analyzed cells.

Obcytes coinjected with α_{1D} , α_2 and β_1 subunit transcripts expressed sustained inward barium currents upon depolarization of the membrane from a holding potential of -90 mV or -50 mV (154 \pm 129 nA, n=21). These currents typically showed little inactivation when test pulses ranging from 140 to 700 msec. were administered. Depolarization to a series of voltages revealed currents that first appeared at approximately -30 mV and peaked at approximately 0 mV.

Application of the DHP Bay K 8644 increased the magnitude of the currents, prolonged the tail currents present upon repolarization of the cell and induced a hyperpolarizing shift in current activation. Bay K 8644 was prepared fresh from a stock solution in DMSO and introduced as a 10x concentrate directly into the 60 μ l bath while the perfusion pump was turned off. The DMSO concentration of the final diluted drug solutions in contact with the cell never exceeded 0.1%. Control experiments showed that 0.1% DMSO had no effect on membrane currents.

Application of the DHP antagonist nifedipine (stock solution prepared in DMSO and applied to the cell as described for application of Bay K 8644) blocked a substantial fraction (91 \pm 6%, n=7) of the inward barium current in oöcytes coinjected with transcripts of the α_{1D} , α_2 and β_1 subunits. A residual inactivating component of the inward barium current typically remained after nifedipine application. The inward barium current was blocked completely by 50 μ M Cd²⁺, but only approximately 15% by 100 μ M Ni²⁺.

The effect of $\omega CgTX$ on the inward barium currents in oöcytes co-injected with transcripts of the $\alpha_{1D},~\alpha_{2},~$ and β_{1} subunits was investigated. $\omega CgTX$ (Bachem, Inc., Torrance CA) was prepared in the 15 mM BaCl₂ bath solution plus 0.1% cytochrome C (Sigma) to serve as a carrier protein. experiments showed that cytochrome C had no effect currents. A series of voltage pulses from a -90 mV holding potential to 0 mV were recorded at 20 msec. intervals. reduce the inhibition of $\omega CgTX$ binding by divalent cations, 73.5 BaCl₂, 15 mΜ made in were recordings tetraethylammonium chloride, and the remaining ingredients identical to the 40 mM Ba2+ recording solution. Bay K 8644 was applied to the cell prior to addition to $\omega CgTX$ in order to determine the effect of $\omega CgTX$ on the DHP-sensitive current component that was distinguished by the prolonged tail currents. The inward barium current was blocked weakly (54 \pm 29%, n=7) and reversibly by relatively high concentrations (10-15 μM) of $\omega CgTX$. The test currents and the accompanying tail currents were blocked progressively within two to three minutes after application of $\omega CgTX$, but both recovered partially as the $\omega CgTX$ was flushed from the bath.

b. Analysis of oöcytes injected with only a transcripts encoding the human neuronal calcium channel α_{1D} or transcripts encoding an α_{1D} and other subunits

The contribution of the α_2 and β_1 subunits to the inward barium current in oöcytes injected with transcripts encoding the α_{1D} , α_2 and β_1 subunits was assessed by expression of the α_{1D} subunit alone or in combination with either the β_1 subunit or the α_2 subunit. In oöcytes injected with only the transcript of a α_{1D} cDNA, no Ba² currents were detected (n=3). In oöcytes injected with transcripts of α_{1D} and β_1 cDNAs, small (108 \pm 39 nA) Ba² currents were detected upon depolarization of the membrane from a holding potential of -90 mV that resembled the currents observed in cells injected with transcripts of α_{1D} , α_2 and β_1 cDNAs, although the magnitude of

the current was less. In two of the four occytes injected with transcripts of the α_{1D} -encoding and β_1 -encoding DNA, the Ba²⁺ currents exhibited a sensitivity to Bay K 8644 that was similar to the Bay K 8644 sensitivity of Ba²⁺ currents expressed in occytes injected with transcripts encoding the α_{1D} α_{1-} , α_{2-} and β_1 subunits.

Three of five oöcytes injected with transcripts encoding the α_{1D} and α_2 subunits exhibited very small Ba²+ currents (15-30 nA) upon depolarization of the membrane from a holding potential of -90 mV. These barium currents showed little or no response to Bay K 8644.

c. Analysis of oöcytes injected with transcripts encoding the human neuronal calcium channel α_2 and/or β_1 subunit

To evaluate the contribution of the α_{1D} α_1 -subunit to the inward barium currents detected in oöcytes co-injected with transcripts encoding the α_{1D} , α_2 and β_1 subunits, oöcytes injected with transcripts encoding the human neuronal calcium channel α_2 and/or β_1 subunits were assayed for barium currents. Oöcytes injected with transcripts encoding the α_2 subunit displayed no detectable inward barium currents (n=5). Oöcytes injected with transcripts encoding a β_1 subunit displayed measurable (54 ± 23 nA, n=5) inward barium currents upon depolarization and oöcytes injected with transcripts encoding the α_2 and β_1 subunits displayed inward barium currents that were approximately 50% larger (80 ± 61 nA, n=18) than those detected in oöcytes injected with transcripts of the β_1 -encoding DNA only.

The inward barium currents in oöcytes injected with transcripts encoding the β_1 subunit or α_2 and β_1 subunits typically were first observed when the membrane was depolarized to -30 mV from a holding potential of -90 mV and peaked when the membrane was depolarized to 10 to 20 mV. Macroscopically, the currents in oöcytes injected with transcripts encoding the α_2 and β_1 subunits or with transcripts encoding the β_1 subunit were indistinguishable. In contrast to the currents in oöcytes co-injected with transcripts of α_{1D} ,

 $\alpha_{\rm 2}$ and $\beta_{\rm 1}$ subunit cDNAs, these currents showed a significant inactivation during the test pulse and a strong sensitivity to the holding potential. The inward barium currents in occytes co-injected with transcripts encoding the α_2 and β_1 subunits usually inactivated to 10-60% of the peak magnitude during a 140-msec pulse and were significantly more sensitive to holding potential than those in occytes co-injected with transcripts encoding the $\alpha_{1D},\ \alpha_2$ and β_1 subunits. Changing the holding potential of the membranes of occytes co-injected with transcripts encoding the α_2 and β_1 subunits from -90 to -50 mV resulted in an approximately 81% (n=11) reduction in the magnitude of the inward barium current of these cells. contrast, the inward barium current measured in occytes coinjected with transcripts encoding the α_{1D} , α_2 and β_1 subunits were reduced approximately 24% (n=11) when the holding potential was changed from -90 to -50 mV.

The inward barium currents detected in occytes injected with transcripts encoding the α_2 and β_1 subunits were pharmacologically distinct from those observed in occytes coinjected with transcripts encoding the α_{1D} , α_{2} and β_{1} subunits. Oöcytes injected with transcripts encoding the α_2 and β_1 currents that displayed inward barium subunits insensitive to Bay K 8644 (n=11). Nifedipine sensitivity was difficult to measure because of the holding potential sensitivity of nifedipine and the current observed in occytes injected with transcripts encoding the α_2 and β_1 subunits. two occytes that were co-injected Nevertheless, transcripts encoding the α_2 and β_1 subunits displayed measurable (25 to 45 nA) inward barium currents depolarized from a holding potential of -50 mV. currents were insensitive to nifedipine (5 to 10 μM). inward barium currents in oöcytes injected with transcripts encoding the α_2 and β_1 subunits showed the same sensitivity to heavy metals as the currents detected in occytes injected with transcripts encoding the α_{1D} , α_{2} and β_{1} subunits.

The inward barium current detected in oöcytes injected with transcripts encoding the human neuronal α_2 and β_1 subunits has pharmacological and biophysical properties that resemble calcium currents in uninjected *Xenopus* oöcytes. Because the amino acids of this human neuronal calcium channel β_1 subunit lack hydrophobic segments capable of forming transmembrane domains, it is unlikely that recombinant β_1 subunits alone can form an ion channel. It is more probable that a homologous endogenous α_1 subunit exists in oöcytes and that the activity mediated by such an α_1 subunit is enhanced by expression of a human neuronal β_1 subunit.

E. Expression of DNA encoding human neuronal calcium channel $\alpha_{1\text{B}},~\alpha_{2\text{B}}$ and $\beta_{1\text{-}2}$ subunits in HEK cells

Transfection of HEK cells

The transient expression of the human neuronal α_{1B-1} , α_{2b} and β_{1-2} subunits was studied in HEK293 cells. The HEK293 cells were grown as a monolayer culture in Dulbecco's modified Eagle's medium (Gibco) containing 5% defined-supplemented bovine calf serum (Hyclone) plus penicillin G (100 U/ml) and steptomycin sulfate (100 μ g/ml). HEK293 cell transfections were mediated by calcium phosphate as described above. Transfected cells were examined for inward Ba²+ currents (I_{Ba}) mediated by voltage-dependent Ca²+ channels.

Cells were transfected (2 x 10 6 per polylysine-coated plate. Standard transfections (10-cm dish) contained 8 μg of pcDNA α_{1B-1} , 5 μg of pHBCaCH α_2 A, 2 μg pHBCaCH β_{1b} RBS(A) (see, Examples II.A.3, IV.B. and III) and 2 μg of CMV β (Clontech) β -glactosidase expression plasmid, and pUC18 to maintain a constant mass of 20 $\mu g/ml$. Cells were analyzed 48 to 72 hours after transfection. Transfection efficiencies ($\pm 10\%$), which were determined by in situ histochemical staining for β -galactosidase activity (Sanes et al. (1986) EMBO J., 5:3133), generally were greater than 50%.

- 2. Electrophysiological analysis of transfectant currents
 - a. Materials and methods

Properties of recombinantly expressed Ca²⁺ channels were studied by whole cell patch-clamp techniques. Recordings were performed on transfected HEK293 cells 2 to 3 days after transfection. Cells were plated at 100,000 to 300,000 cells per polylysine-coated, 35-mm tissue culture dishes (Falcon, Oxnard, CA) 24 hours before recordings. Cells were perfused with 15 mM BaCl₂, 125 mM choline chloride, 1 mM MgCl₂, and 10 mM Hepes (pH = 7.3) adjusted with tetraethylammonium hydroxide (bath solution). Pipettes were filled with 135 mM CsCl, 10 mM EGTA, 10 mM Hepes, 4 mM Mg-adenosine triphosphate (pH = 7.5) adjusted with tetraethylammonium hydroxide. Sylgard (Dow-Corning, Midland, MI)-coated, fire-polished, and filled pipettes had resistances of 1 to 2 megohm before gigohm seals were established to cells.

Bay K 8644 and nifedipine (Research Biochemicals, Natick, MA) were prepared from stock solutions (in dimethyl sulfoxide) and diluted into the bath solution. The dimethyl sulfoxide concentration in the final drug solutions in contact with the cells never exceeded 0.1%. Control experiments showed that 0.1% dimethyl sulfoxide had no efect on membrane currents. $\omega CgTX$ (Bachem, Inc., Torrance CA) was prepared in the 15 mM BaCl₂ bath solution plus 0.1% cytochrome C (Sigma, St. Louis MO) to serve as a carrier protein. Control experiments showed that cytochrome C had no effect on currents. drugs were dissolved in bath solution, and continuously applied by means of puffer pipettes as required for a given experiment. Recordings were performed at room temperature (22° to 25°C). Series resistance compensation (70 to 85%) was employed to minimize voltage error that resulted from pipette access resistance, typically 2 to 3.5 megohm. Current signals were filtered (-3 dB, 4-pole Bessel) at a frequency of 1/4 to 1/5 the sampling rate, which ranged from 0.5 to 3 kHz. Voltage commands were generated and data were acquired with CLAMPEX (pClamp, Axon Instruments, Foster City, CA). reported data are corrected for linear leak and capacitive

components. Exponential fitting of currents was performed with CLAMPFIT (Axon Instruments, Foster City, CA).

b. Results

Transfectants were examined for inward Ba2+ currents (I_{Ba}). Cells cotransfected with DNA encoding $lpha_{\mathrm{1B-1}}$, $lpha_{\mathrm{2b}}$, and $eta_{\mathrm{1-2}}$ subunits expressed high-voltage-activated Ca2+ channels. first appeared when the membrane was depolarized from a holding potential of -90 mV to -20 mV and peaked in magnitude 10 mV. Thirty-nine of 95 cells (12 independent transfections) had $I_{\rm Ba}$ that ranged from 30 to 2700 pA, with a mean of 433 pA. The mean current density was 26 pA/pF, and the highest density was 150 pA/pF. The $I_{\rm Ba}$ typically increased by 2- to 20-fold during the first 5 minutes of recording. Repeated depolarizations during long records often revealed rundown of $I_{\rm Ba}$ usually not exceeding 20% within 10 min. typically activated within 10 ms and inactivated with both a fast time constant ranging from 46 to 105 ms and a slow time constant ranging from 291 to 453 ms (n = 3). showed a complex voltage dependence, such that I_{Ba} elicited at ≥ 20 mV inactivated more slowly than $I_{\rm Ba}$ elicited at lower test voltages, possibly a result of an increase in the magnitude of slow compared to fast inactivation components at higher test voltages.

Recombinant $\alpha_{1B-1}\alpha_{2b}\beta_{1-2}$ channels were sensitive to holding potential. Steady-state inactivation of I_{Ba} , measured after a 30- to 60-s conditioning at various holding potentials, was approximately 50% at holding potential between -60 and -70 mV and approximately 90% at -40 mV. Recovery of I_{Ba} from inactivation was usually incomplete, measuring 55 to 75% of the original magnitude within 1 min. after the holding potential was returned to more negative potentials, possibly indicating some rundown or a slow recovery rate.

Recombinant $\alpha_{1B-1}\alpha_{2b}\beta_{1-2}$ channels were also blocked irreversibly by ω -CgTx concentrations ranging from 0.5 to 10 μ M during the time scale of the experiments. Application of 5 μ M toxin (n = 7) blocked the activity completely within

2 min., and no recovery of $I_{\rm Ba}$ was observed after washing ω -CgTx from the bath for up to 15 min. ${\rm d}^{2+}$ blockage (50 $\mu{\rm M}$) was rapid, complete, and reversible; the DHPs Bay K 8644 (1 $\mu{\rm M}$; n = 4) or nifedipine (5 $\mu{\rm M}$; n = 3) had no discernable effect.

Cells cotransfected with DNA encoding $\alpha_{\text{1B-1}}$, α_{2b} , and $\beta_{\text{1-2}}$ subunits predominantly displayed a single class of saturable, high-affinity ω -CgTx binding The determined sites. dissociation constant (K_d) value was 54.6 \pm 14.5 pM (n = 4). containing Cells transfected with vector the β -galactosidase-encoding DNA or $\alpha_{2b}\beta$ -encoding DNA showed no The binding capacity (B_{max}) of the specific binding. $\alpha_{\text{1B-1}}\alpha_{\text{2b}}\beta$ -transfected cells was 28,710 ± 11,950 sites per cell (n = 4).

These results demonstrate that $\alpha_{1B-1}\alpha_{2b}\beta_{1-2}$ -transfected cells express high-voltage-activated, inactivating Ca²+ channel activity that is irreversibly blocked by ω -CgTx, insensitive to DHPs, and sensitive to holding potential. The activation and inactivation kinetics and voltage sensitivity of the channel formed in these cells are generally consistent with previous characterizations of neuronal N-type Ca²+ channels.

F. Expression of DNA encoding human neuronal calcium channel $\alpha_{\rm 1B-1}$, $\alpha_{\rm 1B-2}$, $\alpha_{\rm 2B}$, $\beta_{\rm 1-2}$ and $\beta_{\rm 1-3}$ subunits in HEK cells

Significant Ba²+ currents were not detected in untransfected HEK293 cells. Furthermore, untransfected HEK293 cells do not express detectable ω -CgTx GVIA binding sites.

In order to approximate the expression of a homogeneous population of trimeric α_{1B} , α_{2b} and β_1 protein complexes in transfected HEK293 cells, the α_{1B} , α_{2b} and β_1 expression levels were altered. The efficiency of expression and assembly of channel complexes at the cell surface were optimized by adjusting the molar ratio of α_{1B} , α_{2b} and β_1 expression plasmids used in the transfections. The transfectants were analyzed for mRNA levels, ω -CgTx GVIA binding and Ca² channel current density in order to determine near optimal channel expression in the absence of immunological reagents for evaluating

protein expression. Higher molar ratios of α_{2b} appeared to increase calcium channel activity.

1. Transfections

HEK293 cells were maintained in DMEM (Gibco #320-1965AJ), 5.5% Defined/Supplemented bovine calf serum (Hyclone #A-2151-L), 100 U/ml penicillin G and 100 μ g/ml streptomycin. phosphate based transient transfections were performed and analyzed as described above. Cells were co-transfected with either 8 μ g pcDNAl $lpha_{ ext{1B-1}}$ (described in Example II.C), 5 μ g pHBCaCH α_2 A (see, Example IV.B.), 2 μ g pHBCaCH β_{1b} RBS(A) (β_{1-2} expression plasmid; see Examples III.A. and IX.E.), and 2 μg $pCMV\beta$ -gal [Clontech, Palo Alto, CA] (2:1.8:1 molar ratio of Ca^{2+} channel subunit expression plasmids) or with 3 μg pcDNA1 $\alpha_{\text{1B-1}}$ or pcDNA1 $\alpha_{\text{1B-2}}$, 11.25 μg pHBCaCH $\alpha_{\text{2}}\text{A}$, 0.75 or 1.0 μg pHBCaCH β_{1b} RBS(A) or pcDNA1 β_{1-3} and 2 μ g pCMV β -gal (2:10.9:1 molar ratio of Ca²⁺ channel subunit expression plasmids). Plasmid pCMV β -gal, a β -galactosidase expression plasmid, was included in the transfections as a marker to permit transfection efficiency estimates by histochemical staining. When less than three subunits were expressed, pCMVPL2, a pCMV promoter-containing vector that lacks a cDNA insert, was substituted to maintain equal moles of pCMV-based DNA in the transfection. pUC18 DNA was used to maintain the total mass of DNA in the transfection at 20 $\mu g/plate$.

RNA from the transfected cells was analyzed by Northern blot analysis for calcium channel subunit mRNA expression using random primed $^{32}\text{P-labeled}$ subunit specific probes. HEK293 cells co-transfected with α_{1B-1} , α_{2b} and β_{1-2} expression plasmids (8, 5 and 2 μg , respectively; molar ratio = 2:1.8:1) did not express equivalent levels of each Ca²+ channel subunit mRNA. Relatively high levels of α_{1B-1} and β_{1-2} mRNAs were expressed, but significantly lower levels of α_{2b} mRNA were expressed. Based on autoradiograph exposures required to produce equivalent signals for all three mRNAs, α_{2b} transcript levels were estimated to be 5 to 10 times lower than α_{1B-1} and

 eta_{1-2} transcript levels. Untransfected HEK293 cells did not express detectable levels of $lpha_{1B-1}$, $lpha_{2b}$, or eta_{1-2} mRNAs.

To achieve equivalent Ca2+ channel subunit mRNA expression levels, a series of transfections was performed with various amounts of $\alpha_{\text{1B-1}}$, α_{2b} and $\beta_{\text{1-2}}$ expression plasmids. Because the $\alpha_{\text{1B-1}}$ and $\beta_{\text{1-2}}$ mRNAs were expressed at very high levels compared to α_{2b} mRNA, the mass of α_{1B-1} and β_{1-2} plasmids was lowered and the mass of α_{2b} plasmid was increased in the transfection experiments. Co-transfection with 3, 11.25 and 0.75 μg of α_{1B} $_{1}$, $lpha_{2b}$ and eta_{1-2} expression plasmids, respectively (molar ratio = 2:10.9:1), approached equivalent expression levels of each Ca $^{2+}$ channel subunit mRNA. The relative molar quantity of α_{2b} expression plasmid to $\alpha_{\text{\tiny 1B-1}}$ and $\beta_{\text{\tiny 1-2}}$ expression plasmids was increased 6-fold. The mass of $\alpha_{\text{\tiny 1B-1}}$ and $\beta_{\text{\tiny 1-2}}$ plasmids in the transfection was decreased 2.67-fold and the mass of α_{2b} plasmid was increased 2.25-fold. The 6-fold molar increase of α_{2b} relative to $\alpha_{1B\text{--}1}$ and $\mathfrak{L}_{1\text{--}2}$ required to achieve near equal abundance mRNA levels is consistent with the previous 5- to 10-fold lower estimate of relative α_{2b} mRNA abundance. ω -CgTx GVIA binding to cells transfected with various amounts of expression plasmids indicated that the 3, 11.25 and 0.75 μg of $lpha_{ ext{1B-1}}$, $lpha_{ ext{2b}}$ and $eta_{ ext{1-2}}$ plasmids, respectively, improved the level of surface expression of channel complexes. increases in the mass of α_{2b} and β_{1-2} expression plasmids while $lpha_{{ ext{\scriptsize 1B-1}}}$ was held constant, and alterations in the mass of the $lpha_{{ ext{\scriptsize 1B-1}}}$ expression plasmid while α_{2b} and β_{1-2} were held constant, indicated that the cell surface expression of $\omega\text{-CgTx}$ GVIA binding sites per cell was nearly optimal. All subsequent transfections were performed with 3, 11.25 and 0.75 μg or 1.0 μg of $lpha_{{ exttt{1B-1}}}$ or $lpha_{{ exttt{1B-2}}}$, $lpha_{{ exttt{2b}}}$ and $eta_{{ exttt{1-2}}}$ or $eta_{{ exttt{1-3}}}$ expression plasmids, respectively.

2. $^{125}\text{I}-\omega\text{-CgTx GVIA}$ binding to transfected cells

Statistical analysis of the K_d and B_{max} values was performed using one-way analysis of variance (ANOVA) followed by the Tukey-Kramer test for multiple pairwise comparisons (ps0.05).

Combinations of human voltage-dependent Ca^{2+} channel subunits, α_{1B-1} , α_{1B-2} , α_{2b} , β_{1-2} and β_{1-3} , were analyzed for saturation binding of $^{125}I-\omega$ -CgTx GVIA. About 200,000 cells were used per assay, except for the α_{1B-1} , α_{1B-2} , $\alpha_{1B-1}\alpha_{2b}$ and $\alpha_{1B-2}\alpha_{2b}$ combinations which were assayed with 1 x 10 6 cells per tube The transfected cells displayed a single-class of saturable, high-affinity binding sites. The values for the dissociation constants (K_d) and binding capacities (B_{max}) were determined for the different combinations. The results are summarized as follows:

Subunit Combination	K _d (pM)	B _{max} (sites/cell)
$\alpha_{\mathtt{1B-1}}\alpha_{\mathtt{2b}}\beta_{\mathtt{1-2}}$	$54.9 \pm 11.1 (n=4)$	45,324 ± 15,606
$lpha_{ ext{ iny 1B-1}}lpha_{ ext{ iny 2b}}eta_{ ext{ iny 1-3}}$	$53.2 \pm 3.6 \ (n=3)$	91,004 ± 37,654
$lpha_{\mathtt{1B-1}}eta_{\mathtt{1-2}}$	$17.9 \pm 1.9 (n=3)$	5,756 ± 2,163
$\alpha_{\mathtt{1B-1}} eta_{\mathtt{1-3}}$	$17.9 \pm 1.6 (n=3)$	8,729 ± 2,980
$lpha_{\mathtt{1B-1}}lpha_{\mathtt{2b}}$	$84.6 \pm 15.3 (n=3)$	2,256 ± 356
$lpha_{ exttt{1B-1}}$	$31.7 \pm 4.2 (n=3)$	757 ± 128
$\alpha_{_{\mathbf{1B-2}}}\alpha_{\mathbf{2b}}\beta_{\mathbf{1-2}}$	$53.0 \pm 4.8 (n=3)$	19,371 ± 3,798
$lpha_{\mathtt{1B-2}}lpha_{\mathtt{2b}}eta_{\mathtt{1-3}}$	$44.3 \pm 8.1 (n=3)$	37,652 ± 8,129
$\alpha_{\mathtt{1B-2}}\beta_{\mathtt{1-2}}$	$16.4 \pm 1.2 \text{ (n=3)}$	2,126 ± 412
$\alpha_{\mathtt{1B-2}}eta_{\mathtt{1-3}}$	$22.2 \pm 5.8 $ (n=3)	2,944 ± 1,168
$\alpha_{\mathtt{1B-2}}\alpha_{\mathtt{2b}}$	N.D. (n=3)	N.D.
α_{1B-2} * N.D. = not detectable	N.D.	N.D.
Coll		

Cells transfected with subunit combinations lacking either the α_{1B-1} or the α_{1B-2} subunit did not exhibit any detectable $^{125}\text{I}-\omega$ -CgTx GVIA binding (≤ 600 sites/cell). $^{125}\text{I}-\omega$ -CgTx GVIA binding to HEK293 cells transfected with α_{1B-2} alone or $\alpha_{1B-2}\alpha_{2b}$ was too low for reliable Scatchard analysis of the data. Comparison of the K_d and B_{max} values revealed several relationships between specific combinations of subunits and the binding affinities and capacities of the transfected cells. In cells transfected with all three subunits, ($\alpha_{1B-1}\alpha_{2b}\beta_{1-2}$ -, $\alpha_{1B-1}\alpha_{2b}\beta_{1-3}$ -, $\alpha_{1B-2}\alpha_{2b}\beta_{1-2}$ -, or $\alpha_{1B-2}\alpha_{2b}\beta_{1-3}$ -transfectants) the K_d values were indistinguishable (p>0.05), ranging from 44.3

 \pm 8.1 pM to 54.9 \pm 11.1 pM. In cells transfected with twosubunit combinations lacking the α_{2b} subunit $(\alpha_{1B-1}\beta_{1-2}, \alpha_{1B-1}\beta_{1-3},$ $\alpha_{{\rm 1B-2}}\beta_{{\rm 1-2}}$ or $\alpha_{{\rm 1B-2}}\beta_{{\rm 1-3}})$ the K_d values were significantly lower than the three-subunit combinations (p<0.01), ranging from 16.4 \pm 1.2 to 22.2 \pm 5.8 pM. Cells transfected with only the $\alpha_{\text{\tiny 1B-1}}$ subunit had a K_d value of 31.7 \pm 4.2 pM, a value that was not different from the two-subunit combinations lacking $lpha_{2b}$ (p<0.05). As with the comparison between the four $\alpha_{1B}\alpha_{2b}\beta_1$ versus $\alpha_{\text{1B}}\beta_{\text{1}}$ combinations, when the $\alpha_{\text{1B-1}}$ was co-expressed with $\alpha_{2b},$ the K_d increased significantly (p<0.05) from 31.7 \pm 4.2 to 84.6 \pm 5.3 pM. These data demonstrate that co-expression of the α_{2b} subunit with α_{1B-1} , $\alpha_{1B-1}\beta_{1-2}$, $\alpha_{1B-1}\beta_{1-3}$, $\alpha_{1B-2}\beta_{1-2}$ or $\alpha_{1B-2}\beta_{1-3}$ subunit combinations results in lower binding affinity of the cell surface receptors for $^{125}I-\omega$ -CgTx GVIA. The B_{max} values of cells transfected with various subunit combinations also differed considerably. Cells transfected with the $lpha_{\scriptscriptstyle{1B-1}}$ subunit alone expressed a low but detectable number of binding sites (approximately 750 binding sites/cell). When the $\alpha_{\scriptscriptstyle 1B-1}$ subunit was co-expressed with the α_{2b} subunit, the binding capacity increased approximately three-fold while co-expression of a $eta_{ exttt{1-}}$ $_{2}$ or $eta_{ ext{1-3}}$ subunit with $lpha_{ ext{1B-1}}$ resulted in 8- to 10-fold higher Cells transfected with all expression of surface binding. three subunits expressed the highest number of cell surface receptors. The binding capacities of cells transfected with $\alpha_{{\rm 1B-1}}\alpha_{{\rm 2b}}\beta_{{\rm 1-3}}$ or $\alpha_{{\rm 1B-2}}\alpha_{{\rm 2b}}\beta_{{\rm 1-3}}$ combinations were approximately two-fold higher than the corresponding combinations containing the $eta_{\scriptscriptstyle 1-2}$ Likewise, cells transfected with $\alpha_{1B-1}\alpha_{2b}\beta_{1-2}$ or subunit. $lpha_{{\scriptscriptstyle 1B}-1}lpha_{{\scriptscriptstyle 2b}}eta_{{\scriptscriptstyle 1-3}}$ combinations expressed approximately 2.5-fold more binding sites per cell than the corresponding combinations containing $\alpha_{\text{1B-2}}$. In all cases, co-expression of the α_{2b} subunit with $\alpha_{\text{\tiny 1B}}$ and $\beta_{\text{\tiny 1}}$ increased the surface receptor density compared to cells transfected with only the corresponding $lpha_{\scriptscriptstyle 1B}$ and eta_1 combinations; approximately 8-fold for $lpha_{1B-1}lpha_{2b}eta_{1-2}$, 10-fold for $\alpha_{\text{1B-1}}\alpha_{\text{2b}}\beta_{\text{1-3}}$, 9-fold for $\alpha_{\text{1B-2}}\alpha_{\text{2b}}\beta_{\text{1-2}}$, and 13-fold for $\alpha_{\text{1B-2}}\alpha_{\text{2b}}\beta_{\text{1-3}}$. Thus, comparison of the ${\bf B}_{\tt max}$ values suggests that the toxin-binding subunit, $\alpha_{\text{1B-1}}$ or $\alpha_{\text{1B-2}}$, is more efficiently expressed and assembled on the cell surface when co-ex-pressed with either the α_{2b} or the $\beta_{\text{1-2}}$ or $\beta_{\text{1-3}}$ subunit, and most efficiently expressed when α_{2b} and β_{1} subunits are present.

3. Electrophysiology

Functional expression of $\alpha_{1B-1}\alpha_{2b}\beta_{1-2}$ and $\alpha_{1B-1}\beta_{1-2}$ subunit combinations was evaluated using the whole-cell recording technique. Transfected cells that had no contacts with surrounding cells and simple morphology were used approximately 48 hours after transfection for recording. The pipette solution was (in mM) 135 CsCl, 10 EgTA, 1 MgCl₂, 10 HEPES, and 4 mM Mg-ATP (pH 7.3, adjusted with TEA-OH). The external solution was (in mM) 15 BaCl₂, 125 Choline Cl, 1 MgCl₂, and 10 HEPES (pH 7.3, adjusted with TEA-OH). ω -CgTx GVIA (Bachem) was prepared in the external solution with 0.1% cytochrome C (Sigma) to serve as a carrier. Control experiments showed that cytochrome C had no effect on the Ba²⁺ current.

The macroscopic electrophysiological properties of Ba²⁺ currents in cells transfected with various amounts of the α_{2b} expression plasmid with the relative amounts of $lpha_{\scriptscriptstyle 1B-1}$ and $eta_{\scriptscriptstyle 1-2}$ plasmids held constant were examined. The amplitudes and densities of the Ba2+ currents (15 mM BaCl2) recorded from whole cells of these transfectants differed dramatically. average currents from 7 to 11 cells of three types of transfections (no $\alpha_{2b};$ 2:1.8:1 $[\alpha_{1B-1}:\alpha_{2b}:\beta_{1-2}]$ molar ratio; and 2:10.9:1 $[\alpha_{1B-1}:\alpha_{2b}:\beta_{1-2}]$ molar ratio) were determined. smallest currents (range: 10 to 205 pA) were recorded when α_{2b} was not included in the transfection, and the largest currents (range: 50 to 8300 pA) were recorded with the 2:10.9:1 ratio of $lpha_{{ ext{\scriptsize 1B-1}}}lpha_{{ ext{\scriptsize 2b}}}eta_{{ ext{\scriptsize 1-2}}}$ plasmids, the ratio that resulted in near equivalent mRNA levels for each subunit transcript. When the amount of $lpha_{2b}$ plasmid was adjusted to yield approximately an equal abundance of subunit mRNAs, the average peak Ba^{2+} current increased from 433 pA to 1,824 pA (4.2-fold) with a corresponding increase in average current density from 26 pA/pF to 127 pA/pF (4.9-fold). This increase is in the presence of a 2.7-fold decrease in the mass of $\alpha_{\text{\tiny 1B-1}}$ and $\beta_{\text{\tiny 1-2}}$ expression plasmids in the transfections.

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In all transfections, the magnitudes of the Ba²⁺ currents did not follow a normal distribution.

To compare the subunit combinations and determine the effects of α_{2h} , the current-voltage properties of cells transfected with $\alpha_{1B-1}\beta_{1-2}$ or with $\alpha_{1B-1}\alpha_{2b}\beta_{1-2}$ in either the 2:1.8:1 $(\alpha_{1B-1}:\alpha_{2b}:\beta_{1-2})$ molar ratio or the 2:10.9:1 $(\alpha_{1B-1}:\alpha_{2b}:\beta_{1-2})$ molar ratio transfectants were examined. The extreme examples of no α_{2b} and 11.25 μ g α_{2b} (2:10.9:1 molar ratio) showed no significant differences in the current voltage plot at test potentials between 0 mV and +40 mV (p<0.05). The slight differences observed at either side of the peak region of the current voltage plot were likely due to normalization. The very small currents observed in the $\alpha_{18-1}\beta_{1-2}$ transfected cells have a substantially higher component of residual leak relative to the barium current that is activated by the test pulse. When the current voltage plots are normalized, this leak is a much greater component than in the $\alpha_{18-1}\alpha_{2b}\beta_{1-2}$ transfected cells and as a result, the current-voltage plot appears broader. This is the most likely explanation of the apparent differences in the current voltage plots, especially given the fact that the current-voltage plot for the $\alpha_{1B-1}\beta_{1-2}$ transfected cells diverge on both sides of the peak. Typically, when the voltagedependence activation is shifted, the entire current-voltage plot is shifted, which was not observed. To qualitatively compare the kinetics of each, the average responses of test pulses from -90 mV to 10 mV were normalized and plotted. significant differences in activation or inactivation kinetics of whole-cell Ba2+ currents were observed with any combination.

G. Expression of DNA encoding human neuronal calcium channel $\alpha_{1E-3}\alpha_{2B}\beta_{1-3}$ and $\alpha_{1E-1}\alpha_{2B}\beta_{1-3}$ subunits in HEK cells Functional expression of the $\alpha_{1E-1}\alpha_{2B}\beta_{1-3}$ and $\alpha_{1E-3}\alpha_{2B}\beta_{1-3}$, as well as α_{1E-3} was evaluated using the whole cell recording technique.

1. Methods

Recordings were performed on transiently transfected HEK 293 cells two days following the transfection, from cells that had no contacts with surrounding cells and which had simple morphology.

The internal solution used to fill pipettes for recording the barium current from the transfected recombinant calcium channels was (in mM) 135 CsCl, 10 EGTA, 1 MgCl₂, 10 HEPES, and 4 mM Mg-ATP (pH 7.4-7.5, adjusted with TEA-OH). The external solution for recording the barium current was (in mM) 15 BaCl2, 150 Choline Cl, 1 MgCl₂, and 10 HEPES and 5 TEA-OH (pH 7.3, adjusted with TEA-OH). In experiments in which Ca2+ was replaced for Ba2+, a Laminar flow chamber was used in order to completely exchange the extracellular solution and prevent any mixing of Ba^{2+} and Ca^{2+} . ω -CgTx GVIA was prepared in the external solution with 0.1% cytochrome C to serve as a carrier, the toxin was applied by pressurized puffer pipette. resistance was compensated 70-85% and currents were analyzed only if the voltage error from series resistance was less than Leak resistance and capacitance was corrected by subtracting the scaled current observed with the P/-4 protocol as implemented by pClamp (Axon Instruments).

2. Electrophysiology Results

Cells transfected with $\alpha_{1E-1}\alpha_{2b}\beta_{1-3}$ or $\alpha_{1E-3}\alpha_{2b}\beta_{1-3}$ showed strong barium currents with whole cell patch clamp recordings. Cells expressing $\alpha_{1E-3}\alpha_{2B}\beta_{1-3}$ had larger peak currents than those expressing $\alpha_{1B-1}\alpha_{2b}\beta_{1-3}$. In addition, the kinetics of activation and inactivation are clearly substantially faster in the cells expressing α_{1E} calcium channels. HEK 293 cells expressing α_{1E-3} alone have a significant degree of functional calcium channels, with properties similar to those expressing $\alpha_{1E}\alpha_{2b}\beta_{1-3}$ but with substantially smaller peak barium currents. Thus, with α_{1E} , the α_2 and β_1 subunits are not required for functional expression of α_{1E} mediated calcium channels, but do substantially increase the number of functional calcium channels.

Examination of the current voltage properties of $\alpha_{1E}\alpha_{2b}\beta_{1.3}$ expressing cells indicates that $\alpha_{1E-3}\alpha_{2b}\beta_{1-3}$ is a high-voltage

activated calcium channel and the peak current is reached at a potential only slightly less positive than other neuronal calcium channels also expressing α_{2b} and β_1 , and α_{1B} and α_{1D} . Current voltage properties of $\alpha_{1E-1}\alpha_{2b}\beta_{1-3}$ and $\alpha_{1E-3}\alpha_{2b}\beta_{1-3}$ are statistically different from those of $\alpha_{1E-1}\alpha_{2b}\beta_{1-3}$. Current voltage curves for $\alpha_{1E-1}\alpha_{2b}\beta_{1-3}$ and $\alpha_{1E-3}\alpha_{2b}\beta_{1-3}$ peak at approximately +5mV, as does the current voltage curve for α_{1E-3} alone.

The kinetics and voltage dependence of inactivation using both prepulse (200 ms) and steady-state inactivation was examined. $\alpha_{\rm IE}$ mediated calcium channels are rapidly inactivated relative to previously cloned calcium channels and other high voltage-activated calcium channels. $\alpha_{\rm IE-3}\alpha_{\rm 2b}\beta_{\rm 1-3}$ mediated calcium channels are inactivated rapidly and are thus sensitive to relatively brief (200 ms) prepulses as well as long prepulses (>20s steady state inactivation), but recover rapidly from steady state inactivation. The kinetics of the rapid inactivation has two components, one with a time constant of approximately 25 ms and the other approximately 400 ms.

To determine whether $\alpha_{\rm IE}$ mediated calcium channels have properties of low voltage activated calcium channels, the details of tail currents activated by a test pulse ranging -60 to +90 mV were measured at -60 mV. Tail currents recorded at -60 mV could be well fit by a single exponential of 150 to 300 μs ; at least an order of magnitude faster than those typically observed with low voltage-activated calcium channels.

HEK 293 cells expressing $\alpha_{1E\cdot3}\alpha_{2b}\beta_{1\cdot3}$ flux more current with Ba^{2+} as the charge carrier and currents carried by Ba^{2+} and Ca^{2+} have different current-voltage properties. Furthermore, the time course of inactivation is slower and the amount of prepulse inactivation less with Ca^{2+} as the charge carrier.

While the invention has been described with some specificity, modifications apparent to those with ordinary skill in the art may be made without departing from the scope of the invention. Since such modifications will be apparent to

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those of skill in the art, it is intended that this invention be limited only by the scope of the appended claims.

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SEQUENCE LISTING

- (1) GENERAL INFORMATION:
 - (i) APPLICANT:
 - (A) NAME: THE SALK INSTITUTE BIOTECHNOLY/INDUSTRIAL ASSOCIATES
 - (B) STREET: 505 COAST BLVD SOUTH, SUITE 300
 - (C) CITY: La Jolla
 - (D) STATE: California
 - (E) COUNTRY: USA
 - (F) POSTAL CODE (ZIP): 92037
 - (ii) TITLE OF INVENTION: HUMAN CALCIUM CHANNEL COMPOSITIONS AND METHODS
 - (iii) NUMBER OF SEQUENCES: 38
 - (iv) COMPUTER READABLE FORM:
 - (A) MEDIUM TYPE: Floppy disk

 - (B) COMPUTER: IBM PC compatible (C) OPERATING SYSTEM: PC-DOS/MS-DOS
 - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
 - (vi) CURRENT APPLICATION DATA:
 - (A) APPLICATION NUMBER:(B) FILING DATE:

 - (C) CLASSIFICATION:
 - (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: 08/149,097
 - (B) FILING DATE: 5-NOV-1993
 - (vii) PRIOR APPLICATION DATA:
 - (A) APPLICATION NUMBER: 08/105,536
 - (B) FILING DATE: 11-AUG-1993
- (2) INFORMATION FOR SEQ ID NO:1:
 - (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 7635 base pairs(B) TYPE: nucleic acid

 - (C) STRANDEDNESS: double
 - (D) TOPOLOGY: linear
 - (ii) MOLECULE TYPE: DNA (genomic)
 - (ix) FEATURE:

 - (A) NAME/KEY: CDS
 (B) LOCATION: 511..6996
 - (ix) FEATURE:

 - (A) NAME/KEY: 5'UTR (B) LOCATION: 1..510
 - (ix) FEATURE:

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(A) NAME/KEY: 3'UTR (B) LOCATION: 6994..7635

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

CHE, DESCRIPTION. SEQ ID NO:1:	
GGGCGAGCGC CTCCGTCCCC GGATGTGAGC TCCGGCTGCC CGCGGTCCCG AGCCA	GCGGC 60
GCGCGGCGG CGCCGGGCAC CGCGGCGGC GGGCAGACGG GCGGG	CATGG 120
GGGGAGCGCC GAGCGGCCCC GGCGGCCGGG CCGGCATCAC CGCGGCGTCT CTCCG	CTAGA 180
GGAGGGGACA AGCCAGTTCT CCTTTGCAGC AAAAAATTAC ATGTATATAT TATTA	AGATA 240
ATATATACAT TGGATTTTAT TTTTTTAAAA AGTTTATTTT GCTCCATTTT TGAAAA	AAGAG 300
AGAGCTTGGG TGGCGAGCGG TTTTTTTTTA AAATCAATTA TCCTTATTTT CTGTTA	ATTTG 360
TCCCCGTCCC TCCCCACCCC CCTGCTGAAG CGAGAATAAG GGCAGGGACC GCGGCT	ICCTA 420
CCTCTTGGTG ATCCCCTTCC CCATTCCGCC CCCGCCCCAA CGCCCAGCAC AGTGCC	CCTGC 480
ACACAGTAGT CGCTCAATAA ATGTTCGTGG ATG ATG ATG ATG ATG ATG ATG ATG Met Met Met Met Met Met Met 1	AAA 534 Lys
AAA ATG CAG CAT CAA CGG CAG CAG CAA GCG GAC CAC GCG AAC GAG G Lys Met Gln His Gln Arg Gln Gln Gln Ala Asp His Ala Asn Glu A 10 15 20	GCA 582 Ala
AAC TAT GCA AGA GGC ACC AGA CTT CCT CTT TCT GGT GAA GGA CCA A Asn Tyr Ala Arg Gly Thr Arg Leu Pro Leu Ser Gly Glu Gly Pro T 30 35	ACT 630 Thr 40
TCT CAG CCG AAT AGC TCC AAG CAA ACT GTC CTG TCT TGG CAA GCT G Ser Gln Pro Asn Ser Ser Lys Gln Thr Val Leu Ser Trp Gln Ala A 45 50 55	GCA 678 Ala
ATC GAT GCT GCT AGA CAG GCC AAG GCT GCC CAA ACT ATG AGC ACC TIle Asp Ala Ala Arg Gln Ala Lys Ala Ala Gln Thr Met Ser Thr S	CT 726 Ser
GCA CCC CCA CCT GTA GGA TCT CTC TCC CAA AGA AAA CGT CAG CAA TA Ala Pro Pro Pro Val Gly Ser Leu Ser Gln Arg Lys Arg Gln Gln To 75 80 85	AC 774 Yr
GCC AAG AGC AAA AAA CAG GGT AAC TCG TCC AAC AGC CGA CCT GCC C Ala Lys Ser Lys Lys Gln Gly Asn Ser Ser Asn Ser Arg Pro Ala A 90 95 100	GC 822 xg
GCC CTT TTC TGT TTA TCA CTC AAT AAC CCC ATC CGA AGA GCC TGC AAA Leu Phe Cys Leu Ser Leu Asn Asn Pro Ile Arg Arg Ala Cys I 110 115	TT 870 le 20
AGT ATA GTG GAA TGG AAA CCA TTT GAC ATA TTT ATA TTA TTG GCT A Ser Ile Val Glu Trp Lys Pro Phe Asp Ile Phe Ile Leu Leu Ala I 125 130 135	TT 918 le

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TTT	GCC Ala	AAT Asn	TGT Cys 140	GTG Val	GCC Ala	TTA Leu	GCT Ala	ATT Ile 145	TAC Tyr	ATC Ile	CCA Pro	TTC Phe	CCT Pro 150	GAA Glu	GAT Asp		966
GAT Asp	TCT Ser	AAT Asn 155	TCA Ser	ACA Thr	AAT Asn	CAT His	AAC Asn 160	TTG Leu	GAA Glu	AAA Lys	GTA Val	GAA Glu 165	TAT Tyr	GCC Ala	TTC Phe	1	.014
CTG Leu	ATT Ile 170	ATT Ile	TTT Phe	ACA Thr	GTC Val	GAG Glu 175	ACA Thr	TTT Phe	TTG Leu	AAG Lys	ATT Ile 180	ATA Ile	GCG Ala	TAT Tyr	GGA Gly	1	.062
Leu 185	Leu	Leu	His	Pro	AAT Asn 190	Ala	Tyr	Val	Arg	Asn 195	Gly	Trp	Asn	Leu	Leu 200	1	.110
GAT Asp	TTT Phe	GTT Val	ATA Ile	GTA Val 205	ATA Ile	GTA Val	GGA Gly	TTG Leu	TTT Phe 210	AGT Ser	GTA Val	ATT Ile	TTG Leu	GAA Glu 215	CAA Gln	1	
TTA Leu	ACC Thr	AAA Lys	GAA Glu 220	ACA Thr	GAA Glu	GGC Gly	GGG Gly	AAC Asn 225	CAC His	TCA Ser	AGC Ser	GGC Gly	AAA Lys 230	TCT Ser	GGA Gly	1	L206
GGC	TTT Phe	GAT Asp 235	GTC Val	AAA Lys	GCC Ala	CTC Leu	CGT Arg 240	GCC Ala	TTT Phe	CGA Arg	GTG Val	TTG Leu 245	CGA Arg	CCA Pro	CTT Leu	1	L254
Arg	Leu 250	Val	Ser	Gly	GTG Val	Pro 255	Ser	Leu	Gln	Val	Val 260	Leu	Asn	Ser	Ile	1	1302
ATA Ile 265	AAA Lys	GCC Ala	ATG Met	GTT Val	CCC Pro 270	CTC Leu	CTT Leu	CAC His	ATA Ile	GCC Ala 275	CTT Leu	TTG Leu	GTA Val	TTA Leu	TTT Phe 280	1	1350
Val	Ile	Ile	Ile	Tyr 285	GCT Ala	Ile	Ile	Gly	Leu 290	Glu	Leu	Phe	Ile	Gly 295	Lys	1	1398
ATG Met	CAC His	AAA Lys	ACA Thr 300	TGT Cys	TTT	TTT	GCT Ala	GAC Asp 305	Ser	GAT Asp	ATC Ile	GTA Val	GCT Ala 310	GAA Glu	GAG Glu	:	1446
GAC Asp	CCA Pro	GCT Ala 315	Pro	TGT Cys	GCG Ala	TTC Phe	TCA Ser 320	GGG Gly	TAA Asn	GGA Gly	CGC	CAG Gln 325	Cys	ACT Thr	GCC Ala	:	1494
AAT Asn	GGC Gly 330	Thr	GAA Glu	TGT Cys	AGG Arg	AGT Ser 335	Gly	TGG	GTT Val	GGC Gly	CCG Pro 340	Asn	GGA Gly	GGC	ATC Ile	;	1542
ACC Thr 345	Asn	TTT Phe	GAT Asp	AAC Asn	TTT Phe 350	Ala	TTT Phe	GCC	ATG Met	Leu 355	Ini	GTG Val	TTT Phe	CAG Gln	TGC Cys 360		1590

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ATC Ile	C ACC	C ATO	G GAG	GGC Gly 365	Trp	ACA Thr	GAC Asp	GTO Val	CTC Leu 370	і Туг	TGG	ATG Met	AAT Asn	GAT Asp 375	GCT Ala	1638
AT(GG# Gly	Y Phe	GAA Glu 380	ι тел	CCC Pro	TGG Trp	GTG Val	TAT Tyr 385	Phe	GTC Val	Ser	CTC Leu	GTC Val 390	Ile	TTT Phe	1686
GGG	TCA Ser	TTT Phe 395	: Pne	GTA Val	CTA Leu	AAT Asn	CTT Leu 400	Val	CTT Leu	GGT	GTA Val	TTG Leu 405	AGC Ser	GGA Gly	GAA Glu	1734
TTC Phe	TCA Ser 410	T)YE	GAA Glu	AGA Arg	GAG Glu	AAG Lys 415	GCA Ala	AAA Lys	GCA Ala	CGG Arg	GGA Gly 420	Asp	TTC Phe	CAG Gln	AAG Lys	1782
CTC Leu 425	- ALY	GAG Glu	AAG Lys	CAG Gln	CAG Gln 430	CTG Leu	GAG Glu	GAG Glu	GAT Asp	CTA Leu 435	AAG Lys	GGC Gly	TAC Tyr	TTG Leu	GAT Asp 440	1830
TGG Trp	ATC Ile	ACC	CAA Gln	GCT Ala 445	GAG Glu	GAC Asp	ATC Ile	GAT Asp	CCG Pro 450	GAG Glu	AAT Asn	GAG Glu	GAA Glu	GAA Glu 455	GGA Gly	1878
GGA Gly	GAG Glu	GAA Glu	GGC Gly 460	AAA Lys	CGA Arg	AAT Asn	ACT Thr	AGC Ser 465	ATG Met	CCC Pro	ACC Thr	AGC Ser	GAG Glu 470	ACT Thr	GAG Glu	1926
TCT Ser	GTG Val	AAC Asn 475	ACA Thr	GAG Glu	AAC Asn	GTC Val	AGC Ser 480	GGT Gly	GAA Glu	GGC Gly	GAG Glu	AAC Asn 485	CGA Arg	GGC Gly	TGC Cys	1974
TGT Cys	GGA Gly 490	AGT Ser	CTC Leu	TGT Cys	CAA Gln	GCC Ala 495	ATC Ile	TCA Ser	AAA Lys	TCC Ser	AAA Lys 500	CTC Leu	AGC Ser	CGA Arg	CGC Arg	2022
TGG Trp 505	CGT Arg	CGC Arg	TGG Trp	AAC Asn	CGA Arg 510	TTC Phe	AAT Asn	CGC Arg	AGA Arg	AGA Arg 515	TGT Cys	AGG Arg	GCC Ala	GCC Ala	GTG Val 520	2070
AAG Lys	TCT Ser	GTC Val	ACG Thr	TTT Phe 525	TAC Tyr	TGG Trp	CTG Leu	GTT Val	ATC Ile 530	GTC Val	CTG Leu	GTG Val	TTT Phe	CTG Leu 535	AAC Asn	2118
ACC Thr	TTA Leu	ACC Thr	ATT Ile 540	TCC Ser	TCT Ser	GAG Glu	CAC His	TAC Tyr 545	AAT Asn	CAG Gln	CCA Pro	Asp	TGG Trp 550	TTG Leu	ACA Thr	2166
CAG Gln	ATT Ile	CAA Gln 555	GAT Asp	ATT Ile	GCC Ala	Asn	AAA Lys 560	GTC Val	CTC Leu	TTG Leu	GCT Ala	CTG Leu 565	TTC Phe	ACC Thr	TGC Cys	2214
GAG Glu	ATG Met 570	CTG Leu	GTA Val	AAA Lys	Met	TAC I Tyr : 575	AGC Ser	TTG Leu	GGC Gly	Leu	CAA Gln 580	GCA '	TAT Tyr	TTC Phe	GTC Val	2262

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TCT Ser 585	CTT Leu	TTC Phe	AAC Asn	CGG Arg	TTT Phe 590	GAT Asp	TGC Cys	TTC Phe	GTG Val	GTG Val 595	TGT Cys	GGT Gly	GGA Gly	ATC Ile	ACT Thr 600	2310
GAG Glu	ACG Thr	ATC Ile	TTG Leu	GTG Val 605	GAA Glu	CTG Leu	GAA Glu	ATC Ile	ATG Met 610	TCT Ser	CCC Pro	CTG Leu	GGG Gly	ATC Ile 615	TCT Ser	2358
GTG Val	TTT Phe	CGG Arg	TGT Cys 620	GTG Val	CGC Arg	CTC Leu	TTA Leu	AGA Arg 625	ATC Ile	TTC Phe	AAA Lys	GTG Val	ACC Thr 630	AGG Arg	CAC His	2406
	ACT Thr															2454
TCC Ser	ATC Ile 650	GCT Ala	TCG Ser	CTG Leu	TTG Leu	CTT Leu 655	CTG Leu	CTT Leu	TTT Phe	CTC Leu	TTC Phe 660	ATT Ile	ATC Ile	ATC Ile	TTT Phe	2502
TCC Ser 665	TTG Leu	CTT Leu	GGG Gly	ATG Met	CAG Gln 670	CTG Leu	TTT Phe	GGC Gly	GGC Gly	AAG Lys 675	TTT Phe	AAT Asn	TTT Phe	GAT Asp	GAA Glu 680	2550
ACG Thr	CAA Gln	ACC Thr	AAG Lys	CGG Arg 685	AGC Ser	ACC Thr	TTT Phe	GAC Asp	AAT Asn 690	TTC Phe	CCT Pro	CAA Gln	GCA Ala	CTT Leu 695	CTC Leu	2598
ACA Thr	GTG Val	TTC Phe	CAG Gln 700	ATC Ile	CTG Leu	ACA Thr	GGC Gly	GAA Glu 705	GAC Asp	TGG Trp	AAT Asn	GCT Ala	GTG Val 710	ATG Met	TAC Tyr	2646
GAT Asp	GGC Gly	ATC Ile 715	ATG Met	GCT Ala	TAC Tyr	GGG Gly	GGC Gly 720	CCA Pro	TCC Ser	TCT Ser	TCA Ser	GGA Gly 725	ATG Met	ATC Ile	GTC Val	2694
TGC Cys	ATC Ile 730	TAC Tyr	TTC Phe	ATC Ile	ATC Ile	CTC Leu 735	TTC Phe	ATT Ile	TGT Cys	GGT Gly	AAC Asn 740	TAT Tyr	ATT Ile	CTA Leu	CTG Leu	2742
AAT Asn 745	GTC Val	TTC Phe	TTG Leu	GCC Ala	ATC Ile 750	GCT Ala	GTA Val	GAC Asp	AAT Asn	TTG Leu 755	GCT Ala	GAT Asp	GCT Ala	GAA Glu	AGT Ser 760	2790
CTG Leu	AAC Asn	ACT Thr	GCT Ala	CAG Gln 765	AAA Lys	GAA Glu	GAA Glu	GCG Ala	GAA Glu 770	GAA Glu	AAG Lys	GAG Glu	AGG Arg	AAA Lys 775	AAG Lys	2838
ATT Ile	GCC Ala	AGA Arg	AAA Lys 780	GAG Glu	AGC Ser	CTA Leu	GAA Glu	AAT Asn 785	Lys	AAG Lys	AAC Asn	AAC Asn	AAA Lys 790	CCA Pro	GAA Glu	2886
.GTC Val	AAC Asn	CAG Gln 795	Ile	GCC Ala	AAC Asn	AGT Ser	GAC Asp 800	Asn	AAG Lys	GTT Val	ACA Thr	ATT Ile 805	GAT Asp	GAC Asp	TAT Tyr	2934

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AGA Arg	GAA Glu 810	GIU	GA7 1 Asp	GAA Glu	ASP	AAG Lys 815	Asp	CCC Pro	TAI Tyr	CCG Pro	CCI Pro 820	суя	GAT S Asp	GTO Val	CCA Pro	2982
GTA Val 825	GIA	GAA Glu	A GAG	GAA Glu	GAG Glu 830	Glu	GAG Glu	GAG Glu	GAG Glu	GAT Asp 835	Glu	CCI Pro	GAG Glu	GTI Val	CCT Pro 840	3030
GCC	GGA Gly	CCC Pro	CGT Arg	CCT Pro 845	Arg	AGG Arg	ATC	TCG Ser	GAG Glu 850	Leu	AAC Asn	ATG Met	AAG Lys	GAA Glu 855	AAA Lys	3078
ATT	GCC	CCC Pro	ATC Ile 860	Pro	GAA Glu	GGG Gly	AGC Ser	GCT Ala 865	Phe	TTC Phe	ATT Ile	CTT Leu	AGC Ser 870	AAG Lys	ACC Thr	3126
AAC Asn	CCG Pro	Ile 875	Arg	GTA Val	GGC Gly	TGC Cys	CAC His 880	AAG Lys	CTC Leu	ATC Ile	AAC Asn	CAC His 885	CAC His	ATC Ile	TTC Phe	3174
ACC Thr	AAC Asn 890	CTC Leu	ATC Ile	CTT	GTC Val	TTC Phe 895	ATC Ile	ATG Met	CTG Leu	AGC Ser	AGT Ser 900	GCT Ala	GCC Ala	CTG Leu	GCC Ala	3222
GCA Ala 905	GAG Glu	GAC Asp	CCC Pro	ATC Ile	CGC Arg 910	AGC Ser	CAC His	TCC Ser	TTC Phe	CGG Arg 915	AAC Asn	ACG Thr	ATA Ile	CTG Leu	GGT Gly 920	3270
TAC Tyr	TTT Phe	GAC Asp	TAT Tyr	GCC Ala 925	TTC Phe	ACA Thr	GCC Ala	ATC Ile	TTT Phe 930	ACT Thr	GTT Val	GAG Glu	ATC Ile	CTG Leu 935	TTG Leu	3318
AAG Lys	ATG Met	ACA Thr	ACT Thr 940	TTT Phe	GGA Gly	GCT Ala	TTC Phe	CTC Leu 945	CAC His	AAA Lys	GGG Gly	GCC Ala	TTC Phe 950	TGC Cys	AGG Arg	3366
AAC Asn	TAC Tyr	TTC Phe 955	AAT Asn	TTG Leu	CTG Leu	GAT Asp	ATG Met 960	CTG Leu	GTG Val	GTT Val	GGG Gly	GTG Val 965	TCT Ser	CTG Leu	GTG Val	3414
TCA Ser	TTT Phe 970	GGG Gly	ATT Ile	CAA Gln	TCC Ser	AGT Ser 975	GCC Ala	ATC Ile	TCC Ser	GTT Val	GTG Val 980	AAG Lys	ATT Ile	CTG Leu	AGG Arg	3462
GTC Val 985	TTA Leu	AGG Arg	GTC Val	CTG Leu	CGT Arg 990	CCC Pro	CTC Leu	AGG Arg	GCC Ala	ATC Ile 995	AAC Asn	AGA Arg	GCA Ala	Lys	GGA Gly 1000	3510
CTT Leu	AAG Lys	CAC His	GTG Val	GTC Val 1005	CAG Gln	TGC Cys	GTC Val	TTC Phe	GTG Val 1010	Ala	ATC Ile	CGG Arg	ACC Thr	ATC Ile 1015	Gly	3558
AAC Asn	ATC Ile	Met	ATC Ile 1020	Val	ACC Thr	ACC Thr	Leu	CTG Leu 1025	Gln	TTC Phe	ATG Met	Phe	GCC Ala 1030	Cys	ATC Ile	3606

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GGG Gly	GTC Val	CAG Gln 1035	Leu	TTC Phe	AAG Lys	GGG Gly	AAG Lys 1040	Phe	TAT Tyr	CGC Arg	TGT Cys	ACG Thr 1045	Asp	GAA Glu	GCC Ala	3654
		AAC Asn					Arg					Leu				3702
GGG Gly 106	Asp	GTT Val	GAC Asp	AGT Ser	CCT Pro 1070	Val	GTC Val	CGT Arg	GAA Glu	CGG Arg 1075	Ile	TGG Trp	CAA Gln	AAC Asn	AGT Ser 1080	3750
		AAC Asn			Asn					Met					Thr	3798
		ACG Thr		Glu					Leu					Ile		3846
		GGA Gly 1115	Glu					Ile					Val			3894
		TTC Phe					Ile					Phe				3942
	Ile	TTT Phe				Val					Gln					3990
AAA Lys	GAG Glu	TAT Tyr	AAG Lys	AAC Asn 116	Cys	GAG Glu	CTG Leu	GAC Asp	AAA Lys 117	Asn	CAG Gln	CGT Arg	CAG Gln	TGT Cys 117	Val	4038
GAA Glu	TAC Tyr	GCC Ala	TTG Leu 118	Lys	GCA Ala	CGT Arg	CCC Pro	TTG Leu 118	Arg	AGA Arg	TAC Tyr	ATC Ile	CCC Pro 1190	Lys	AAC Asn	4086
CCC Pro	TAC Tyr	CAG Gln 119	Tyr	AAG Lys	TTC Phe	TGG Trp	TAC Tyr 120	Val	GTG Val	AAC Asn	TCT Ser	TCG Ser 120	Pro	TTC Phe	GAA Glu	4134
TAC Tyr	ATG Met 121	ATG Met	TTT Phe	GTC Val	CTC Leu	ATC Ile 121	Met	CTC Leu	AAC Asn	ACA Thr	CTC Leu 122	Cys	TTG Leu	GCC Ala	ATG Met	4182
CAG Gln 122	His	TAC Tyr	GAG Glu	CAG Gln	TCC Ser 123	Lys	ATG Met	TTC Phe	AAT Asn	GAT Asp 123	Ala	ATG Met	GAC Asp	ATT Ile	CTG Leu 1240	4230
AAC Asn	ATG Met	GTC Val	TTC Phe	ACC Thr 124	Gly	GTG Val	TTC Phe	ACC Thr	GTC Val 125	Glu	ATG Met	GTT Val	TTG Leu	AAA Lys 125	Val	4278

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ATC Ile	GCA Ala	A TTI	AAG Lys 126	PIC	AAG Lys	GGG Gly	TAT	TTT Phe 126	Ser	GAC Asp	GCC Ala	TGG Trp	AAC Asn 127	Thi	TTT Phe	432
GAC Asp	Ser	CTC Leu 127	TTE	GTA Val	ATC Ile	GGC	AGC Ser 128	Ile	ATA Ile	GAC Asp	GTG Val	GCC Ala 128	Leu	AGC Ser	GAA Glu	4374
GCA Ala	GAC Asp 129	PIO	ACT Thr	GAA Glu	AGT Ser	GAA Glu 129	Asn	GTC Val	CCT	GTC Val	CCA Pro 130	Thr	GCT Ala	ACA Thr	CCT	4422
GGG Gly 130	MSII	TCT Ser	GAA Glu	GAG Glu	AGC Ser 131	Asn	AGA Arg	ATC Ile	TCC Ser	ATC Ile 131	Thr	TTT Phe	TTC Phe	CGT Arg	CTT Leu 1320	4470
TTC Phe	CGA Arg	GTG Val	ATG Met	CGA Arg 132	TTG Leu 5	GTG Val	AAG Lys	CTT Leu	CTC Leu 133	Ser	AGG Arg	GGG Gly	GAA Glu	GGC Gly 133	Ile	4518
CGG Arg	ACA Thr	TTG Leu	CTG Leu 134	TTP	ACT Thr	TTT Phe	ATT Ile	AAG Lys 134!	Phe	TTT Phe	CAG Gln	GCG Ala	CTC Leu 135	Pro	TAT Tyr	4566
GTG Val	GCC Ala	CTC Leu 135	ьeп	ATA Ile	GCC Ala	ATG Met	CTG Leu 1360	Phe	TTC Phe	ATC Ile	TAT Tyr	GCG Ala 1365	Val	ATT Ile	GGC Gly	4614
ATG Met	CAG Gln 137	Mec	TTT Phe	GGG Gly	AAA Lys	GTT Val 1375	Ala	ATG Met	AGA Arg	GAT Asp	AAC Asn 1380	Asn	CAG Gln	ATC Ile	AAT Asn	4662
AGG Arg 1385	MSII	AAT Asn	AAC Asn	TTC Phe	CAG Gln 1390	Thr	TTT Phe	CCC Pro	CAG Gln	GCG Ala 1395	Val	CTG Leu	CTG Leu	CTC Leu	TTC Phe 1400	4710
AGG Arg	TGT Cys	GCA Ala	ACA Thr	GGT Gly 1405	GAG Glu	GCC Ala	TGG Trp	CAG Gln	GAG Glu 1410	Ile	ATG Met	CTG Leu	GCC Ala	TGT Cys 1415	Leu	4758
CCA Pro	GGG Gly	AAG Lys	CTC Leu 1420	Cys	GAC Asp	CCT Pro	Glu	TCA Ser 1425	Asp	TAC Tyr	AAC Asn	CCC Pro	GGG Gly 1430	Glu	GAG Glu	4806
CAT His	ACA Thr	TGT Cys 1435	GIA	AGC Ser	AAC Asn	Phe .	GCC Ala 1440	Ile	GTC Val	TAT Tyr	TTC Phe	ATC Ile 1445	Ser	TTT Phe	TAC Tyr	4854
ATG Met	CTC Leu 1450	Cys	GCA Ala	TTT Phe	Leu	ATC I Ile 1455	ATC .	AAT Asn	CTG Leu	Phe	GTG Val 1460	Ala	GTC Val	ATC Ile	ATG Met	4902
GAT Asp 1465	Asn	TTC Phe	GAC Asp	Tyr	CTG . Leu ' 1470	ACC (Thr)	CGG (Arg)	GAC ' Asp '	\mathtt{Trp}	TCT Ser 1475	ATT Ile	TTG Leu	GGG Gly	CCT Pro	CAC His 1480	4950

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CAT TTA GAT GAA His Leu Asp Glu	TTC AAA AGA A Phe Lys Arg I 1485	ATA TGG TCA GAA Ile Trp Ser Glu 1490	TAT GAC CCT GAG Tyr Asp Pro Glu 149!	Ala
AAG GGA AGG ATA Lys Gly Arg Ile 1500	Lys His Leu A	GAT GTG GTC ACT Asp Val Val Thr 1505	CTG CTT CGA CGC Leu Leu Arg Arg 1510	ATC 5046 Ile
CAG CCT CCC CTG Gln Pro Pro Leu 1515	Gly Phe Gly I	AAG TTA TGT CCA Lys Leu Cys Pro 1520	CAC AGG GTA GCG His Arg Val Ala 1525	TGC 5094 Cys
AAG AGA TTA GTT Lys Arg Leu Val 1530	GCC ATG AAC A Ala Met Asn M 1535	ATG CCT CTC AAC Met Pro Leu Asn	AGT GAC GGG ACA Ser Asp Gly Thr 1540	GTC 5142 Val
ATG TTT AAT GCA Met Phe Asn Ala 1545	ACC CTG TTT G Thr Leu Phe A 1550	GCT TTG GTT CGA Ala Leu Val Arg 1555	Thr Ala Leu Lys	ATC 5190 Ile 1560
AAG ACC GAA GGG Lys Thr Glu Gly	AAC CTG GAG C Asn Leu Glu G 1565	CAA GCT AAT GAA Gln Ala Asn Glu 1570	GAA CTT CGG GCT Glu Leu Arg Ala 157	Val
ATA AAG AAA ATT Ile Lys Lys Ile 158	Trp Lys Lys T	ACC AGC ATG AAA Thr Ser Met Lys 1585	TTA CTT GAC CAA Leu Leu Asp Gln 1590	GTT 5286 Val
GTC CCT CCA GCT Val Pro Pro Ala 1595	Gly Asp Asp G	GAG GTA ACC GTG Glu Val Thr Val 1600	GGG AAG TTC TAT Gly Lys Phe Tyr 1605	GCC 5334 Ala
ACT TTC CTG ATA Thr Phe Leu Ile 1610	CAG GAC TAC T Gln Asp Tyr I 1615	TTT AGG AAA TTC Phe Arg Lys Phe	AAG AAA CGG AAA Lys Lys Arg Lys 1620	GAA 5382 Glu
CAA GGA CTG GTG Gln Gly Leu Val 1625	GGA AAG TAC (Gly Lys Tyr 1 1630	CCT GCG AAG AAC Pro Ala Lys Asn 1635	ACC ACA ATT GCC Thr Thr Ile Ala	CTA 5430 Leu 1640
CAG GCG GGA TTA Gln Ala Gly Leu	AGG ACA CTG (Arg Thr Leu I 1645	CAT GAC ATT GGG His Asp Ile Gly 1650	CCA GAA ATC CGG Pro Glu Ile Arg 165	Arg
GCT ATA TCG TGT Ala Ile Ser Cys 166	Asp Leu Gln	GAT GAC GAG CCT Asp Asp Glu Pro 1665	GAG GAA ACA AAA Glu Glu Thr Lys 1670	CGA 5526 Arg
GAA GAA GAA GAT Glu Glu Glu Asp 1675	Asp Val Phe	AAA AGA AAT GGT Lys Arg Asn Gly 1680	GCC CTG CTT GGA Ala Leu Leu Gly 1685	AAC 5574 Asn
CAT GTC AAT CAT His Val Asn His 1690	GTT AAT AGT (Val Asn Ser 1	Asp Arg Arg Asp	TCC CTT CAG CAG Ser Leu Gln Gln 1700	ACC 5622 Thr

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1705	171	Leu His Va 0	I Gln Arg Pi 1715	CT TCA ATT CC	Pro 1720
rad ber Asp	1725	Pro Leu Ph	e Pro Pro Al 1730	A GGA AAT TCC a Gly Asn Ser 173	Val 15
TGT CAT AAC Cys His Asn	CAT CAT AAC His His Asn 1740	CAT AAT TC His Asn Se 17	тте стугу	G CAA GTT CCC s Gln Val Pro 1750	ACC 5766 Thr
TCA ACA AAT Ser Thr Asn 1755	THE WALL THE	AAT AAT GC Asn Asn Ala 1760	C AAT ATG TC a Asn Met Se	C AAA GCT GCC r Lys Ala Ala 1765	CAT 5814 His
GGA AAG CGG Gly Lys Arg 1770	CCC AGC ATT Pro Ser Ile	GGG AAC CT Gly Asn Let 1775	F GAG CAT GT 1 Glu His Va 17	G TCT GAA AAT l Ser Glu Asn 80	GGG 5862 Gly
CAT CAT TCT His His Ser 1785	TCC CAC AAG Ser His Lys 1790	HIS ASP ATO	G GAG CCT CAG Glu Pro Gl: 1795	G AGA AGG TCC n Arg Arg Ser	AGT 5910 Ser 1800
GTG AAA AGA Val Lys Arg	ACC CGC TAT Thr Arg Tyr 1805	TAT GAA ACT Tyr Glu Thi	TAC ATT AGG Tyr Ile Arg 1810	G TCC GAC TCA G Ser Asp Ser 181:	Gly
TIP OLG CIN I	CTC CCA ACT Leu Pro Thr 1820	ATT TGC CGG Ile Cys Arg 182	Glu Asp Pro	A GAG ATA CAT O Glu Ile His 1830	GGC 6006
TAT TTC AGG (Tyr Phe Arg / 1835	GAC CCC CAC Asp Pro His	TGC TTG GGG Cys Leu Gly 1840	GAG CAG GAG Glu Gln Glu	TAT TTC AGT Tyr Phe Ser 1845	AGT 6054 Ser
GAG GAA TGC T Glu Glu Cys T 1850	lyr Giu Asp	GAC AGC TCG Asp Ser Ser 1855	CCC ACC TGG Pro Thr Trp 186	Ser Arg Gln	AAC 6102 Asn
TAT GGC TAC T Tyr Gly Tyr T 1865	TAC AGC AGA T Tyr Ser Arg 1870	TAC CCA GGC Tyr Pro Gly	AGA AAC ATO Arg Asn Ile 1875	GAC TCT GAG Asp Ser Glu	AGG 6150 Arg 1880
CCC CGA GGC T Pro Arg Gly T	AC CAT CAT (Yr His His H 1885	CCC CAA GGA Pro Gln Gly	TTC TTG GAG Phe Leu Glu 1890	GAC GAT GAC Asp Asp Asp 1895	Ser
CCC GTT TGC T Pro Val Cys T	AT GAT TCA (Yr Asp Ser A	CGG AGA TCT Arg Arg Ser 1909	Pro Arg Arg	CGC CTA CTA Arg Leu Leu 1910	CCT 6246 Pro
CCC ACC CCA G Pro Thr Pro A 1915	CA TCC CAC C la Ser His A	CGG AGA TCC arg Arg Ser 1920	TCC TTC AAC Ser Phe Asn	TTT GAG TGC Phe Glu Cys 1925	CTG 6294 Leu

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CGC CGG CAG Arg Arg Gln 1930	AGC AGC CAG Ser Ser Gln	GAA GAG GI Glu Glu Va 1935	al Pro Ser	TCT CCC ATC Ser Pro Ile 1940	TTC CCC Phe Pro	6342
CAT CGC ACG His Arg Thr 1945	GCC CTG CCT Ala Leu Pro 195	Leu His Le	TA ATG CAG eu Met Gln 1955	Gln Gln Ile	ATG GCA Met Ala 1960	6390
GTT GCC GGC Val Ala Gly	CTA GAT TCA Leu Asp Ser 1965	AGT AAA GO Ser Lys Al	CC CAG AAG la Gln Lys 1970	TAC TCA CCG Tyr Ser Pro	AGT CAC Ser His 1975	6438
TCG ACC CGG Ser Thr Arg	TCG TGG GCC Ser Trp Ala 1980	Thr Pro Pr	CA GCA ACC ro Ala Thr 985	CCT CCC TAC Pro Pro Tyr 1990	Arg Asp	6486
TGG ACA CCG Trp Thr Pro 199	TGC TAC ACC Cys Tyr Thr 5	CCC CTG AT Pro Leu Il 2000	TC CAA GTG le Gln Val	GAG CAG TCA Glu Gln Ser 2005	GAG GCC Glu Ala	6534
CTG GAC CAG Leu Asp Gln 2010	GTG AAC GGC Val Asn Gly	AGC CTG CC Ser Leu Pr 2015	ro Ser Leu	CAC CGC AGC His Arg Ser 2020	TCC TGG Ser Trp	6582
TAC ACA GAC Tyr Thr Asp 2025	GAG CCC GAC Glu Pro Asp 203	Ile Ser Ty	AC CGG ACT yr Arg Thr 2035	Phe Thr Pro	GCC AGC Ala Ser 2040	6630
CTG ACT GTC Leu Thr Val	CCC AGC AGC Pro Ser Ser 2045	TTC CGG AF	AC AAA AAC sn Lys Asn 2050	AGC GAC AAG Ser Asp Lys	CAG AGG Gln Arg 2055	6678
AGT GCG GAC Ser Ala Asp	AGC TTG GTG Ser Leu Val 2060	Glu Ala Va	TC CTG ATA al Leu Ile 065	TCC GAA GGC Ser Glu Gly 207	Leu Gly	6726
CGC TAT GCA Arg Tyr Ala 207	AGG GAC CCA Arg Asp Pro 5	AAA TTT GT Lys Phe Va 2080	TG TCA GCA al Ser Ala	ACA AAA CAC Thr Lys His 2085	GAA ATC Glu Ile	6774
GCT GAT GCC Ala Asp Ala 2090	TGT GAC CTC Cys Asp Leu	ACC ATC GATT Thr Ile As	AC GAG ATG sp Glu Met	GAG AGT GCA Glu Ser Ala 2100	GCC AGC Ala Ser	6822
ACC CTG CTT Thr Leu Leu 2105	AAT GGG AAC Asn Gly Asr 211	Val Arg P	CC CGA GCC ro Arg Ala 2115	Asn Gly Asp	GTG GGC Val Gly 2120	6870
CCC CTC TCA Pro Leu Ser	CAC CGG CAC His Arg Glr 2125	GAC TAT GA Asp Tyr G	AG CTA CAG lu Leu Gln 2130	GAC TTT GGT Asp Phe Gly	CCT GGC Pro Gly 2135	6918
TAC AGC GAC Tyr Ser Asp	GAA GAG CCA Glu Glu Pro 2140	Asp Pro G	GGG AGG GAT Sly Arg Asp 145	GAG GAG GAC Glu Glu Asp 215	Leu Ala	6966

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Asp Glu Met Ile Cys Ile Thr Thr Leu 2155 2160	7013
ACTGGCTCTG GCCTCAGGTG GGGCGCAGGA GAGCCAGGGG AAAAGTGCCT CATAGTTAGG	7073
AAAGTTTAGG CACTAGTTGG GAGTAATATT CAATTAATTA GACTTTTGTA TAAGAGATGT	7133
CATGCCTCAA GAAAGCCATA AACCTGGTAG GAACAGGTCC CAAGCGGTTG AGCCTGGCAG	7193
AGTACCATGC GCTCGGCCCC AGCTGCAGGA AACAGCAGGC CCCGCCCTCT CACAGAGGAT	7253
GGGTGAGGAG GCCAGACCTG CCCTGCCCCA TTGTCCAGAT GGGCACTGCT GTGGAGTCTG	7313
CTTCTCCCAT GTACCAGGGC ACCAGGCCCA CCCAACTGAA GGCATGGCGG CGGGGTGCAG	7373
GGGAAAGTTA AAGGTGATGA CGATCATCAC ACCTGTGTCG TTACCTCAGC CATCGGTCTA	7433
GCATATCAGT CACTGGGCCC AACATATCCA TTTTTAAACC CTTTCCCCCA AATACACTGC	7493
GTCCTGGTTC CTGTTTAGCT GTTCTGAAAT ACGGTGTGTA AGTAAGTCAG AACCCAGCTA	7553
CCAGTGATTA TTGCGAGGGC AATGGGACCT CATAAATAAG GTTTTCTGTG ATGTGACGCC	7613
AGTTTACATA AGAGAATATC AC	7635
(2) INFORMATION FOR SEQ ID NO:2:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 104 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA (genomic)	
(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1102	
<pre>(ix) FEATURE: (A) NAME/KEY: misc_feature (B) LOCATION: 1104 (D) OTHER INFORMATION: /note= "A 104-nucleotide alternative exon of alpha-1D."</pre>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:	
GTA AAT GAT GCG ATA GGA TGG GAA TGG CCA TGG GTG TAT TTT GTT AGT Val Asn Asp Ala Ile Gly Trp Glu Trp Pro Trp Val Tyr Phe Val Ser 1 5 10 15	48
CTG ATC ATC CTT GGC TCA TTT TTC GTC CTT AAC CTG GTT CTT GGT GTC Leu Ile Ile Leu Gly Ser Phe Phe Val Leu Asn Leu Val Leu Gly Val	96

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Leu	Ser	GG														10.
(2)	INFO	ORMAT	NOI	FOR	SEQ	ID 1	10:3	:								
	(i)	(1	A) LI 3) T C) S	engti YPE : IRANI	HARAC H: 69 nucl DEDNI DGY:	575 l Leic ESS:	ase acio sino	pai:	rs							
	(ii)	MOI	LECUI	LE T	YPE:	DNA	(ger	omic	2)							
	(ix)		A) N	AME/I	KEY:		5492									
	(xi)	SEÇ	QUENC	CE DI	ESCR	PTIC	ON: 8	SEQ I	ID NO	0:3:						
		AAT Asn														4.8
		AAC Asn														90
		GCA Ala 35														144
		TCG Ser														192
ATG Met 65	GGC Gly	AGC Ser	GCT Ala	GGC Gly	AAT Asn 70	GCG Ala	ACC Thr	ATC Ile	TCC Ser	ACA Thr 75	GTC Val	AGC Ser	TCC Ser	ACG Thr	CAG Gln 80	240
CGG Arg	AAG Lys	CGC Arg	CAG Gln	CAA Gln 85	TAT Tyr	GGG Gly	AAA Lys	CCC Pro	AAG Lys 90	AAG Lys	CAG Gln	GGC	AGC Ser	ACC Thr 95	ACG Thr	28
GCC Ala	ACA Thr	CGC Arg	CCG Pro 100	CCC Pro	CGA Arg	GCC Ala	Leu	CTC Leu 105	TGC Cys	CTG Leu	ACC Thr	CTG Leu	AAG Lys 110	AAC Asn	CCC Pro	33
ATC Ile	CGG Arg	AGG Arg 115	GCC Ala	TGC Cys	ATC Ile	AGC Ser	ATT Ile 120	GTC Val	GAA Glu	TGG Trp	AAA Lys	CCA Pro 125	TTT Phe	GAA Glu	ATA Ile	384
ATT Ile	ATT Ile 130	TTA Leu	CTG Leu	ACT Thr	ATT Ile	TTT Phe 135	GCC Ala	AAT Asn	TGT Cys	GTG Val	GCC Ala 140	TTA Leu	GCG Ala	ATC Ile	TAT Tyr	43:

ATT CCC TTT CCA GAA GAT GAT TCC AAC GCC ACC AAT TCC AAC CTG GAA

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Ile 145	Pro	Phe	Pro	Glu	Asp 150	Asp	Ser	Asr	Ala	Thr 155	Asr	ı Ser	Asn	Leu	Glu 160	
CGA Arg	GTG Val	GAA Glu	TAT Tyr	Leu 165	Phe	CTC Leu	ATA	ATI	TTT Phe 170	Thr	GTO Val	GAA Glu	GCG Ala	TTT Phe 175	TTA Leu	528
AAA Lys	GTA Val	ATC	GCC Ala 180	Tyr	GGA Gly	CTC Leu	CTC Leu	TTI Phe 185	His	Pro	AAT Asn	GCC Ala	TAC Tyr 190	Leu	CGC Arg	576
AAC Asn	GGC	TGG Trp 195	Asn	CTA Leu	CTA Leu	GAT Asp	TTT Phe 200	Ile	ATT	GTG Val	GTT Val	GTG Val 205	GGG Gly	CTT Leu	TTT Phe	624
AGT Ser	GCA Ala 210	тте	TTA Leu	GAA Glu	CAA Gln	GCA Ala 215	ACC Thr	AAA Lys	GCA Ala	GAT Asp	GGG Gly 220	GCA Ala	AAC Asn	GCT Ala	CTC Leu	672
GGA Gly 225	GGG	AAA Lys	GGG Gly	GCC Ala	GGA Gly 230	TTT Phe	GAT Asp	GTG Val	AAG Lys	GCG Ala 235	CTG Leu	AGG Arg	GCC Ala	TTC Phe	CGC Arg 240	720
GTG Val	CTG Leu	CGC Arg	CCC Pro	CTG Leu 245	CGG Arg	CTG Leu	GTG Val	TCC Ser	GGA Gly 250	GTC Val	CCA Pro	AGT Ser	CTC Leu	CAG Gln 255	GTG Val	768
GTC Val	CTG Leu	AAT Asn	TCC Ser 260	ATC Ile	ATC Ile	AAG Lys	GCC Ala	ATG Met 265	GTC Val	CCC Pro	CTG Leu	CTG Leu	CAC His 270	ATC Ile	GCC Ala	816
CTG Leu	CTT Leu	GTG Val 275	CTG Leu	TTT Phe	GTC Val	ATC Ile	ATC Ile 280	ATC Ile	TAC Tyr	GCC Ala	ATC Ile	ATC Ile 285	GGC Gly	TTG Leu	GAG Glu	864
CTC Leu	TTC Phe 290	ATG Met	GGG Gly	AAG Lys	ATG Met	CAC His 295	AAG Lys	ACC Thr	TGC Cys	TAC Tyr	AAC Asn 300	CAG Gln	GAG Glu	GGC Gly	ATA Ile	912 "
GCA Ala 305	GAT Asp	GTT Val	CCA Pro	GCA Ala	GAA Glu 310	GAT Asp	GAC Asp	CCT Pro	TCC Ser	CCT Pro 315	TGT Cys	GCG Ala	CTG Leu	GAA Glu	ACG Thr 320	960
GGC Gly	CAC His	GGG Gly	CGG Arg	CAG Gln 325	TGC Cys	CAG Gln	AAC Asn	GGC Gly	ACG Thr 330	GTG Val	TGC Cys	AAG Lys	CCC Pro	GGC Gly 335	TGG Trp	1008
GAT Asp	GGT Gly	CCC Pro	AAG Lys 340	CAC His	GGC Gly	ATC Ile	ACC Thr	AAC Asn 345	TTT Phe	GAC Asp	AAC Asn	TTT Phe	GCC Ala 350	TTC Phe	GCC Ala	1056
ATG Met	CTC Leu	ACG Thr 355	GTG Val	TTC Phe	CAG Gln	TGC Cys	ATC Ile 360	ACC Thr	ATG Met	GAG Glu	GGC Gly	TGG Trp 365	ACG Thr	GAC Asp	GTG Val	1104
CTG	TAC	TGG	GTC	TAA	GAT	GCC	GTA	GGA	AGG	GAC	TGG	ccc	TGG	ATC	TAT	1152

Leu	Tyr 370	Trp	Val	Asn	Asp	Ala 375	Val	Gly	Arg	Asp	Trp 380	Pro	Trp	Ile	Tyr	
					ATC Ile 390											1200
					GGA Gly											1248
					CAG Gln											1296
					CTG Leu											1344
					GAA Glu											1392
					ATG Met 470											1440
					GAA Glu											1488
					GAA Glu											1536
					CTG Leu											1584
					TGG Trp											1632
					GTC Val 550											1680
CTC Leu	AAC Asn	ACG Thr	CTC Leu	ACC Thr 565	ATT Ile	GCC Ala	TCT Ser	GAG Glu	CAC His 570	TAC Tyr	AAC Asn	CAG Gln	CCC Pro	AAC Asn 575	TGG Trp	1728
					GAC Asp											1776
ACG	GCA	GAG	ATG	CTC	CTG	AAG	ATG	TAC	AGC	CTG	GGC	CTG	CAG	GCC	TAC	1824

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Thr	Ala	Glu 595	Met	Leu	Leu	Lys	Met 600	Tyr	Ser	Leu	Gly	Leu 605	Gln	Ala	Tyr	
TTC Phe	GTG Val 610	Ser	CTC Leu	TTC Phe	AAC Asn	CGC Arg 615	TTT Phe	GAC Asp	TGC Cys	TTC Phe	GTC Val 620	GTG Val	TGT Cys	GGC Gly	GGC Gly	1872
ATC Ile 625	Leu	GAG Glu	ACC Thr	ATC Ile	CTG Leu 630	GTG Val	GAG Glu	ACC Thr	AAG Lys	ATC Ile 635	ATG Met	TCC Ser	CCA Pro	CTG Leu	GGC Gly 640	1920
ATC Ile	TCC Ser	GTG Val	CTC Leu	AGA Arg 645	TGC Cys	GTC Val	CGG Arg	CTG Leu	CTG Leu 650	AGG Arg	ATT Ile	TTC Phe	AAG Lys	ATC Ile 655	ACG Thr	1968
AGG Arg	TAC Tyr	TGG Trp	AAC Asn 660	TCC Ser	TTG Leu	AGC Ser	AAC Asn	CTG Leu 665	GTG Val	GCA Ala	TCC Ser	TTG Leu	CTG Leu 670	AAC Asn	TCT Ser	2016
GTG Val	CGC Arg	TCC Ser 675	ATC Ile	GCC Ala	TCC Ser	CTG Leu	CTC Leu 680	CTT Leu	CTC Leu	CTC Leu	TTC Phe	CTC Leu 685	TTC Phe	ATC Ile	AŤC Ile	2064
ATC Ile	TTC Phe 690	TCC Ser	CTC Leu	CTG Leu	GGG Gly	ATG Met 695	CAG Gln	CTC Leu	TTT Phe	GGA Gly	GGA Gly 700	AAG Lys	TTC Phe	AAC Asn	TTT Phe	2112
GAT Asp 705	GAG Glu	ATG Met	CAG Gln	ACC Thr	CGG Arg 710	AGG Arg	AGC Ser	ACA Thr	TTC Phe	GAT Asp 715	AAC Asn	TTC Phe	CCC Pro	CAG Gln	TCC Ser 720	2160
CTC Leu	CTC Leu	ACT Thr	GTG Val	TTT Phe 725	CAG Gln	ATC Ile	CTG Leu	ACC Thr	GGG Gly 730	GAG Glu	GAC Asp	TGG Trp	AAT Asn	TCG Ser 735	GTG Val	2208
ATG Met	TAT Tyr	GAT Asp	GGG Gly 740	ATC Ile	ATG Met	GCT Ala	TAT Tyr	GGG Gly 745	GGC Gly	CCC Pro	TCT Ser	TTT Phe	CCA Pro 750	GGG Gly	ATG Met	2256
TTA Leu	GTC Val	TGT Cys 755	ATT Ile	TAC Tyr	TTC Phe	ATC Ile	ATC Ile 760	CTC Leu	TTC Phe	ATC Ile	TGT Cys	GGA Gly 765	AAC Asn	TAT Tyr	ATC Ile	2304
CTA Leu	CTG Leu 770	AAT Asn	GTG Val	TTC Phe	TTG Leu	GCC Ala 775	ATT Ile	GCT Ala	GTG Val	GAC Asp	AAC Asn 780	CTG Leu	GCT Ala	GAT Asp	GCT Ala	2352
GAG Glu 785	AGC Ser	CTC Leu	ACA Thr	TCT Ser	GCC Ala 790	CAA Gln	AAG Lys	GAG Glu	GAG Glu	GAA Glu 795	GAG Glu	GAG Glu	AAG Lys	GAG Glu	AGA Arg 800	2400
AAG Lys	AAG Lys	CTG Leu	GCC Ala	AGG Arg 805	ACT Thr	GCC Ala	AGC Ser	CCA Pro	GAG Glu 810	AAG Lys	AAA Lys	CAA Gln	GAG Glu	TTG Leu 815	GTG Val	2448
GAG	AAG	CCG	GCA	GTG	GGG	GAA	TCC	AAG	GAG	GAG	AAG	ATT	GAG	CTG	AAA	2496

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Glu	Lys	Pro	Ala 820	Val	Gly	Glu	Ser	Lys 825	Glu	Glu	Lys	Ile	Glu 830	Leu	Lys	
TCC Ser	ATC Ile	ACG Thr 835	GCT Ala	GAC Asp	GGA Gly	GAG Glu	TCT Ser 840	CCA Pro	CCC Pro	GCC Ala	ACC Thr	AAG Lys 845	ATC Ile	AAC Asn	ATG Met	2544
GAT Asp	GAC Asp 850	CTC Leu	CAG Gln	CCC Pro	AAT Asn	GAA Glu 855	AAT Asn	GAG Glu	GAT Asp	AAG Lys	AGC Ser 860	CCC Pro	TAC Tyr	CCC Pro	AAC Asn	2592
CCA Pro 865	GAA Glu	ACT Thr	ACA Thr	GGA Gly	GAA Glu 870	GAG Glu	GAT Asp	GAG Glu	GAG Glu	GAG Glu 875	CCA Pro	GAG Glu	ATG Met	CCT Pro	GTC Val 880	2640
					CCA Pro											2688
					GCC Ala											2736
					TGC Cys											-2784
					TTC Phe											2832
					CAC His 950											2880
					ACC Thr											2928
					GCT Ala											2976
					GAC Asp			Val					Leu			3024
		Ile			AGT Ser		Ile					Ile				3072
	Arg				CCC Pro 1030	Leu					Arg					3120
AAG	CAT	GTG	GTT	CAG	TGT	GTG	TTT	GTC	GCC	ATC	CGG	ACC	ATC	GGG	AAC	3168

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Lys	His	Val	Val	Gln 104	Cys 5	Val	Phe	Val	Ala 105		Arg	Thr	Ile	Gl _y 105	Asn 5	
ATC Ile	GTG Val	ATT	GTC Val 106	Thr	ACC Thr	CTG Leu	CTG Leu	CAG Gln 106	Phe	ATG Met	TTT Phe	GCC Ala	TGC Cys 107	Ile	GGG	3216
GTC Val	CAG Gln	CTC Leu 107	Phe	AAG Lys	GGA Gly	AAG Lys	CTG Leu 108	Tyr	ACC	TGT Cys	TCA Ser	GAC Asp 108	Ser	TCC	AAG Lys	3264
CAG Gln	ACA Thr 109	GIu	GCG Ala	GAA Glu	TGC Cys	AAG Lys 109	Gly	AAC Asn	TAC Tyr	ATC Ile	ACG Thr	Tyr	AAA Lys	GAC Asp	GGG GJy	3312
GAG Glu 110	GTT Val 5	GAC Asp	CAC His	CCC	ATC Ile 111	Ile	CAA Gln	CCC Pro	CGC Arg	AGC Ser 111	Trp	GAG Glu	AAC Asn	AGC Ser	AAG Lys 1120	3360
TTT	GAC Asp	TTT Phe	GAC Asp	AAT Asn 1125	Val	CTG Leu	GCA Ala	GCC Ala	ATG Met 113	Met	GCC Ala	CTC Leu	TTC Phe	ACC Thr 113	Val	3408
TCC Ser	ACC Thr	TTC Phe	GAA Glu 114	Gly	TGG Trp	CCA Pro	GAG Glu	CTG Leu 114	Leu	TAC Tyr	CGC Arg	TCC Ser	ATC Ile 115	Asp	TCC Ser	3456
CAC His	ACG Thr	GAA Glu 115	Asp	AAG Lys	GGC Gly	CCC Pro	ATC Ile 1160	Tyr	AAC Asn	TAC Tyr	CGT Arg	GTG Val 1165	Glu	ATC Ile	TCC Ser	3504
ATC Ile	TTC Phe 1170	Phe	ATC Ile	ATC Ile	TAC Tyr	ATC Ile 1175	Ile	ATC Ile	ATC Ile	GCC Ala	TTC Phe 1180	Phe	ATG Met	ATG Met	AAC Asn	3552
ATC Ile 1185	TTC Phe	GTG Val	GGC Gly	TTC Phe	GTC Val 1190	Ile	GTC Val	ACC Thr	TTT Phe	CAG Gln 1195	Glu	CAG Gln	GGG Gly	GAG Glu	CAG Gln 1200	3600
GAG Glu	TAC Tyr	AAG Lys	AAC Asn	TGT Cys 1205	Glu	CTG Leu	GAC Asp	AAG Lys	AAC Asn 1210	Gln	CGA Arg	CAG Gln	TGC Cys	GTG Val 1215	Glu	3648
TAC Tyr	GCC Ala	Leu	AAG Lys 1220	Ala	CGG Arg	CCC Pro	Leu	CGG Arg 1225	Arg	TAC Tyr	ATC Ile	CCC Pro	AAG Lys 1230	Asn	CAG Gln	3696
CAC His	CAG Gln	TAC Tyr 1235	Lys	GTG Val	TGG Trp	Tyr	GTG Val 1240	Val	AAC Asn	TCC Ser	Thr	TAC Tyr 1245	Phe	GAG Glu	TAC Tyr	3744
CTG Leu	ATG Met 1250	Phe	GTC Val	CTC . Leu	Ile	CTG Leu 1255	Leu	AAC Asn	ACC Thr	Ile	TGC Cys 1260	Leu	GCC Ala	ATG Met	CAG Gln	3792
CAC	TAC	GGC	CAG .	AGC '	TGC	CTG	TTC .	AAA	ATC	GCC	ATG .	AAC	ATC	CTC	AAC	3840

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His 1265		Gly	Gln	Ser	Cys 1270		Phe	Lys	Ile	Ala 1275		Asn	Ile	Leu	Asn 1280	
ATG Met	CTC Leu	TTC Phe	ACT Thr	GGC Gly 1285	Leu	TTC Phe	ACC Thr	GTG Val	GAG Glu 1290	ATG Met	ATC Ile	CTG Leu	AAG Lys	CTC Leu 1295	Ile	3888
GCC Ala	TTC Phe	AAA Lys	CCC Pro 1300	Lys	GGT Gly	TAC Tyr	TTT Phe	AGT Ser 1305	Asp	CCC Pro	TGG Trp	AAT Asn	GTT Val 1310	Phe	GAC Asp	3936
TTC Phe	CTC Leu	ATC Ile 1315	Val	ATT Ile	GGC Gly	AGC Ser	ATA Ile 1320	Ile	GAC Asp	GTC Val	ATT Ile	CTC Leu 1325	Ser	GAG Glu	ACT Thr	3984
AAT Asn	CCA Pro 1330	Ala	GAA Glu	CAT His	ACC Thr	CAA Gln 1335	Cys	TCT Ser	CCC Pro	TCT Ser	ATG Met 1340	Asn	GCA Ala	GAG Glu	GAA Glu	4032
AAC Asn 1345	Ser	CGC Arg	ATC Ile	TCC Ser	ATC Ile 1350	Thr	TTC Phe	TTC Phe	CGC Arg	CTG Leu 1355	Phe	CGG Arg	GTC Val	ATG Met	CGT Arg 1360	4080
CTG Leu	GTG Val	AAG Lys	CTG Leu	CTG Leu 1365	Ser	CGT Arg	GGG Gly	GAG Glu	GGC Gly 1370	ATC Ile	CGG Arg	ACG Thr	CTG Leu	CTG Leu 1375	Trp	4128
ACC Thr	TTC Phe	ATC Ile	AAG Lys 1380	Ser	TTC Phe	CAG Gln	GCC Ala	CTG Leu 1389	Pro	TAT Tyr	GTG Val	GCC Ala	CTC Leu 1390	Leu	ATC Ile	4176
GTG Val	ATG Met	CTG Leu 1395	Phe	TTC Phe	ATC Ile	TAC Tyr	GCG Ala 1400	Val	ATC Ile	GGG Gly	ATG Met	CAG Gln 1405	Val	TTT Phe	GGG Gly	4224
AAA Lys	ATT Ile 1410	Ala	CTG Leu	AAT Asn	GAT Asp	ACC Thr 141	Thr	GAG Glu	ATC Ile	AAC Asn	CGG Arg 1420	Asn	AAC Asn	AAC Asn	TTT Phe	4272
CAG Gln 1425	Thr	TTC Phe	CCC Pro	CAG Gln	GCC Ala 143	Val	CTG Leu	CTC Leu	CTC Leu	TTC Phe 143	Arg	TGT Cys	GCC Ala	ACC Thr	GGG Gly 1440	4320
GAG Glu	GCC Ala	TGG Trp	CAG Gln	GAC Asp 144	Ile	ATG Met	CTG Leu	GCC Ala	TGC Cys 145	ATG Met O	CCA Pro	GGC Gly	AAG Lys	AAG Lys 145	Cys	4368
GCC Ala	CCA Pro	GAG Glu	TCC Ser 146	Glu	CCC Pro	AGC Ser	AAC Asn	AGC Ser 146	Thr	GAG Glu	GGT Gly	GAA Glu	ACA Thr 147	Pro	TGT Cys	4416
GGT Gly	AGC Ser	AGC Ser 147	Phe	GCT Ala	GTC Val	TTC Phe	TAC Tyr 148	Phe	ATC Ile	AGC Ser	TTC Phe	TAC Tyr 148	Met	CTC Leu	TGT Cys	4464
GCC	TTC	CTG	ATC	ATC	AAC	CTC	TTT	GTA	GCT	GTC	ATC	ATG	GAC	AAC	TTT	4512

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Ala Phe Leu Ile Ile Asn Leu Phe Val Ala Val Ile Met Asp Asn Phe 1490 1495 1500	
GAC TAC CTG ACA AGG GAC TGG TCC ATC CTT GGT CCC CAC CAC CTG GAT Asp Tyr Leu Thr Arg Asp Trp Ser Ile Leu Gly Pro His His Leu Asp 1505 1510 1520	4560
GAG TTT AAA AGA ATC TGG GCA GAG TAT GAC CCT GAA GCC AAG GGT CGT Glu Phe Lys Arg Ile Trp Ala Glu Tyr Asp Pro Glu Ala Lys Gly Arg 1525 1530 1535	4608
ATC AAA CAC CTG GAT GTG GTG ACC CTC CTC CGG CGG ATT CAG CCG CCA Ile Lys His Leu Asp Val Val Thr Leu Leu Arg Arg Ile Gln Pro Pro 1540 1545 1550	4656
CTA GGT TTT GGG AAG CTG TGC CCT CAC CGC GTG GCT TGC AAA CGC CTG Leu Gly Phe Gly Lys Leu Cys Pro His Arg Val Ala Cys Lys Arg Leu 1555 1560 1565	4704
GTC TCC ATG AAC ATG CCT CTG AAC AGC GAC GGG ACA GTC ATG TTC AAT Val Ser Met Asn Met Pro Leu Asn Ser Asp Gly Thr Val Met Phe Asn 1570 1580	4752
GCC ACC CTG TTT GCC CTG GTC AGG ACG GCC CTG AGG ATC AAA ACA GAA Ala Thr Leu Phe Ala Leu Val Arg Thr Ala Leu Arg Ile Lys Thr Glu 1585 1590 1595 1600	4800
GGG AAC CTA GAA CAA GCC AAT GAG GAG CTG CGG GCG ATC ATC AAG AAG Gly Asn Leu Glu Gln Ala Asn Glu Glu Leu Arg Ala Ile Ile Lys Lys 1605 1610 1615	4848
ATC TGG AAG CGG ACC AGC ATG AAG CTG CTG GAC CAG GTG GTG CCC CCT Ile Trp Lys Arg Thr Ser Met Lys Leu Leu Asp Gln Val Val Pro Pro 1620 1625 1630	4896
GCA GGT GAT GAG GTC ACC GTT GGC AAG TTC TAC GCC ACG TTC CTG Ala Gly Asp Asp Glu Val Thr Val Gly Lys Phe Tyr Ala Thr Phe Leu 1635 1640 1645	4944
ATC CAG GAG TAC TTC CGG AAG TTC AAG AAG CGC AAA GAG CAG GGC CTT Ile Gln Glu Tyr Phe Arg Lys Phe Lys Lys Arg Lys Glu Gln Gly Leu 1650 1660	4992
GTG GGC AAG CCC TCC CAG AGG AAC GCG CTG TCT CTG CAG GCT GGC TTG Val Gly Lys Pro Ser Gln Arg Asn Ala Leu Ser Leu Gln Ala Gly Leu 1665 1670 1680	5040
CGC ACA CTG CAT GAC ATC GGG CCT GAG ATC CGA CGG GCC ATC TCT GGA Arg Thr Leu His Asp Ile Gly Pro Glu Ile Arg Arg Ala Ile Ser Gly 1685 1690 1695	5088
GAT CTC ACC GCT GAG GAG GAG CTG GAC AAG GCC ATG AAG GAG GCT GTG Asp Leu Thr Ala Glu Glu Glu Leu Asp Lys Ala Met Lys Glu Ala Val 1700 1705 1710	5136
TCC GCT GCT TCT GAA GAT GAC ATC TTC AGG AGG GCC GGT GGC CTG TTC	5184

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Ser	Ala	Ala 1715		Glu	Asp	Asp	Ile 1720		Arg	Arg	Ala	Gly 1725		Leu	Phe	
GGC Gly	AAC Asn 1730	His	GTC Val	AGC Ser	TAC Tyr	TAC Tyr 1735	Gln	AGC Ser	GAC Asp	GGC Gly	CGG Arg 1740	Ser	GCC Ala	TTC Phe	CCC Pro	5232
	Thr				CAG Gln 1750	Arg					Asn					5280
AGC Ser	CAG Gln	GGC Gly	GAC Asp	ACT Thr 1765	GAG Glu	TCG Ser	CCA Pro	TCC Ser	CAC His 1770	Glu	AAG Lys	CTG Leu	GTG Val	GAC Asp 1775	Ser	5328
ACC Thr	TTC Phe	ACC Thr	CCG Pro 1780	Ser	AGC Ser	TAC Tyr	TCG Ser	TCC Ser 1785	Thr	GGC Gly	TCC Ser	AAC Asn	GCC Ala 1790	Asn	ATC Ile	5376
AAC Asn	AAC Asn	GCC Ala 1795	Asn	AAC Asn	ACC Thr	GCC Ala	CTG Leu 1800	Gly	CGC Arg	CTC Leu	CCT Pro	CGC Arg 1805	Pro	GCC Ala	GGC Gly	5424
TAC Tyr	CCC Pro 1810	Ser	ACA Thr	GTC Val	AGC Ser	ACT Thr 1815	Val	GAG Glu	GGC Gly	CAC His	GGG Gly 1820	Pro	CCC Pro	TTG Leu	TCC Ser	5472
CCT Pro 1825	Ala	ATC Ile	CGG Arg	GTG Val	CAG Gln 1830	Glu	GTG Val	GCG Ala	TGG Trp	AAG Lys 1835	Leu	AGC Ser	TCC Ser	AAC Asn	AGG Arg 1840	5520
TGC Cys	CAC His	TCC Ser	CGG Arg	GAG Glu 1845	AGC Ser	CAG Gln	GCA Ala	GCC Ala	ATG Met 1850	Ala	CGT Arg	CAG Gln	GAG Glu	GAG Glu 185	Thr	5568
TCT Ser	CAG Gln	GAT Asp	GAG Glu 1860	Thr	TAT Tyr	GAA Glu	GTG Val	AAG Lys 186	Met	AAC Asn	CAT His	GAC Asp	ACG Thr 1870	Glu	GCC Ala	5616
TGC Cys	AGT Ser	GAG Glu 1875	Pro	AGC Ser	CTG Leu	CTC Leu	TCC Ser 1880	Thr	GAG Glu	ATG Met	CTC Leu	TCC Ser 1885	Tyr	CAG Gln	GAT Asp	5664
GAC Asp	GAA Glu 1890	Asn	CGG Arg	CAA Gln	CTG Leu	ACG Thr 189	Leu	CCA Pro	GAG Glu	GAG Glu	GAC Asp 190	Lys	AGG Arg	GAC Asp	ATC Ile	5712
CGG Arg 190	Gln	TCT Ser	CCG Pro	AAG Lys	AGG Arg 191	Gly	TTC Phe	CTC Leu	CGC Arg	TCT Ser 191	Ala	TCA Ser	CTA Leu	GGT Gly	CGA Arg 1920	5760
AGG Arg	GCC Ala	TCC Ser	TTC Phe	CAC His 192	CTG Leu 5	GAA Glu	TGT Cys	CTG Leu	AAG Lys 193	Arg	CAG Gln	AAG Lys	GAC Asp	CGA Arg 193	Gly	5808
GGA	GAC	ATC	TCT	CAG	AAG	ACA	GTC	CTG	ccc	TTG	CAT	CTG	GTT	CAT	CAT	5856

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Gly	Asp	, Ile	Ser 194	Gln 0	Lys	Thr	Val	Leu 194	Pro 5	Leu	His	Leu	Val		His	
CAG Gln	GCA Ala	TTG Leu 195	Ala	GTG Val	GCA Ala	GGC	CTG Leu 196	Ser	CCC	CTC Leu	CTC	CAG Gln 196	Arg	AGC Ser	CAT His	5904
TCC Ser	CCT Pro 197	ALA	TCA Ser	TTC Phe	CCT Pro	AGG Arg 197	Pro	TTT Phe	GCC Ala	ACC Thr	CCA Pro 198	Pro	GCC Ala	ACA Thr	CCT Pro	5952
GGC Gly 198	ser	CGA Arg	GGC Gly	TGG Trp	CCC Pro 199	Pro	CAG Gln	CCC Pro	GTC Val	CCC Pro 199	Thr	CTG Leu	CGG Arg	CTT Leu	GAG Glu 2000	6000
GGG Gly	GTC Val	GAG Glu	TCC Ser	AGT Ser 200	Glu	AAA Lys	CTC Leu	AAC Asn	AGC Ser 201	Ser	TTC Phe	CCA Pro	TCC Ser	ATC Ile 201	His	6048
TGC Cys	GGC Gly	TCC Ser	TGG Trp 202	Ala	GAG Glu	ACC Thr	ACC Thr	CCC Pro 202	Gly	GGC Gly	GGG Gly	GGC Gly	AGC Ser 203	Ser	GCC Ala	6096
GCC Ala	CGG Arg	AGA Arg 203	Val	CGG Arg	CCC Pro	GTC Val	TCC Ser 2040	Leu	ATG Met	GTG Val	CCC Pro	AGC Ser 2045	Gln	GCT Ala	GGG Gly	6144
GCC Ala	CCA Pro 2050	GIY	AGG Arg	CAG Gln	TTC Phe	CAC His 2055	Gly	AGT Ser	GCC Ala	AGC Ser	AGC Ser 2060	CTG Leu	GTG Val	GAA Glu	GCG Ala	6192
GTC Val 2070	ьeп	ATT Ile	TCA Ser	GAA Glu	GGA Gly 2075	Leu	GGG Gly	CAG Gln	TTT Phe	GCT Ala 2080	Gln	GAT Asp	CCC Pro	AAG Lys	TTC Phe 2085	6240
ATC Ile	GAG Glu	GTC Val	ACC Thr	ACC Thr 2090	Gln	GAG Glu	CTG Leu	GCC Ala	GAC Asp 2095	Ala	TGC Cys	GAC Asp	ATG Met	ACC Thr 2100	Ile	6288
GAG Glu	GAG Glu	ATG Met	GAG Glu 2105	Ser	GCG Ala	GCC Ala	Asp	AAC Asn 2110	Ile	CTC Leu	AGC Ser	GGG Gly	GGC Gly 2115	Ala	CCA Pro	6336
CAG Gln	AGC Ser	CCC Pro 2120	Asn	GGC Gly	GCC Ala	Leu	TTA Leu 2125	Pro	TTT Phe	GTG Val	AAC Asn	TGC Cys 2130	Arg	GAC Asp	GCG Ala	6384
GGG Gly	CAG Gln 2135	Asp	CGA Arg	GCC Ala	GGG Gly	GGC Gly 2135	GAA Glu	GAG Glu	GAC Asp	Ala	GGC Gly 2140	TGT Cys	GTG Val	CGC Arg	GCG Ala	6432
CGG Arg 2145	GIA	CGA Arg	CCG Pro	Ser	GAG Glu 2150	Glu (GAG Glu	CTC Leu	Gln	GAC Asp 2155	Ser	AGG (GTC Val	TAC Tyr	GTC Val 2160	6480
AGC	AGC	CTG	TAGT	GGGC	GC T	GCCA	GATG	C GG	GCTT	TTTT	TTA	TTTG'	TTT	CAAT	GTTCCT	6539

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ser ser neu	
AATGGGTTCG TTTCAGAAGT GCCTCACTGT TCTCGT	6575
(2) INFORMATION FOR SEQ ID NO:4:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 133 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:	
AGACCACGGC TTCCTCGAAT CTTGCGCGAA GCCGCCGGCCA TCGGAGGAG GGATTAATCC	60
AGACCCGCCG GGGGGTGTTT TCACATTTCT TCCTCTTCGTG GCTGCTCCT CCTATTAAAA	120
CCATTTTGG TCC	133
(2) INFORMATION FOR SEQ ID NO:5:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 89 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:	
CGCTGAGGGC CTTCCGCGTG CTGCGCCCCC TGCGGCTGGT GTCCGGAGTC CCAAGTCTCC	60
AGGTGGTCCT GAATTCCATC ATCAAGGCC	89
(2) INFORMATION FOR SEQ ID NO:6:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 84 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
<pre>(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 184 (D) OTHER INFORMATION: /note= "An alternative exon of</pre>	

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:6:

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His Tyr Phe Cys Asp Ala Trp Asn Thr Phe Asp Ala Leu Ile Val Val 1 5 10 15	4
GGT AGC ATT GTT GAT ATA GCA ATC ACC GAG GTA AAC Gly Ser Ile Val Asp Ile Ala Ile Thr Glu Val Asn 20 25	8
(2) INFORMATION FOR SEQ ID NO:7:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 7362 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1447163	
(ix) FEATURE: (A) NAME/KEY: 5'UTR (B) LOCATION: 1143	
(ix) FEATURE: (A) NAME/KEY: 3'UTR (B) LOCATION: 71617362	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:	
GCGGCGGCGG CTGCGGCGGT GGGGCCGGGC GAGGTCCGTG CGGTCCCGGC GGCTCCGTGG	60
CTGCTCCGCT CTGAGCGCCT GCGCGCCCCG CGCCCTCCCT GCCGGGGCCG CTGGGCCGGG	120
GATGCACGCG GGGCCCGGGA GCC ATG GTC CGC TTC GGG GAC GAG CTG GGC Met Val Arg Phe Gly Asp Glu Leu Gly 1 5	170
GGC CGC TAT GGA GGC CCC GGC GGC GGA GAG CGG GCC CGG GGC GGC	218
GCC GGC GGG GGG GGC CCG GGT CCC GGG GGG	266
GGG GTC CTC TAC AAG CAA TCG ATC GCG CAG CGC GCG CGG ACC ATG GCG LTG Val Leu Tyr Lys Gln Ser Ile Ala Gln Arg Ala Arg Thr Met Ala 45 50 55	314
TG TAC AAC CCC ATC CCG GTC AAG CAG AAC TGC TTC ACC GTC AAC CGC eu Tyr Asn Pro Ile Pro Val Lys Gln Asn Cys Phe Thr Val Asn Arg	362

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TCG Ser	CTC Leu 75	TTC Phe	GTC Val	TTC Phe	AGC Ser	GAG Glu 80	GAC Asp	AAC Asn	GTC Val	GTC Val	Arg 85	AAA Lys	TAC Tyr	GCG Ala	AAG Lys	410
CGC Arg 90	ATC Ile	ACC Thr	GAG Glu	TGG Trp	CCT Pro 95	CCA Pro	TTC Phe	GAG Glu	AAT Asn	ATG Met 100	ATC Ile	CTG Leu	GCC Ala	ACC Thr	ATC Ile 105	458
ATC Ile	GCC Ala	AAC Asn	TGC Cys	ATC Ile 110	GTG Val	CTG Leu	GCC Ala	CTG Leu	GAG Glu 115	CAG Gln	CAC His	CTC Leu	CCT Pro	GAT Asp 120	GGG Gly	506
GAC Asp	AAA Lys	ACG Thr	CCC Pro 125	ATG Met	TCC Ser	GAG Glu	CGG Arg	CTG Leu 130	GAC Asp	GAC Asp	ACG Thr	GAG Glu	CCC Pro 135	TAT Tyr	TTC Phe	554
ATC Ile	GGG	ATC Ile 140	TTT Phe	TGC Cys	TTC Phe	GAG Glu	GCA Ala 145	GGG Gly	ATC Ile	AAA Lys	ATC Ile	ATC Ile 150	GCT Ala	CTG Leu	GGC Gly	602
TTT Phe	GTC Val 155	TTC Phe	CAC His	AAG Lys	GGC Gly	TCT Ser 160	TAC Tyr	CTG Leu	CGG Arg	AAC Asn	GGC Gly 165	TGG Trp	AAC Asn	GTC Val	ATG Met	650
GAC Asp 170	TTC Phe	GTG Val	GTC Val	GTC Val	CTC Leu 175	ACA Thr	GGG Gly	ATC Ile	CTT Leu	GCC Ala 180	ACG Thr	GCT Ala	GGA Gly	ACT Thr	GAC Asp 185	698
TTC Phe	GAC Asp	CTG Leu	CGA Arg	ACA Thr 190	CTG Leu	AGG Arg	GCT Ala	GTG Val	CGT Arg 195	GTG Val	CTG Leu	AGG Arg	CCC Pro	CTG Leu 200	AAG Lys	746
CTG Leu	GTG Val	TCT Ser	GGG Gly 205	ATT Ile	CCA Pro	AGT Ser	TTG Leu	CAG Gln 210	GTG Val	GTG Val	CTC Leu	AAG Lys	TCC Ser 215	ATC Ile	ATG Met	794
AAG Lys	GCC Ala	ATG Met 220	GTT Val	CCA Pro	CTC Leu	CTG Leu	CAG Gln 225	ATT	GGG Gly	CTG Leu	CTT Leu	CTC Leu 230	TTC Phe	TTT Phe	GCC Ala	842
ATC Ile	CTC Leu 235	ATG Met	TTT Phe	GCC Ala	ATC Ile	ATT Ile 240	GGC Gly	CTG Leu	GAG Glu	TTC Phe	TAC Tyr 245	ATG Met	GGC Gly	AAG Lys	TTC Phe	890
CAC His 250	Lys	GCC Ala	TGT Cys	TTC Phe	CCC Pro 255	AAC Asn	AGC Ser	ACA Thr	GAT Asp	GCG Ala 260	GAG Glu	CCC Pro	GTG Val	GGT Gly	GAC Asp 265	938
TTC Phe	CCC Pro	TGT Cys	GGC Gly	AAG Lys 270	Glu	GCC Ala	CCA Pro	GCC Ala	CGG Arg 275	Leu	TGC Cys	GAG Glu	GGC Gly	GAC Asp 280	ACT Thr	986
GAG Glu	TGC Cys	CGG Arg	GAG Glu 285	Tyr	TGG	CCA Pro	GGA Gly	CCC Pro 290	Asn	TTT Phe	GGC	ATC Ile	ACC Thr 295	ASD	TTT Phe	1034

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GAC Asp	AAT Asn	ATC Ile 300	ьeu	TTT Phe	GCC Ala	ATC	TTG Leu 305	Thr	GTG Val	TTC Phe	CAG Gln	TGC Cys 310	Ile	ACC Thr	ATG Met		1082
GAG Glu	GGC Gly 315	Trp	ACT Thr	GAC Asp	ATC	CTC Leu 320	Tyr	TAA naA	ACA Thr	AAC Asn	GAT Asp 325	Ala	GCC	GGC	AAC Asn		1130
ACC Thr 330	Trp	AAC Asn	TGG	CTC Leu	TAC Tyr 335	TTC Phe	ATC Ile	CCT Pro	CTC Leu	ATC Ile 340	Ile	ATC Ile	GGC	TCC	TTC Phe 345		1178
TTC Phe	ATG Met	CTC	AAC Asn	CTG Leu 350	GTG Val	CTG Leu	GGC Gly	GTG Val	CTC Leu 355	TCG Ser	GGG Gly	GAG Glu	TTT Phe	GCC Ala 360	AAG Lys		1226
GAG Glu	CGA Arg	GAG Glu	AGG Arg 365	GTG Val	GAG Glu	AAC Asn	CGC Arg	CGC Arg 370	GCC Ala	TTC Phe	CTG Leu	AAG Lys	CTG Leu 375	CGC Arg	CGG Arg		1274
CAG Gln	CAG Gln	CAG Gln 380	ATC Ile	GAG Glu	CGA Arg	GAG Glu	CTC Leu 385	AAC Asn	GGG Gly	TAC	CTG Leu	GAG Glu 390	TGG Trp	ATC Ile	TTC Phe		1322
ьуs	395	GIU	GIU	Val	Met	CTG Leu 400	Ala	Glu	Glu	Asp	Arg 405	Asn	Ala	Glu	Glu		1370
AAG Lys 410	TCC Ser	CCT Pro	TTG Leu	GAC Asp	GTG Val 415	CTG Leu	AAG Lys	AGA Arg	GCG Ala	GCC Ala 420	ACC Thr	AAG Lys	AAG Lys	AGC Ser	AGA Arg 425		1418
AAT Asn	GAC Asp	CTG Leu	ATC Ile	CAC His 430	GCA Ala	GAG Glu	GAG Glu	GGA Gly	GAG Glu 435	GAC Asp	CGG Arg	TTT Phe	GCA Ala	GAT Asp 440	CTC Leu		1466
Cys	Ата	vaI	445	ser	Pro	TTC Phe	Ala	Arg 450	Ala	Ser	Leu	Lys	Ser 455	Gly	Lys	,	1514
ACA Thr	GAG Glu	AGC Ser 460	TCG Ser	TCA Ser	TAC Tyr	TTC Phe	CGG Arg 465	AGG Arg	AAG Lys	GAG Glu	AAG Lys	ATG Met 470	TTC Phe	CGG Arg	TTT Phe		1562
TTT Phe	ATC Ile 475	CGG Arg	CGC Arg	ATG Met	GTG Val	AAG Lys 480	GCT Ala	CAG Gln	AGC Ser	TTC Phe	TAC Tyr 485	TGG Trp	GTG Val	GTG Val	CTG Leu	:	1610
TGC Cys 490	GTG Val	GTG Val	GCC Ala	CTG Leu	AAC Asn 495	ACA Thr	CTG Leu	TGT Cys	GTG Val	GCC Ala 500	ATG Met	GTG Val	CAT His	TAC Tyr	AAC Asn 505	:	1658
CAG Gln	CCG Pro	CGG Arg	Arg	CTT Leu 510	ACC Thr	ACG Thr	ACC Thr	CTG Leu	TAT Tyr 515	TTT Phe	GCA Ala	GAG Glu	TTT Phe	GTT Val 520	TTC Phe	:	1706

CTG Leu	GGT Gly	CTC Leu	TTC Phe 525	CTC Leu	ACA Thr	GAG Glu	ATG Met	TCC Ser 530	CTG Leu	AAG Lys	ATG Met	TAT Tyr	GGC Gly 535	CTG Leu	GGG Gly	1754
CCC Pro	AGA Arg	AGC Ser 540	TAC Tyr	TTC Phe	CGG Arg	TCC Ser	TCC Ser 545	TTC Phe	AAC Asn	TGC Cys	TTC Phe	GAC Asp 550	TTT Phe	GGG Gly	GTC Val	1802
ATC Ile	GTG Val 555	GGG Gly	AGC Ser	GTC Val	TTT Phe	GAA Glu 560	GTG Val	GTC Val	TGG Trp	GCG Ala	GCC Ala 565	ATC Ile	AAG Lys	CCG Pro	GGA Gly	1850
AGC Ser 570	TCC Ser	TTT Phe	GGG Gly	ATC Ile	AGT Ser 575	GTG Val	CTG Leu	CGG Arg	GCC Ala	CTC Leu 580	CGC Arg	CTG Leu	CTG Leu	AGG Arg	ATC Ile 585	1898
TTC Phe	AAA Lys	GTC Val	ACG Thr	AAG Lys 590	TAC Tyr	TGG Trp	AGC Ser	TCC Ser	CTG Leu 595	CGG Arg	AAC Asn	CTG Leu	GTG Val	GTG Val 600	TCC Ser	1946
CTG Leu	CTG Leu	AAC Asn	TCC Ser 605	ATG Met	AAG Lys	TCC Ser	ATC Ile	ATC Ile 610	AGC Ser	CTG Leu	CTC Leu	TTC Phe	TTG Leu 615	CTC Leu	TTC Phe	1994
CTG Leu	TTC Phe	ATT Ile 620	GTG Val	GTC Val	TTC Phe	GCC Ala	CTG Leu 625	CTG Leu	GGG Gly	ATG Met	CAG Gln	CTG Leu 630	TTT Phe	GGG Gly	GGA Gly	2042
CAG Gln	TTC Phe 635	AAC Asn	TTC Phe	CAG Gln	GAT Asp	GAG Glu 640	ACT Thr	CCC Pro	ACA Thr	ACC Thr	AAC Asn 645	TTC Phe	GAC Asp	ACC Thr	TTC Phe	2090
CCT Pro 650	GCC Ala	GCC Ala	ATC Ile	CTC Leu	ACT Thr 655	GTC Val	TTC Phe	CAG Gln	ATC Ile	CTG Leu 660	ACG Thr	GGA Gly	GAG Glu	GAC Asp	TGG Trp 665	2138
AAT Asn	GCA Ala	GTG Val	ATG Met	TAT Tyr 670	CAC His	GGG Gly	ATC Ile	GAA Glu	TCG Ser 675	CAA Gln	GGC Gly	GGC Gly	GTC Val	AGC Ser 680	AAA Lys	2186
GGC Gly	ATG Met	TTC Phe	TCG Ser 685	TCC Ser	TTT Phe	TAC Tyr	TTC Phe	ATT Ile 690	GTC Val	CTG Leu	ACA Thr	CTG Leu	TTC Phe 695	GGA Gly	AAC Asn	2234
TAC Tyr	ACT Thr	CTG Leu 700	Leu	AAT Asn	GTC Val	TTT Phe	CTG Leu 705	Ala	ATC Ile	GCT Ala	GTG Val	GAC Asp 710	Asn	CTG Leu	GCC Ala	2282
AAC Asn	GCC Ala 715	Gln	GAG Glu	CTG Leu	ACC Thr	AAG Lys 720	Asp	GAA Glu	GAG Glu	GAG Glu	ATG Met 725	GIU	GAA Glu	GCA Ala	GCC Ala	2330
AAT Asn 730	Gln	AAG Lys	CTT Leu	GCT Ala	CTG Leu 735	CAA Gln	AAG Lys	GCC Ala	AAA Lys	GAA Glu 740	Val	GCT Ala	GAA Glu	GTC Val	AGC Ser 745	2378

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CCC	ATG Met	TCI Ser	GCC Ala	GCG Ala 750	ASI	: ATC	TCC	ATC Ile	GCC Ala 755	Ala	AGG Arg	G CAG	CAG Gln	AAC Asn 760	TCG Ser	2426
GCC Ala	AAG Lys	GCG Ala	CGC Arg 765	Ser	GTG Val	TGG Trp	GAG Glu	CAG Gln 770	Arg	GCC Ala	AGC Ser	CAG Gln	CTA Leu 775	Arg	CTG Leu	2474
CAG Gln	AAC	CTG Leu 780	Arg	GCC Ala	AGC Ser	TGC Cys	GAG Glu 785	GCG Ala	CTG Leu	TAC	AGC Ser	GAG Glu 790	Met	GAC Asp	CCC Pro	2522
GAG Glu	GAG Glu 795	Arg	CTG Leu	CGC Arg	TTC Phe	GCC Ala 800	ACT Thr	ACG Thr	CGC Arg	CAC His	CTG Leu 805	Arg	CCC Pro	GAC Asp	ATG Met	2570
AAG Lys 810	ACG Thr	CAC	CTG Leu	GAC Asp	CGG Arg 815	CCG Pro	CTG Leu	GTG Val	GTG Val	GAG Glu 820	CTG Leu	GGC	CGC Arg	GAC Asp	GGC Gly 825	2618
GCG Ala	CGG Arg	GGG	CCC Pro	GTG Val 830	GGA Gly	GGC Gly	AAA Lys	GCC Ala	CGA Arg 835	CCT Pro	GAG Glu	GCT Ala	GCG Ala	GAG Glu 840	GCC Ala	2666
CCC Pro	GAG Glu	GGC Gly	GTC Val 845	GAC Asp	CCT Pro	CCG Pro	CGC Arg	AGG Arg 850	CAC His	CAC His	CGG Arg	CAC His	CGC Arg 855	GAC Asp	AAG Lys	2714
GAC Asp	AAG Lys	ACC Thr 860	CCC Pro	GCG Ala	GCG Ala	GGG Gly	GAC Asp 865	CAG Gln	GAC Asp	CGA Arg	GCA Ala	GAG Glu 870	GCC Ala	CCG Pro	AAG Lys	2762
GCG Ala	GAG Glu 875	AGC Ser	GGG Gly	GAG Glu	CCC Pro	GGT Gly 880	GCC Ala	CGG Arg	GAG Glu	GAG Glu	CGG Arg 885	CCG Pro	CGG Arg	CCG Pro	CAC His	2810
CGC Arg 890	AGC Ser	CAC His	AGC Ser	AAG Lys	GAG Glu 895	GCC Ala	GCG Ala	GGG Gly	CCC Pro	CCG Pro 900	GAG Glu	GCG Ala	CGG Arg	AGC Ser	GAG Glu 905	2858
CGC Arg	GGC Gly	CGA Arg	GGC Gly	CCA Pro 910	GGC Gly	CCC Pro	GAG Glu	GGC Gly	GGC Gly 915	CGG Arg	CGG Arg	CAC His	CAC His	CGG Arg 920	CGC Arg	2906
GGC	TCC Ser	CCG Pro	GAG Glu 925	GAG Glu	GCG Ala	GCC Ala	GAG Glu	CGG Arg 930	GAG Glu	CCC Pro	CGA Arg	CGC Arg	CAC His 935	CGC Arg	GCG Ala	2954
CAC His	CGG Arg	CAC His 940	CAG Gln	GAT Asp	CCG Pro	Ser	AAG Lys 945	GAG Glu	TGC Cys	GCC Ala	GGC Gly	GCC Ala 950	AAG Lys	GGC Gly	GAG Glu	3002
CGG Arg	CGC Arg 955	GCG Ala	CGG Arg	CAC His	CGC Arg	GGC Gly 960	GGC Gly	CCC Pro	CGA Arg	GCG Ala	GGG Gly 965	CCC Pro	CGG Arg	GAG Glu	GCG Ala	3050

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GAG AGC Glu Ser 970	GGG GAG Gly Glu	GAG CCG Glu Pro 975	Ala Ai	g CGG g Arg	CAC His	CGG Arg 980	GCC Ala	CGG Arg	CAC His	AAG Lys	GCG Ala 985	3098
CAG CCT Gln Pro	GCT CAC Ala His	GAG GCT Glu Ala 990	GTG GA	G AAG u Lys	GAG Glu 995	ACC Thr	ACG Thr	GAG Glu	AAG Lys	GAG Glu 100	Ala	3146
ACG GAG Thr Glu	AAG GAG Lys Glu 100	Ala Glu	ATA GI	G GAA 1 Glu 101	Ala	GAC Asp	AAG Lys	GAA Glu	AAG Lys 101	Glu	CTC Leu	3194
CGG AAC Arg Asn	CAC CAG His Gln 1020	CCC CGG Pro Arg	Glu Pr	A CAC O His 25	TGT Cys	GAC Asp	CTG Leu	GAG Glu 1030	Thr	AGT Ser	GGG Gly	3242
ACT GTG Thr Val 103	ACT GTG Thr Val 5	GGT CCC Gly Pro	ATG CA Met Hi 1040	C ACA s Thr	CTG Leu	CCC Pro	AGC Ser 104	Thr	TGT Cys	CTC Leu	CAG Gln	3290
AAG GTG Lys Val 1050	GAG GAA Glu Glu	CAG CCA Gln Pro 105	Glu As	T GCA p Ala	GAC Asp	AAT Asn 1060	Gln	CGG Arg	AAC Asn	GTC Val	ACT Thr 1065	3338
CGC ATG Arg Met	GGC AGT Gly Ser	CAG CCC Gln Pro 1070	CCA GA Pro As	C CCG p Pro	AAC Asn 107	Thr	ATT Ile	GTA Val	CAT His	ATC Ile 1080	Pro	3386
GTG ATG Val Met	CTG ACG Leu Thr 108	Gly Pro	CTT GG Leu Gl	G GAA y Glu 109	Ala	ACG Thr	GTC Val	GTT Val	CCC Pro 1095	Ser	GGT Gly	3434
AAC GTG Asn Val	GAC CTG Asp Leu 1100	GAA AGC Glu Ser	CAA GC Gln Al 11	a Glu	GGG Gly	AAG Lys	AAG Lys	GAG Glu 1110	Val	GAA Glu	GCG Ala	3482
GAT GAC Asp Asp 1115	GTG ATG Val Met	AGG AGC Arg Ser	GGC CC Gly Pr 1120	C CGG o Arg	CCT Pro	ATC Ile	GTC Val 112	Pro	TAC Tyr	AGC Ser	TCC Ser	3530
	TGT TTA Cys Leu		Thr As				Arg					3578
ATC GTG Ile Val	ACC ATG Thr Met	AGG TAC Arg Tyr 1150	TTC GA Phe Gl	G GTG u Val	GTC Val 1155	Ile	CTC Leu	GTG Val	GTC Val	ATC Ile 1160	Ala	. 3626
	AGC ATC Ser Ile 116	Ala Leu			Asp					Asp		3674
	AAC AAC Asn Asn 1180			r Leu					Thr			3722

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															•	
TTT Phe	ACC Thr 119	TTT Phe 5	GAG Glu	ATG Met	GTG Val	ATA Ile 120	Lys	ATG Met	ATC Ile	GAC Asp	TTG Leu 120	Gly	CTG Leu	CTG Leu	CTT Leu	3770
CAC His 121	Pro	GGA Gly	GCC Ala	TAT Tyr	TTC Phe 121!	Arg	GAC Asp	TTG Leu	TGG Trp	AAC Asn 122	Ile	CTG Leu	GAC Asp	TTC Phe	ATT Ile 1225	3818
GTG Val	GTC Val	AGT Ser	GGC Gly	GCC Ala 1230	Leu	GTG Val	GCG Ala	TTT Phe	GCT Ala 123	Phe	TCA Ser	GGA Gly	TCC Ser	AAA Lys 1240	Gly	3866
AAA Lys	GAC Asp	ATC Ile	AAT Asn 1245	Thr	ATC Ile	AAG Lys	TCT Ser	CTG Leu 1250	Arg	GTC Val	CTT Leu	CGT Arg	GTC Val 1255	Leu	CGG Arg	3914
CCC Pro	CTC Leu	AAG Lys 1260	Thr	ATC Ile	AAA Lys	CGG Arg	CTG Leu 1265	Pro	AAG Lys	CTC Leu	AAG Lys	GCT Ala 1270	Val	TTT Phe	GAC Asp	3962
TGT Cys	GTG Val 1275	GTG Val	AAC Asn	TCC Ser	CTG Leu	AAG Lys 1280	Asn	GTC Val	CTC Leu	AAC Asn	ATC Ile 1285	Leu	ATT Ile	GTC Val	TAC Tyr	4010
ATG Met 1290	Leu	TTC Phe	ATG Met	TTC Phe	ATA Ile 1295	Phe	GCC Ala	GTC Val	ATT Ile	GCG Ala 1300	Val	CAG Gln	CTC Leu	TTC Phe	AAA Lys 1305	4058
GGG	AAG Lys	TTT Phe	TTC Phe	TAC Tyr 1310	Cys	ACA Thr	GAT Asp	GAA Glu	TCC Ser 1315	Lys	GAG Glu	CTG Leu	GAG Glu	AGG Arg 1320	Asp	4106
TGC Cys	AGG Arg	GGT Gly	CAG Gln 1325	Tyr	TTG Leu	GAT Asp	TAT Tyr	GAG Glu 1330	Lys	GAG Glu	GAA Glu	GTG Val	GAA Glu 1335	Ala	CAG Gln	4154
CCC Pro	AGG Arg	CAG Gln 1340	\mathtt{Trp}	AAG Lys	AAA Lys	TAC Tyr	GAC Asp 1345	Phe	CAC His	TAC Tyr	GAC Asp	AAT Asn 1350	Val	CTC Leu	TGG Trp	4202
GCT Ala	CTG Leu 1355	CTG Leu	ACG Thr	CTG Leu	TTC Phe	ACA Thr 1360	Val	TCC Ser	ACG Thr	GGA Gly	GAA Glu 1365	Gly	TGG Trp	CCC Pro	ATG Met	4250
	Leu	AAA Lys		Ser		Asp					Glu			Pro		4298
		TAC Tyr	Arg		Glu			Ile		Tyr			Tyr		Val	4346
GTC Val	TTT Phe	CCC Pro	TTC Phe 1405	Phe	TTC Phe	GTC Val	Asn	ATC Ile 1410	Phe	GTG Val	GCT Ala	Leu	ATC Ile 1415	Ile	ATC Ile	4394

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ACC TTC CAG GAG CATTLE Phe Gln Glu Glu 1420	in Gly Asp Lys \ 1425	Val Met Ser Glu	1430	GIU
AAG AAC GAG AGG GG Lys Asn Glu Arg Al 1435	CT TGC ATT GAC 1 la Cys Ile Asp I 1440	TTC GCC ATC AGC Phe Ala Ile Ser 144!	Ala Lys Pro	CTG 4490 Leu
ACA CGG TAC ATG CG Thr Arg Tyr Met P: 1450	CC CAA AAC CGG (ro Gln Asn Arg (1455	CAG TCG TTC CAG Gln Ser Phe Gln 1460	TAT AAG ACG Tyr Lys Thr	TGG 4538 Trp 1465
ACA TTT GTG GTC TO Thr Phe Val Val So	CC CCG CCC TTT (er Pro Pro Phe (470	GAA TAC TTC ATC Glu Tyr Phe Ile 1475	ATG GCC ATG Met Ala Met 1480	ITE
GCC CTC AAC ACT G Ala Leu Asn Thr V 1485	al Val Leu Met I	ATG AAG TTC TAT Met Lys Phe Tyr 1490	GAT GCA CCC Asp Ala Pro 1495	TAT 4634 Tyr
GAG TAC GAG CTG A' Glu Tyr Glu Leu M 1500	TG CTG AAA TGC o et Leu Lys Cys : 1505	Leu Asn Ile Val	TTC ACA TCC Phe Thr Ser 1510	ATG 4682 Met
TTC TCC ATG GAA T Phe Ser Met Glu C 1515	GC GTG CTG AAG . ys Val Leu Lys 1520	ATC ATC GCC TTT Ile Ile Ala Phe 152	GIA ANT TER	AAC 4730 Asn
TAT TTC AGA GAT G Tyr Phe Arg Asp A 1530	CC TGG AAT GTC la Trp Asn Val 1535	TTT GAC TTT GTC Phe Asp Phe Val 1540	ACT GTG TTG Thr Val Leu	GGA 4778 Gly 1545
AGT ATT ACT GAT A Ser Ile Thr Asp I	TT TTA GTA ACA le Leu Val Thr 550	GAG ATT GCG GAA Glu Ile Ala Glu 1555	ACG AAC AAT Thr Asn Asn 156	Pne
ATC AAC CTC AGC T Ile Asn Leu Ser P 1565	TC CTC CGC CTC The Leu Arg Leu	TTT CGA GCT GCG Phe Arg Ala Ala 1570	CGG CTG ATC Arg Leu Ile 1575	AAG 4874 Lys
CTG CTC CGC CAG G Leu Leu Arg Gln G 1580	GC TAC ACC ATC ly Tyr Thr Ile 1585	Arg Ile Leu Leu	TGG ACC TTT Trp Thr Phe 1590	GTC 4922 Val
CAG TCC TTC AAG G Gln Ser Phe Lys A 1595	SCC CTG CCC TAC Ala Leu Pro Tyr 1600	GTG TGT CTG CTC Val Cys Leu Leu 160	I TIE MIG MEC	CTG 4970 Leu
TTC TTC ATC TAC G Phe Phe Ile Tyr A 1610	GCC ATC ATC GGC Ala Ile Ile Gly 1615	ATG CAG GTG TTT Met Gln Val Phe 1620	GGG AAT ATT Gly Asn Ile	GCC 5018 Ala 1625
CTG GAT GAT GAC A	ACC AGC ATC AAC Thr Ser Ile Asn 1630	CGC CAC AAC AAC Arg His Asn Asn 1635	TTC CGG ACG Phe Arg Thi	. File

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тт	ים כיש	A GC	C CT(~ 3 m/	- cmc											
Le	u Gl	n Al	a Le		Leu	Lev	TT(AGG Arg	y se:	C GC r Ala	C ACC	G GGC r Gly	GA(Gl) 16!	ı Ala	C TGG a Trp	5114
CA Hi	C GA s Gl	G ATO	C 14C	G CTC	TCC Ser	TGC Cys	CTC Leu 166	ı sei	C AAC	C CAC	G GCC	TGI Cys 167	Asp	GA(G CAG	5162
GC Al	C AA a As 16		C ACC	GAG Glu	TGT Cys	GGA Gly 168	sei	GAC Asp	TTT Phe	GCC Ala	TAC Tyr 168	Phe	TAC	TTO Phe	GTC Val	5210
TC Se 16	C TT r Ph 90	C ATO	TTC Phe	CTG Leu	TGC Cys 169	ser	TTT Phe	CTG Leu	ATO Met	TTC Lev	l Asn	CTC Leu	TTT Phe	GTC Val	GCT Ala 1705	5258
GT(Va	G ATO	C ATO	GAC Asp	AAT Asn 171	T-116	GAG Glu	TAC Tyr	CTC	ACG Thr	Arg	GAC Asp	TCT Ser	TCC Ser	ATC Ile	CTA Leu	5306
GG! Gly	r cc: y Pro	CAC His	CAC His	Leu	GAT Asp	GAG Glu	TTC Phe	ATC Ile 173	Arg	GTC Val	TGG Trp	GCT Ala	GAA Glu 173	Tyr	GAC Asp	5354
Pro	G GCT	GCG Ala 174	. Cys	GGG Gly	CGC Arg	ATC Ile	AGT Ser 174	iyr	AAT Asn	GAC • Asp	ATG Met	TTT Phe 1750	Glu	ATG Met	CTG Leu	5402
AA/ Lys	CAC His	1-1-0	TCC Ser	CCG Pro	CCT Pro	CTG Leu 1760	GTÅ	CTG Leu	GGG Gly	AAG Lys	AAA Lys 176	Cys	CCT Pro	GCT Ala	CGA Arg	5450
GTT Val 177	710	TAC	AAG Lys	CGC Arg	CTG Leu 1775	val	CGC Arg	ATG Met	AAC Asn	ATG Met 178	Pro	ATC Ile	TCC Ser	AAC Asn	GAG Glu 1785	5498
GAC Asp	ATG Met	ACT Thr	GTT Val	CAC His 1790	Pne	ACG Thr	TCC Ser	ACG Thr	CTG Leu 179	Met	GCC Ala	CTC Leu	ATC Ile	CGG Arg 1800	Thr	5546
GCA Ala	CTG Leu	GAG Glu	ATC Ile 1805	AAG Lys	CTG Leu	GCC Ala	CCA Pro	GCT Ala 1810	Gly	ACA Thr	AAG Lys	Gln	CAT His 1815	Gln	TGT Cys	5594
GAC Asp	GCG Ala	GAG Glu 1820	ьeu	AGG Arg	AAG Lys	Glu	ATT Ile 1825	Ser	GTT Val	GTG Val	TGG Trp	GCC Ala 1830	Asn	CTG Leu	CCC Pro	5642
CAG Gln	AAG Lys 183	Inr	TTG Leu	GAC Asp	Leu :	CTG Leu 1840	GTA Val	CCA Pro	CCC Pro	CAT His	AAG Lys 1845	Pro .	GAT Asp	GAG Glu	ATG Met	5690
ACA Thr 185	val	GGG Gly	AAG Lys	GTT Val	TAT (Tyr 1 1855	GCA (Ala)	GCT Ala	CTG Leu	Met	ATA Ile 1860	Phe	GAC (Asp)	TTC Phe	Tyr	AAG Lys 1865	. 5738

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CAG Gln	AAC Asn	AAA Lys	ACC Thr	ACC Thr 1870	Arg	GAC Asp	CAG Gln	ATG Met	CAG Gln 1875	Gln	GCT Ala	CCT Pro	GGA Gly	GGC Gly 1880	Leu	5786
TCC Ser	CAG Gln	ATG Met	GGT Gly 1885	Pro	GTG Val	TCC Ser	CTG Leu	TTC Phe 1890	His	CCT Pro	CTG Leu	AAG Lys	GCC Ala 1899	Thr	CTG Leu	5834
GAG Glu	CAG Gln	ACA Thr 1900	Gln	CCG Pro	GCT Ala	GTG Val	CTC Leu 1905	Arg	GGA Gly	GCC Ala	CGG Arg	GTT Val 1910	Phe	CTT Leu	CGA Arg	5882
CAG Gln	AAG Lys 1915	Ser	TCC Ser	ACC Thr	TCC Ser	CTC Leu 1920	Ser	AAT Asn	GGC Gly	Gly	GCC Ala 192	ATA [.] Ile	CAA Gln	AAC Asn	CAA Gln	5930
GAG Glu 1930	Ser	GGC Gly	ATC Ile	AAA Lys	GAG Glu 1935	Ser	GTC Val	TCC Ser	TGG Trp	GGC Gly 1940	Thr	CAA Gln	AGG Arg	ACC Thr	CAG Gln 1945	5978
GAT Asp	GCA Ala	CCC Pro	CAT His	GAG Glu 1950	Ala	AGG Arg	CCA Pro	CCC Pro	CTG Leu 195	Glu	CGT Arg	GGC Gly	CAC His	TCC Ser 1960	Thr	6026
GAG Glu	ATC Ile	CCT Pro	GTG Val 196!	Gly	CGG Arg	TCA Ser	GGA Gly	GCA Ala 1970	Leu	GCT Ala	GTG Val	GAC Asp	GTT Val 1975	Gln	ATG Met	6074
CAG Gln	AGC Ser	ATA Ile 1980	Thr	CGG Arg	AGG Arg	GGC Gly	CCT Pro 198	Asp	GGG Gly	GAG Glu	CCC Pro	CAG Gln 1990	Pro	GGG Gly	CTG Leu	6122
GAG Glu	AGC Ser 199	Gln	GGT Gly	CGA Arg	GCG Ala	GCC Ala 200	Ser	ATG Met	CCC Pro	CGC Arg	CTT Leu 200	GCG Ala 5	GCC Ala	GAG Glu	ACT Thr	6170
CAG Gln 201	Pro	GTC Val	ACA Thr	GAT Asp	GCC Ala 201	Ser	CCC Pro	ATG Met	AAG Lys	CGC Arg 202	Ser	ATC Ile	TCC Ser	ACG Thr	CTG Leu 2025	6218
GCC Ala	CAG Gln	CGG Arg	CCC Pro	CGT Arg 203	Gly	ACT Thr	CAT His	CTT	TGC Cys 203	Ser	ACC Thr	ACC Thr	CCG Pro	GAC Asp 204	Arg	6266
CCA Pro	CCC Pro	CCT Pro	AGC Ser 204	Gln	GCG Ala	TCG Ser	TCG Ser	CAC His 205	His	CAC His	CAC His	CAC His	CGC Arg 205	Cys	CAC His	6314
CGC Arg	CGC Arg	AGG Arg 206	Asp	AGG Arg	AAG Lys	CAG Gln	AGG Arg 206	Ser	CTG Leu	GAG Glu	AAG Lys	GGG Gly 207	Pro	AGC Ser	CTG Leu	6362
TCT Ser	GCC Ala 207	Asp	ATG Met	GAT Asp	GGC Gly	GCA Ala 208	Pro	AGC Ser	AGT Ser	GCT	GTG Val 208	GIY	CCG Pro	GGG Gly	CTG Leu	6410

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CCC CCG GGA GAG GGG CCT ACA GGC TGC CGG CGG GAA CGA GAG CGC CGG Pro Pro Gly Glu Gly Pro Thr Gly Cys Arg Arg Glu Arg Glu Arg Arg 2090 2095 2100 2105	6458
CAG GAG CGG GGC CGG TCC CAG GAG CGG AGG CAG CCC TCA TCC TCC Gln Glu Arg Gly Arg Ser Gln Glu Arg Gln Pro Ser Ser Ser Ser 2110 2115 2120	6506
TCG GAG AAG CAG CGC TTC TAC TCC TGC GAC CGC TTT GGG GGC CGT GAG Ser Glu Lys Gln Arg Phe Tyr Ser Cys Asp Arg Phe Gly Gly Arg Glu 2125 2130 2135	6554
CCC CCG AAG CCC AAG CCC TCC CTC AGC AGC CAC CCA ACG TCG CCA ACA Pro Pro Lys Pro Lys Pro Ser Leu Ser Ser His Pro Thr Ser Pro Thr 2140 2145 2150	6602
GCT GGC CAG GAG CCG GGA CCC CAC CCA CAG GGC AGT GGT TCC GTG AAT Ala Gly Gln Glu Pro Gly Pro His Pro Gln Gly Ser Gly Ser Val Asn 2155 2160 2165	6650
GGG AGC CCC TTG CTG TCA ACA TCT GGT GCT AGC ACC CCC GGC CGC GGT Gly Ser Pro Leu Leu Ser Thr Ser Gly Ala Ser Thr Pro Gly Arg Gly 2170 2185	6698
GGG CGG AGG CAG CTC CCC CAG ACG CCC CTG ACT CCC CGC CCC AGC ATC Gly Arg Arg Gln Leu Pro Gln Thr Pro Leu Thr Pro Arg Pro Ser Ile 2190 2195 2200	6746
ACC TAC AAG ACG GCC AAC TCC TCA CCC ATC CAC TTC GCC GGG GCT CAG Thr Tyr Lys Thr Ala Asn Ser Ser Pro Ile His Phe Ala Gly Ala Gln 2205 2210 2215	6794
ACC AGC CTC CCT GCC TTC TCC CCA GGC CGG CTC AGC CGT GGG CTT TCC Thr Ser Leu Pro Ala Phe Ser Pro Gly Arg Leu Ser Arg Gly Leu Ser 2220 2225 2230	6842
GAA CAC AAC GCC CTG CTG CAG AGA GAC CCC CTC AGC CAG CCC CTG GCC Glu His Asn Ala Leu Leu Gln Arg Asp Pro Leu Ser Gln Pro Leu Ala 2235 2240 2245	6890
CCT GGC TCT CGA ATT GGC TCT GAC CCT TAC CTG GGG CAG CGT CTG GAC Pro Gly Ser Arg Ile Gly Ser Asp Pro Tyr Leu Gly Gln Arg Leu Asp 2250 2260 2265	6938
AGT GAG GCC TCT GTC CAC GCC CTG CCT GAG GAC ACG CTC ACT TTC GAG Ser Glu Ala Ser Val His Ala Leu Pro Glu Asp Thr Leu Thr Phe Glu 2270 2275 2280	6986
GAG GCT GTG GCC ACC AAC TCG GGC CGC TCC TCC AGG ACT TCC TAC GTG Glu Ala Val Ala Thr Asn Ser Gly Arg Ser Ser Arg Thr Ser Tyr Val 2285 2290 2295	7034
TCC TCC CTG ACC TCC CAG TCT CAC CCT CTC CGC CGC GTG CCC AAC GGT Ser Ser Leu Thr Ser Gln Ser His Pro Leu Arg Arg Val Pro Asn Gly 2300 2305 2310	7082

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TAC CAC TGC ACC CTG GGA CTC AGC TCG GGT GGC CGA GCA CGG CAC AGC Tyr His Cys Thr Leu Gly Leu Ser Ser Gly Gly Arg Ala Arg His Ser 2315 2320 2325	713
TAC CAC CAC CCT GAC CAA GAC CAC TGG TGC TAGCTGCACC GTGACCGCTC Tyr His His Pro Asp Gln Asp His Trp Cys 2330 2335 234	. ⁷¹⁸⁰
AGACGCCTGC ATGCAGCAGG CGTGTGTTCC AGTGGATGAG TTTTATCATC CACACGGGGC	7240
AGTCGGCCCT CGGGGGAGGC CTTGCCCACC TTGGTGAGGC TCCTGTGGCC CCTCCCTCCC	7300
CCTCCTCCCC TCTTTTACTC TAGACGACGA ATAAAGCCCT GTTGCTTGAG TGTACGTACC	7360
GC	7362
(2) INFORMATION FOR SEQ ID NO:8:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 7175 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1446857	
(ix) FEATURE: (A) NAME/KEY: 5'UTR (B) LOCATION: 1143	
(ix) FEATURE: (A) NAME/KEY: 3'UTR (B) LOCATION: 68557175	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:	
GCGGCGGCGG CTGCGGCGGT GGGGCCGGGC GAGGTCCGTG CGGTCCCGGC GGCTCCGTGG	60
CTGCTCCGCT CTGAGCGCCT GCGCGCCCCG CGCCCTCCCT GCCGGGGCCG CTGGGCCGGG	120
GATGCACGCG GGGCCCGGGA GCC ATG GTC CGC TTC GGG GAC GAG CTG GGC Met Val Arg Phe Gly Asp Glu Leu Gly 1 5	170
GGC CGC TAT GGA GGC CCC GGC GGC GGA GAG CGG GCC CGG GGC GGC	211
GCC GGC GGG GGG GGC CCG GGT CCC GGG GGG	266

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CGG Arg	GTC Val	CTC Leu	TAC Tyr 45	. TAs	CAA Gln	TCG Ser	ATC Ile	GCG Ala 50	Gln	CGC Arg	GCG Ala	CGG Arg	ACC Thr	Met	GCG Ala	314
CTG Leu	TAC	AAC Asn 60	Pro	ATC Ile	CCG Pro	GTC Val	AAG Lys 65	Gln	AAC Asn	TGC Cys	TTC Phe	ACC Thr	Val	AAC Asn	CGC	362
TCG Ser	CTC Leu 75	Pne	GTC Val	TTC	AGC Ser	GAG Glu 80	GAC Asp	AAC Asn	GTC Val	GTC Val	CGC Arg 85	Lys	TAC Tyr	GCG Ala	AAG Lys	410
CGC Arg 90	ATC Ile	ACC	GAG Glu	TGG Trp	CCT Pro 95	CCA Pro	TTC Phe	GAG Glu	AAT Asn	ATG Met 100	Ile	CTG Leu	GCC Ala	ACC Thr	ATC Ile 105	458
ATC Ile	GCC Ala	AAC Asn	TGC Cys	ATC Ile 110	GTG Val	CTG Leu	GCC Ala	CTG Leu	GAG Glu 115	CAG Gln	CAC His	CTC Leu	CCT Pro	GAT Asp 120	GGG Gly	506
GAC Asp	AAA Lys	ACG Thr	CCC Pro 125	ATG Met	TCC Ser	GAG Glu	CGG Arg	CTG Leu 130	GAC Asp	GAC Asp	ACG Thr	GAG Glu	CCC Pro 135	TAT Tyr	TTC Phe	554
ATC Ile	GGG Gly	ATC Ile 140	TTT Phe	TGC Cys	TTC Phe	GAG Glu	GCA Ala 145	GGG Gly	ATC Ile	AAA Lys	ATC Ile	ATC Ile 150	GCT Ala	CTG Leu	GGC Gly	602
TTT Phe	GTC Val 155	TTC Phe	CAC His	AAG Lys	GGC Gly	TCT Ser 160	TAC Tyr	CTG Leu	CGG Arg	AAC Asn	GGC Gly 165	TGG Trp	AAC Asn	GTC Val	ATG Met	650
GAC Asp 170	TTC Phe	GTG Val	GTC Val	GTC Val	CTC Leu 175	ACA Thr	GGG Gly	ATC Ile	CTT Leu	GCC Ala 180	ACG Thr	GCT Ala	GGA Gly	ACT Thr	GAC Asp 185	698
rrc Phe	GAC Asp	CTG Leu	CGA Arg	ACA Thr 190	CTG Leu	AGG Arg	GCT Ala	GTG Val	CGT Arg 195	GTG Val	CTG Leu	AGG Arg	CCC Pro	CTG Leu 200	AAG Lys	746
CTG Leu	GTG Val	TCT Ser	GGG Gly 205	ATT Ile	CCA Pro	AGT Ser	TTG Leu	CAG Gln 210	GTG Val	GTG Val	CTC Leu	AAG Lys	TCC Ser 215	ATC Ile	ATG Met	794
AAG Lys	GCC Ala	ATG Met 220	GTT Val	CCA Pro	CTC Leu	CTG Leu	CAG Gln 225	ATT Ile	GGG Gly	CTG Leu	CTT Leu	CTC Leu 230	TTC Phe	TTT Phe	GCC Ala	842
TC le	CTC Leu 235	ATG Met	TTT Phe	GCC Ala	ATC Ile	ATT Ile 240	GGC Gly	CTG Leu	GAG Glu	Phe	TAC Tyr 245	ATG Met	GGC Gly	AAG Lys	TTC Phe	890
CAC lis !50	AAG Lys	GCC Ala	TGT Cys	Phe	CCC Pro 255	AAC Asn	AGC Ser	ACA Thr	Asp	GCG Ala 260	GAG Glu	CCC Pro	GTG Val	GGT Gly	GAC Asp 265	938

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TTC Phe	CCC Pro	TGT Cys	GGC Gly	AAG Lys 270	GAG Glu	GCC Ala	CCA Pro	GCC Ala	CGG Arg 275	CTG Leu	TGC Cys	GAG Glu	GGC	GAC Asp 280	ACT Thr	986	5
GAG Glu	TGC Cys	CGG Arg	GAG Glu 285	TAC Tyr	TGG Trp	CCA Pro	GGA Gly	CCC Pro 290	AAC Asn	TTT Phe	GGC Gly	ATC Ile	ACC Thr 295	AAC Asn	TTT Phe	1034	1
GAC Asp	AAT Asn	ATC Ile 300	CTG Leu	TTT Phe	GCC Ala	ATC Ile	TTG Leu 305	ACG Thr	GTG Val	TTC Phe	CAG Gln	TGC Cys 310	ATC Ile	ACC Thr	ATG Met	1082	2
GAG Glu	GGC Gly 315	TGG Trp	ACT Thr	GAC Asp	ATC Ile	CTC Leu 320	TAT Tyr	AAT Asn	ACA Thr	AAC Asn	GAT Asp 325	GCG Ala	GCC Ala	GGC Gly	AAC Asn	1130)
ACC Thr 330	TGG Trp	AAC Asn	TGG Trp	CTC Leu	TAC Tyr 335	TTC Phe	ATC Ile	CCT Pro	CTC Leu	ATC Ile 340	ATC Ile	ATC Ile	GGC Gly	TCC Ser	TTC Phe 345	1178	3
TTC Phe	ATG Met	CTC Leu	AAC Asn	CTG Leu 350	GTG Val	CTG Leu	GGC Gly	GTG Val	CTC Leu 355	TCG Ser	GGG Gly	GAG Glu	TTT Phe	GCC Ala 360	AAG Lys	1226	5
GAG Glu	CGA Arg	GAG Glu	AGG Arg 365	GTG Val	GAG Glu	AAC Asn	CGC Arg	CGC Arg 370	GCC Ala	TTC Phe	CTG Leu	AAG Lys	CTG Leu 375	CGC Arg	CGG Arg	1274	1
CAG Gln	CAG Gln	CAG Gln 380	ATC Ile	GAG Glu	CGA Arg	GAG Glu	CTC Leu 385	AAC Asn	GGG Gly	TAC Tyr	CTG Leu	GAG Glu 390	TGG Trp	ATC Ile	TTC Phe	1322	2
AAG Lys	GCG Ala 395	GAG Glu	GAA Glu	GTC Val	ATG Met	CTG Leu 400	GCC Ala	GAG Glu	GAG Glu	GAC Asp	AGG Arg 405	AAT Asn	GCA Ala	GAG Glu	GAG Glu	1370	D
AAG Lys 410	TCC Ser	CCT Pro	TTG Leu	GAC Asp	GTG Val 415	CTG Leu	AAG Lys	AGA Arg	GCG Ala	GCC Ala 420	ACC Thr	AAG Lys	AAG Lys	AGC Ser	AGA Arg 425	141	В
AAT Asn	GAC Asp	CTG Leu	ATC Ile	CAC His 430	GCA Ala	GAG Glu	GAG Glu	GGA Gly	GAG Glu 435	GAC Asp	CGG Arg	TTT Phe	GCA Ala	GAT Asp 440	CTC Leu	146	6
TGT Cys	GCT Ala	GTT Val	GGA Gly 445	Ser	CCC Pro	TTC Phe	GCC Ala	CGC Arg 450	Ala	AGC Ser	CTC Leu	AAG Lys	AGC Ser 455	GGG Gly	AAG Lys	151	4
ACA Thr	GAG Glu	AGC Ser 460	Ser	TCA Ser	TAC	TTC Phe	CGG Arg 465	Arg	AAG Lys	GAG Glu	AAG Lys	ATG Met 470	Pne	CGG Arg	TTT Phe	156	2
TTT Phe	ATC Ile 475	Arg	CGC Arg	ATG Met	GTG Val	AAG Lys 480	Ala	CAG Gln	AGC Ser	TTC	TAC Tyr 485	Trp	GTG Val	GTG Val	CTG Leu	161	.0

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TGC Cys 490	val	GTG Val	GCC Ala	CTG Leu	AAC Asn 495	Thr	CTG	TG1 Cys	GTG Val	GCC Ala	Met	GTG Val	CAT His	TAC	AAC Asn 505		1658
GII	Pro	Arg	Arg	510	Thr	Thr	Thr	Leu	515	Phe	: Ala	Glu	Phe	Val 520			1706
Dea	. Gly	Leu	525	reu	Thr	Glu	Met	530	Leu	Lys	Met	TAT	Gly 535	Leu	Gly		1754
FIU	ALG	540	TYE	Pne	Arg	Ser	545	Phe	Asn	Cys	Phe	GAC Asp 550	Phe	Gly	Val		1802
116	555	GIY	ser	vai	Pne	560	Val	Val	Trp	Ala	Ala 565		Lys	Pro	Gly		1850
570	ser	Pne	GIA	TTE	575	Val	Leu	Arg	Ala	Leu 580	Arg	CTG Leu	Leu	Arg	Ile 585		1898
PME	цуѕ	vai	THE	590	lyr	Trp	Ser	Ser	Leu 595	Arg	Asn	CTG Leu	Val	Val 600	Ser		1946
Leu	Leu	Asn	605	Met	Lys	Ser	Ile	Ile 610	Ser	Leu	Leu	TTC Phe	Leu 615	Leu	Phe		1994
Leu	Pne	620	Val	Val	Phe	Ala	Leu 625	Leu	Gly	Met	Gln	CTG Leu 630	Phe	Gly	Gly		2042
GIII	635	ASI	Pne	GIn	Asp	G1u 640	Thr	Pro	Thr	Thr	Asn 645	TTC Phe	Asp	Thr	Phe		2090
CCT Pro 650	GCC Ala	GCC Ala	ATC Ile	CTC Leu	ACT Thr 655	GTC Val	TTC Phe	CAG Gln	ATC Ile	CTG Leu 660	ACG Thr	GGA Gly	GAG Glu	GAC Asp	TGG Trp 665		2138
AAT Asn	GCA Ala	GTG Val	ATG Met	TAT Tyr 670	CAC His	GGG Gly	ATC Ile	GAA Glu	TCG Ser 675	CAA Gln	GGC Gly	GGC Gly	Val	AGC Ser 680	AAA Lys	:	2186
GGC Gly	ATG Met	Phe	TCG Ser 685	TCC Ser	TTT Phe	TAC Tyr	Phe	ATT Ile 690	GTC Val	CTG Leu	ACA Thr	CTG Leu	TTC Phe 695	GGA Gly	AAC Asn	:	2234
TAC Tyr	Thr	CTG Leu 700	CTG . Leu .	AAT Asn	GTC Val	Phe	CTG Leu 705	GCC Ala	ATC Ile	GCT Ala	GTG Val	GAC . Asp . 710	AAC Asn	CTG Leu	GCC Ala	:	2282

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AAC Asn	GCC Ala 715	CAA Gln	GAG Glu	CTG Leu	ACC Thr	AAG Lys 720	GAT Asp	GAA Glu	GAG Glu	GAG Glu	ATG Met 725	GAA Glu	GAA Glu	GCA Ala	GCC Ala		2330
AAT Asn 730	CAG Gln	AAG Lys	CTT Leu	GCT Ala	CTG Leu 735	CAA Gln	AAG Lys	GCC Ala	AAA Lys	GAA Glu 740	GTG Val	GCT Ala	GAA Glu	GTC Val	AGC Ser 745	-	2378
CCC Pro	ATG Met	TCT Ser	GCC Ala	GCG Ala 750	AAC Asn	ATC Ile	TCC Ser	ATC Ile	GCC Ala 755	GCC Ala	AGG Arg	CAG Gln	CAG Gln	AAC Asn 760	TCG Ser		2426
GCC Ala	AAG Lys	GCG Ala	CGC Arg 765	TCG Ser	GTG Val	TGG Trp	GAG Glu	CAG Gln 770	CGG Arg	GCC Ala	AGC Ser	CAG Gln	CTA Leu 775	CGG Arg	CTG Leu		2474
CAG Gln	AAC Asn	CTG Leu 780	CGG Arg	GCC Ala	AGC Ser	TGC Cys	GAG Glu 785	GCG Ala	CTG Leu	TAC Tyr	AGC Ser	GAG Glu 790	ATG Met	GAC Asp	CCC Pro		2522
GAG Glu	GAG Glu 795	CGG Arg	CTG Leu	CGC Arg	TTC Phe	GCC Ala 800	ACT Thr	ACG Thr	CGC Arg	CAC His	CTG Leu 805	CGG Arg	CCC Pro	GAC Asp	ATG Met		2570
AAG Lys 810	ACG Thr	CAC His	CTG Leu	GAC Asp	CGG Arg 815	CCG Pro	CTG Leu	GTG Val	GTG Val	GAG Glu 820	CTG Leu	GGC Gly	CGC Arg	GAC Asp	GGC Gly 825		2618
GCG Ala	CGG Arg	GGG Gly	CCC Pro	GTG Val 830	GGA Gly	GGC Gly	AAA Lys	GCC Ala	CGA Arg 835	CCT Pro	GAG Glu	GCT Ala	GCG Ala	GAG Glu 840	GCC Ala		2666
CCC Pro	GAG Glu	GGC Gly	GTC Val 845	GAC Asp	CCT Pro	CCG Pro	CGC Arg	AGG Arg 850	CAC His	CAC His	CGG Arg	CAC His	CGC Arg 855	GAC Asp	AAG Lys		2714
GAC Asp	AAG Lys	ACC Thr 860	CCC Pro	GCG Ala	GCG Ala	GGG Gly	GAC Asp 865	CAG Gln	GAC Asp	CGA Arg	GCA Ala	GAG Glu 870	GCC Ala	CCG Pro	AAG Lys		2762
GCG Ala	GAG Glu 875	AGC Ser	GGG Gly	GAG Glu	CCC Pro	GGT Gly 880	GCC Ala	CGG Arg	GAG Glu	GAG Glu	CGG Arg 885	CCG Pro	CGG Arg	CCG Pro	CAC His		2810
CGC Arg 890	AGC Ser	CAC His	AGC Ser	AAG Lys	GAG Glu 895	GCC Ala	GCG Ala	GGG Gly	CCC Pro	CCG Pro 900	GAG Glu	GCG Ala	CGG Arg	AGC Ser	GAG Glu 905		2858
CGC Arg	GGC Gly	CGA Arg	GGC Gly	CCA Pro 910	GGC Gly	CCC Pro	GAG Glu	GGC Gly	GGC Gly 915	CGG Arg	CGG Arg	CAC His	CAC His	CGG Arg 920	CGC Arg		2906
GGC Gly	TCC Ser	CCG Pro	GAG Glu 925	Glu	GCG Ala	GCC Ala	GAG Glu	CGG Arg 930	Glu	CCC	CGA Arg	CGC	CAC His 935	CGC Arg	GCG Ala		2954

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CAC His	CGG Arg	CAC His	GTI.	GAT Asp	CCG Pro	AGC Ser	AAG Lys 945	Glu	TGC Cys	GCC Ala	GGC Gly	GCC Ala 950	Lys	GGG Gly	GAG Glu	3002
CGG Ar g	Arg 955	ALC	G CGG Arg	CAC His	CGC Arg	GGC Gly 960	GGC Gly	CCC	CGA Arg	GCG	GGG Gly 965	Pro	CGG Arg	GAG Glu	GCG Ala	3050
GAG Glu 970	261	GGG	GAG Glu	GAG Glu	Pro 975	Ата	CGG Arg	CGG Ar g	CAC His	CGG Arg 980	Ala	CGG Arg	CAC	AAG Lys	GCG Ala 985	3098
CAG Gln	CCT Pro	GCT Ala	CAC His	GAG Glu 990	Ala	GTG Val	GAG Glu	AAG Lys	GAG Glu 995	ACC Thr	ACG Thr	GAG Glu	AAG Lys	GAG Glu 100	GCC Ala 0	3146
ACG Thr	GAG Glu	AAG Lys	GAG Glu 100	Ата	GAG Glu	ATA Ile	GTG Val	GAA Glu 101	Ala	GAC Asp	AAG Lys	GAA Glu	AAG Lys 101	Glu	CTC Leu	3194
CGG Arg	AAC Asn	CAC His 102	GIII	CCC	CGG Arg	GAG Glu	CCA Pro 102	Hls	TGT Cys	GAC Asp	CTG Leu	GAG Glu 103	Thr	AGT Ser	GGG Gly	3242
ACT Thr	GTG Val 103	THE	GTG Val	GGT Gly	CCC Pro	ATG Met 1040	His	ACA Thr	CTG Leu	CCC Pro	AGC Ser 1045	Thr	TGT Cys	CTC Leu	CAG Gln	3290
AAG Lys 1050	val	GAG Glu	GAA Glu	CAG Gln	CCA Pro 1055	GAG Glu	GAT Asp	GCA Ala	GAC Asp	AAT Asn 1060	Gln	CGG Arg	AAC Asn	GTC Val	ACT Thr 1065	3338
CGC Arg	ATG Met	GGC Gly	AGT Ser	CAG Gln 1070	Pro	CCA Pro	GAC Asp	CCG Pro	AAC Asn 1075	Thr	ATT Ile	GTA Val	CAT His	ATC Ile 1080	Pro	3386
GTG Val	ATG Met	CTG Leu	ACG Thr 1085	Gly	CCT Pro	CTT Leu	GGG Gly	GAA Glu 1090	Ala	ACG Thr	GTC Val	GTT Val	CCC Pro 1095	Ser	GGT Gly	3434
AAC Asn	GTG Val	GAC Asp 1100	Leu	GAA Glu	AGC Ser	Gln	GCA Ala 1105	Glu	GGG Gly	AAG Lys	Lys	GAG Glu 1110	Val	GAA Glu	GCG Ala	3482
GAT Asp	GAC Asp 1115	Val	ATG Met	AGG Arg	Ser	GGC Gly 1120	CCC Pro	CGG Arg	CCT Pro	Ile	GTC Val 1125	CCA Pro	TAC Tyr	AGC Ser	TCC Ser	3530
ATG Met 1130	Pne	TGT Cys	TTA Leu	Ser	CCC Pro 1135	Thr .	AAC Asn	CTG Leu	Leu	CGC Arg 1140	Arg	TTC Phe	TGC Cys	CAC His	TAC Tyr 1145	3578
ATC	GTG Val	ACC Thr	Met	AGG Arg 1150	Tyr	TTC (Phe (GAG Glu	Val	GTC Val 1155	ATT Ile	CTC (Leu	GTG Val	Val	ATC Ile 1160	Ala	3626

TTG AGC AGC Leu Ser Ser	ATC GCC CTG Ile Ala Leu 1165	Ala Ala	GAG GAC Glu Asp 1170	CCA GTG Pro Val	CGC ACA Arg Thr 1175	Asp Se	CG 3674 er
CCC AGG AAC Pro Arg Asn 1180	Asn Ala Leu	AAA TAC (Lys Tyr : 1185	Leu Asp	TAC ATT Tyr Ile	TTC ACT Phe Thr 1190	GGT G1 Gly Va	CC 3722
TTT ACC TTT Phe Thr Phe 1195	GAG ATG GTG Glu Met Val	ATA AAG I Ile Lys I 1200	ATG ATC Met Ile	GAC TTG Asp Leu 1205	Gly Leu	CTG CT Leu Le	TT 3770 eu
CAC CCT GGA His Pro Gly 1210	GCC TAT TTC Ala Tyr Phe 121	Arg Asp	TTG TGG Leu Trp	AAC ATT Asn Ile 1220	CTG GAC Leu Asp	Phe Il	TT 3818 Le 225
GTG GTC AGT Val Val Ser	GGC GCC CTG Gly Ala Leu 1230	GTG GCG 'Val Ala	TTT GCT Phe Ala 1235	Phe Ser	Gly Ser	AAA GO Lys Gl 1240	3866 Ly
AAA GAC ATC Lys Asp Ile	AAT ACC ATC Asn Thr Ile 1245	Lys Ser	CTG AGA Leu Arg 1250	GTC CTT Val Leu	CGT GTC Arg Val 1255	Leu Ai	GG 3914 CG
CCC CTC AAG Pro Leu Lys 1260	Thr Ile Lys	CGG CTG Arg Leu 1265	Pro Lys	CTC AAG Leu Lys	GCT GTG Ala Val 1270	TTT GA	AC 3962 sp
TGT GTG GTG Cys Val Val 1275	AAC TCC CTG Asn Ser Leu	AAG AAT Lys Asn 1280	GTC CTC Val Leu	AAC ATC Asn Ile 1285	Leu Ile	GTC TI Val T	AC 4010 yr
ATG CTC TTC Met Leu Phe 1290	ATG TTC ATA Met Phe Ile 129	Phe Ala	GTC ATT Val Ile	GCG GTG Ala Val 1300	CAG CTC Gln Leu	Phe Ly	AA 4058 ys 305
GGG AAG TTT Gly Lys Phe	TTC TAC TGC Phe Tyr Cys 1310	ACA GAT Thr Asp	GAA TCC Glu Ser 131	Lys Glu	CTG GAG Leu Glu	AGG G Arg A 1320	AC 4106 sp
TGC AGG GGT Cys Arg Gly	CAG TAT TTG Gln Tyr Leu 1325	GAT TAT Asp Tyr	GAG AAG Glu Lys 1330	GAG GAA Glu Glu	GTG GAA Val Glu 1335	ALA G	AG 4154 ln
CCC AGG CAG Pro Arg Gln 134	TGG AAG AAA Trp Lys Lys 0	TAC GAC Tyr Asp 1345	Phe His	TAC GAC Tyr Asp	AAT GTG Asn Val 1350	CTC To Leu T	GG 4202 rp
GCT CTG CTG Ala Leu Leu 1355	ACG CTG TTC Thr Leu Phe	ACA GTG Thr Val 1360	TCC ACG Ser Thr	GGA GAA Gly Glu 136	Gly Trp	CCC A Pro M	TG 4250 et
GTG CTG AAA Val Leu Lys 1370	CAC TCC GTG His Ser Val	Asp Ala	ACC TAT Thr Tyr	GAG GAG Glu Glu 1380	CAG GGT Gln Gly	Pro S	GC 4298 er 385

CCT Pro	GGG Gly	TAC Tyr	CGC Arg	ATG Met 139	Glu	CTG Leu	TCC Ser	ATC Ile	TTC Phe 139	Tyr	GTG Val	GTC Val	TAC Tyr	TTT Phe 140		4346
GTC Val	TTT Phe	CCC Pro	TTC Phe 140	Phe	TTC Phe	GTC Val	AAC Asn	ATC Ile 1410	Phe	GTG Val	GCT Ala	TTG Leu	ATC Ile 141	Ile	ATC Ile	4394
ACC Thr	TTC Phe	CAG Gln 1420	Glu	CAG Gln	GGG Gly	GAC Asp	AAG Lys 142	Val	ATG Met	TCT Ser	GAA Glu	TGC Cys 1430	Ser	CTG Leu	GAG Glu	4442
AAG Lys	AAC Asn 1435	GAG Glu	AGG Arg	GCT Ala	TGC Cys	ATT Ile 1440	Asp	TTC Phe	GCC Ala	ATC Ile	AGC Ser 144	Ala	AAA Lys	CCC Pro	CTG Leu	4490
ACA Thr 1450	Arg	TAC Tyr	ATG Met	CCC Pro	CAA Gln 1455	Asn	CGG Arg	CAG Gln	TCG Ser	TTC Phe 1460	Gln	TAT Tyr	AAG Lys	ACG Thr	TGG Trp 1465	4538
ACA Thr	TTT Phe	GTG Val	GTC Val	TCC Ser 1470	Pro	CCC Pro	TTT Phe	GAA Glu	TAC Tyr 1475	Phe	ATC Ile	ATG Met	GCC Ala	ATG Met 1480	Ile	4586
GCC Ala	CTC Leu	AAC Asn	ACT Thr 1485	Val	GTG Val	CTG Leu	ATG Met	ATG Met 1490	Lys	TTC Phe	TAT Tyr	GAT Asp	GCA Ala 1495	Pro	TAT Tyr	4634
GAG Glu	TAC Tyr	GAG Glu 1500	Leu	ATG Met	CTG Leu	AAA Lys	TGC Cys 1505	Leu	AAC Asn	ATC Ile	GTG Val	TTC Phe 1510	Thr	TCC Ser	ATG Met	4682
TTC Phe	TCC Ser 1515	ATG Met	GAA Glu	TGC Cys	GTG Val	CTG Leu 1520	Lys	ATC Ile	ATC Ile	GCC Ala	TTT Phe 1525	Gly	GTG Val	CTG Leu	AAC Asn	4730
TAT Tyr 1530	Phe	AGA Arg	GAT Asp	GCC Ala	TGG Trp 1535	Asn	GTC Val	TTT Phe	GAC Asp	TTT Phe 1540	Val	ACT Thr	GTG Val	TTG Leu	GGA Gly 1545	4778
AGT Ser	ATT Ile	ACT Thr	GAT Asp	ATT Ile 1550	Leu	GTA Val	ACA Thr	GAG Glu	ATT Ile 1555	Ala	GAA Glu	ACG Thr	AAC Asn	AAT Asn 1560	Phe	4826
		CTC Leu		Phe					Arg					Ile		4874
		CGC Arg 1580	Gln			Thr		Arg					Thr			4922
		TTC Phe					Tyr					Ile				4970

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TTC Phe	Phe	Ile	Tyr	Ala	Ile 1615	Ile	Gly	Met	GIN	1620	Pne	GIY	ASII	116	1625	5018
CTG :	GAT Asp	GAT Asp	GAC Asp	ACC Thr 1630	Ser	ATC Ile	AAC Asn	CGC Arg	CAC His 1635	HPII	AAC Asn	TTC Phe	CGG Arg	ACG Thr 1640		5066
TTG Leu	CAA Gln	GCC Ala	CTG Leu 1645	Met	CTG Leu	CTG Leu	TTC Phe	AGG Arg 1650	ser	GCC Ala	ACG Thr	GGG Gly	GAG Glu 1655	WIG	TGG Trp	5114
CAC His	GAG Glu	ATC Ile 1660	Met	CTG Leu	TCC Ser	TGC Cys	CTG Leu 166	Ser	AAC Asn	CAG Gln	GCC Ala	TGT Cys 1670	Map	GAG Glu	CAG Gln	5162
Ala	AAT Asn 167	Ala	ACC Thr	GAG Glu	TGT Cys	GGA Gly 1680	ser	GAC Asp	TTT Phe	GCC Ala	TAC Tyr 168	FILE	TAC Tyr	TTC Phe	GTC Val	5210
TCC Ser 1690	Phe	ATC Ile	TTC Phe	CTG Leu	TGC Cys 169	Ser	TTT Phe	CTG Leu	ATG Met	TTG Leu 1700	Asn	CTC Leu	TTT Phe	GTG Val	GCT Ala 1705	5258
GTG Val	ATC Ile	ATG Met	GAC Asp	AAT Asn 171	Phe	GAG Glu	TAC Tyr	CTC Leu	ACG Thr 171	CGG Arg 5	GAC Asp	TCT Ser	TCC	ATC Ile 172	шеи	5306
GGT Gly	CCT Pro	CAC His	CAC His 172	Leu	GAT Asp	GAG Glu	TTC Phe	ATC Ile 173	Arg	GTC Val	TGG Trp	GCT Ala	GAA Glu 173		GAC Asp	5354
CCG Pro	GCT Ala	GCG Ala 174	Cys	GGG Gly	CGC Arg	ATC Ile	AGT Ser 174	TYL	AAT Asn	GAC Asp	ATG Met	TTT Phe 175		ATG Met	CTG Leu	5402
AAA Lys	CAC His	Met	TCC	CCG	CCT Pro	CTG Leu 176	GLY	CTG Leu	GGG Gly	AAG Lys	AAA Lys 176	- Cys	CCT Pro	GCT Ala	CGA Arg	5450
GTT Val 177	Ala	TAC	: AAG : Lys	CGC	CTG Leu 177	ı vaı	CGC	ATG Met	AAC Asr	ATG Met 178		ATC Ile	TCC Ser	AAC Asn	GAG Glu 1785	5498
GAC Asp	ATC Met	ACI Thr	GTI Val	CAC His	s Phe	ACG Thr	TCC Sei	C ACC	CTC Lev 179	1 Mec	GCC Ala	CTC Lev	ATC	CGC Arg 180	ACG Thr	5546
GCA Ala	CTC	GAC	ATC 1 Ile 180	E Ly	G CT(GCC Ala	C CC	A GCT 5 Ala 18:	2 61	G ACA y Thi	A AAC Lys	G CAC	CAT His 183	CAC Gli L5	TGT Cys	5594
GAC Asp	GCC Ala	G GAG a Gli 18:	ı Leı	AG Ar	g AA(G GA(3 AT 1 Il 18	e 5e.	C GT r Va	r GTO l Val	TG(G GCC p Ala 18:		r CT	G CCC u Pro	5642

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CA(Gl:	G AA n Ly 18		T TT r Le	G GA u Ası	C TTO P Lev	G CTC 1 Leu 184	va.	A CCA	A CC	C CA	r AAG s Ly: 184	s Pro	T GA	T GA p Gl	G ATG u Met	5690
ACI Thi 185	A GTO C Va;	G GG l Gl	G AAG y Lya	G GT1 s Val	TA1 Ty1 185	. Ala	GCT Ala	CTO Lev	ATO Met	3 ATA 116 186	≥ Pne	GA(C TTO Pho	C TA e Ty	C AAG r Lys 186	
		- - ,		187	0	Asp	GII	. Met	187	i Glr 75	1 Ala	Pro	Gly	/ Gl;		5786
			188	5	• • • • • • • • • • • • • • • • • • • •	DEI	neu	189	0	PIC	. ren	Lys	189	Th:	C CTG r Leu	5834
GAG Glu	Gln	ACI Thi		CCG Pro	GCT Ala	GTG Val	CTC Leu 190	Arg	GGA Gly	GCC Ala	CGG	GTT Val 191	Phe	CTT Let	CGA Arg	5882
	191	5			261	1920	o O	ASI	GIY	GIĀ	A1a 192	Ile 5	Gln	Asr	CAA Gln	5930
GAG Glu 193		GGC	ATC Ile	AAA Lys	GAG Glu 193	Set	GTC Val	TCC Ser	TGG Trp	GGC Gly 194	Thr	CAA Gln	AGG Arg	ACC Thr	CAG Gln 1945	5978
			1112	1950)	Arg	Pro	Pro	Leu 195	Glu 5	Arg	Gly	His	Ser 196	0	6026
GAG Glu	ATC Ile	CCT Pro	GTG Val 196	GTÀ	CGG Arg	TCA Ser	GGA Gly	GCA Ala 1970	ren	GCT Ala	GTG Val	GAC Asp	GTT Val 197	Gln	ATG Met	6074
CAG Gln	AGC Ser	ATA Ile 198	ACC Thr	CGG Arg	AGG Arg	GIY	CCT Pro 1985	Asp	GGG Gly	GAG Glu	CCC Pro	CAG Gln 1990	Pro	GGG Gly	CTG Leu	6122
GAG Glu	AGC Ser 1995	GTU	GGT Gly	CGA Arg	Ala	GCC Ala 2000	Ser	ATG Met	CCC Pro	CGC Arg	CTT Leu 2005	Ala	GCC Ala	GAG Glu	ACT Thr	6170
CAG Gln 2010	PIO	GTC Val	ACA Thr	Asp	GCC Ala 2015	AGC Ser	CCC Pro	ATG Met	Lys	CGC Arg 2020	Ser	ATC Ile	TCC Ser	ACG Thr	CTG Leu 2025	6218
GCC Ala	CAG Gln	CGG Arg	CCC Pro	CGT Arg 2030	GTA .	ACT (CAT His	Leu	TGC Cys 2035	Ser	ACC Thr	ACC Thr	CCG Pro	GAC Asp 2040	Arg	6266
CCA Pro	CCC Pro	CCT Pro	AGC Ser 2045	Gin .	GCG ' Ala :	TCG : Ser :	Ser 1	CAC (His] 2050	CAC His :	CAC His	CAC His	His ?	CGC Arg 2055	Cys	CAC His	6314

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Arg Arg Asp Arg Lys Gln Arg Ser Leu Glu Lys Gly Pro Ser Leu 2060 2065 2070	52
TCT GCC GAT ATG GAT GGC GCA CCA AGC AGT GCT GTG GGG CCG GGG CTG Ser Ala Asp Met Asp Gly Ala Pro Ser Ser Ala Val Gly Pro Gly Leu 2075 2080 641	LO
CCC CCG GGA GAG GGG CCT ACA GGC TGC CGG CGG GAA CGA GAG CGC CGG Pro Pro Gly Glu Gly Pro Thr Gly Cys Arg Arg Glu Arg Glu Arg Arg 2090 2100 2105	58
CAG GAG CGG GGC CGG TCC CAG GAG CGG AGG CAG CCC TCA TCC TCC TCC Gln Glu Arg Gly Arg Ser Gln Glu Arg Gln Pro Ser Ser Ser Ser 2110 2115 2120)6
TCG GAG AAG CAG CGC TTC TAC TCC TGC GAC CGC TTT GGG GGC CGT GAG Ser Glu Lys Gln Arg Phe Tyr Ser Cys Asp Arg Phe Gly Gly Arg Glu 2125 2130 2135	54
CCC CCG AAG CCC AAG CCC TCC CTC AGC AGC CAC CCA ACG TCG CCA ACA Pro Pro Lys Pro Lys Pro Ser Leu Ser Ser His Pro Thr Ser Pro Thr 2140 2145 2150	02
GCT GGC CAG GAG CCG GGA CCC CAC CCA CAG GCC GGC TCA GCC GTG GGC Ala Gly Gln Glu Pro Gly Pro His Pro Gln Ala Gly Ser Ala Val Gly 2155 2160 2165	50
TTT CCG AAC ACA ACG CCC TGC TGC AGA GAG ACC CCC TCA GCC AGC CCC Phe Pro Asn Thr Thr Pro Cys Cys Arg Glu Thr Pro Ser Ala Ser Pro 2170 2180 2185	98
TGG CCC CTG GCT CTC GAA TTG GCT CTG ACC CTT ACC TGG GGC AGC GTC Trp Pro Leu Ala Leu Glu Leu Ala Leu Thr Leu Thr Trp Gly Ser Val 2190 2195 2200	46
TGG ACA GTG AGG CCT CTG TCC ACG CCC TGC CTG AGG ACA CGC TCA CTT Trp Thr Val Arg Pro Leu Ser Thr Pro Cys Leu Arg Thr Arg Ser Leu 2205 2210 2215	94
TCG AGG AGG CTG TGG CCA CCA ACT CGG GCC GCT CCT CCA GGA CTT CCT Ser Arg Arg Leu Trp Pro Pro Thr Arg Ala Ala Pro Pro Gly Leu Pro 2220 2225 2230 .	342
ACG TGT CCT CCC TGACCTCCCA GTCTCACCCT CTCCGCCGCG TGCCCAACGG 68 Thr Cys Pro Pro 2235	394
TTACCACTGC ACCCTGGGAC TCAGCTCGGG TGGCCGAGCA CGGCACAGCT ACCACCACCC 69	954
	014
	07
	13

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AG	ACG/	ACGAZ	TAI	AGCC	CTG	TTG	TTGA	GT G	TAC	TACC	G C						7185
				N FC													7175
			EQUE (A) (B) (C)	NCE LENG TYPE STRA TOPO	CHAR TH: : nu NDED	ACTE 1546 clei NESS	RIST bas c ac	ICS: e pa id uble	irs								
	(i	i) M	OLEC	ULE	TYPE	: DN	A (g	enom	ic)								
	(i	x) F	EATU (A) (B)	RE : NAME LOCA'	/KEY TION	: CD	S .143'	7									
	(i:		EATU (A) I (B) I	RE: NAME, LOCA:	/KEY FION	: 3'1 : 14:	UTR 35:	L546									
	(x:	i) s	EQUE	NCE I	DESCI	RIPT	ON:	SEQ	ID 1	10:9:	:						
1			_,	5	5		. 561	. MI	10	PIC) 1 <u>y</u> 1	Pro	Pro	Ser 15			48
			20				. wat	25	Ser	Pro	GII	Gly	30 30	Туг	AGC Ser		96
_	_	35	1	-43	- 110	Llys	40	ser	Asp	GIA	Ser	Thr 45	Ser	Ser	GAT Asp		144
	50			JC1	rne	55	Arg	GIN	GIÀ	ser	Ala 60	Glu	Ser	Tyr	ACC Thr		192
AGC Ser 65	CGT Arg	CCA Pro	TCA Ser	GAC Asp	TCT Ser 70	GAT Asp	GTA Val	TCT Ser	CTG Leu	GAG Glu 75	GAG Glu	GAC Asp	CGG Arg	GAA Glu	GCC Ala 80	:	240
TTA Leu	AGG Arg	AAG Lys	GAA Glu	GCA Ala 85	GAG Glu	CGC Arg	CAG Gln	GCA Ala	TTA Leu 90	GCG Ala	CAG Gln	CTC Leu	GAG Glu	AAG Lys 95	GCC Ala		288
AAG Lys	ACC Thr	AAG Lys	CCA Pro 100	GTG Val	GCA Ala	TTT Phe	GCT Ala	GTG Val 105	CGG Arg	ACA Thr	AAT Asn	GTT Val	GGC Gly 110	TAC Tyr	AAT Asn	3	336
Pro	TCT Ser	CCA Pro 115	GGG Gly	GAT Asp	GAG Glu	GTG Val	CCT Pro 120	GTG Val	CAG Gln	GGA Gly	GTG Val	GCC Ala 125	ATC Ile	ACC Thr	TTC Phe	3	84

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														C 3 C	TCC	4	32
GAG Glu	CCC Pro 130	AAA Lys	GAC Asp	TTC Phe	Leu	CAC His 135	ATC Ile	AAG Lys	GAG Glu	AAA Lys	TAC Tyr 140	TAA Asn	AAT Asn	Asp	Trp	•	32
TGG Trp 145	ATC Ile	GGG Gly	CGG Arg	CTG Leu	GTG Val 150	AAG Lys	GAG Glu	GGC Gly	TGT Cys	GAG Glu 155	GTT Val	GGC Gly	TTC Phe	ATT Ile	CCC Pro 160	4	80
AGC Ser	CCC Pro	GTC Val	AAA Lys	CTG Leu 165	GAC Asp	AGC Ser	CTT Leu	CGC Arg	CTG Leu 170	CTG Leu	CAG Gln	GAA Glu	CAG Gln	AAG Lys 175	CTG Leu	5	528
CGC Arg	CAG Gln	AAC Asn	CGC Arg 180	CTC Leu	GGC Gly	TCC Ser	AGC Ser	AAA Lys 185	TCA Ser	GGC Gly	GAT Asp	AAC Asn	TCC Ser 190	AGT Ser	TCC Ser	S	576
AGT Ser	CTG Leu	GGA Gly 195	GAT Asp	GTG Val	GTG Val	ACT Thr	GGC Gly 200	ACC Thr	CGC Arg	CGC Arg	CCC Pro	ACA Thr 205	CCC Pro	CCT Pro	GCC Ala	6	524
AGT Ser	GCC Ala 210	Lys	CAG Gln	AAG Lys	CAG Gln	AAG Lys 215	TCG Ser	ACA Thr	GAG Glu	CAT His	GTG Val 220	CCC Pro	CCC Pro	TAT Tyr	GAC Asp		672
GTG Val 225	Val	CCT Pro	TCC Ser	ATG Met	AGG Arg 230	CCC Pro	ATC Ile	ATC Ile	CTG Leu	GTG Val 235	GGA Gly	CCG Pro	TCG Ser	CTC Leu	AAG Lys 240	•	720
GGC	TAC	GAG Glu	GTT Val	ACA Thr 245	Asp	ATG Met	ATG Met	CAG Gln	AAA Lys 250	ATG	TTA Leu	TTT Phe	GAC Asp	TTC Phe 255			768
AAC Lys	CAT His	CGG Arg	TTI Phe	Asp	GGC Gly	AGG Arg	ATC	TCC Ser 265	116	ACT Thr	CGT	GTG Val	ACG Thr 270		GAT Asp		816
ATT Ile	TCC Ser	CTC Lev	ı Ala	AAG Lys	CGC Arg	TCA Ser	GTT Val	Let	AAC Asn	AAC Asn	CCC Pro	AGC Ser 285	-7-	CAC His	ATC		864
ATC Ile	ATT = 11e	e Glı	G CG(TCC Ser	AAC Asr	ACA Thr 295	Arg	TCC Sei	AGC Sei	CTC Lev	GCT 1 Ala 300		GTG Val	Glr	AGT Ser		912
GA: Gl: 30	ı Ile	C GA	G CG	A ATO	TT(e Phe 31(e GIV	CTO	G GCC	C CGG	ACC Thi		r CAC	TTC Lev	GT(GCT L Ala 320		960
•		T GC p Al	T GA	C ACC p Th: 32	ril	C AA' e Ası	r CA	c cc s Pr	A GCO Ala 33	u	G CT n Le	G TCC u Se:	C AAG	G ACC S Th: 33	TCG r Ser		1008
CT Le	G GC u Al	C CC a Pr	C AT o Il 34	e 11	T GT e Va	T TAG	C AT	C AA e Ly 34	3 11	C AC e Th	C TC r Se	T CC	C AA o Ly 35		A CTT l Leu		1056

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CAA Gln	AGG Arg	CTC Leu 355	ATC Ile	AAG Lys	TCC Ser	CGA Arg	GGA Gly 360	AAG Lys	TCT Ser	CAG Gln	TCC Ser	AAA Lys 365	CAC His	CTC Leu	AAT Asn	1	104
GTC Val	CAA Gln 370	ATA Ile	GCG Ala	GCC Ala	TCG Ser	GAA Glu 375	AAG Lys	CTG Leu	GCA Ala	CAG Gln	TGC Cys 380	CCC Pro	CCT Pro	GAA Glu	ATG Met	1:	152
TTT Phe 385	GAC Asp	ATC Ile	ATC Ile	CTG Leu	GAT Asp 390	GAG Glu	AAC Asn	CAA Gln	TTG Leu	GAG Glu 395	GAT Asp	GCC Ala	TGC Cys	GAG Glu	CAT His 400	12	200
CTG Leu	GCG Ala	GAG Glu	TAC Tyr	TTG Leu 405	GAA Glu	GCC Ala	TAT Tyr	TGG Trp	AAG Lys 410	GCC Ala	ACA Thr	CAC His	CCG Pro	CCC Pro 415	AGC Ser	12	248
AGC Ser	ACG Thr	CCA Pro	CCC Pro 420	AAT Asn	CCG Pro	CTG Leu	CTG Leu	AAC Asn 425	CGC Arg	ACC Thr	ATG Met	GCT Ala	ACC Thr 430	GCA Ala	GCC Ala	12	296
CTG Leu	GCT Ala	GCC Ala 435	AGC Ser	CCT Pro	GCC Ala	CCT Pro	GTC Val 440	TCC Ser	AAC Asn	CTC Leu	CAG Gln	GTA Val 445	CAG Gln	GTG Val	CTC Leu	13	44
Thr	TCG Ser 450	CTC Leu	AGG Arg	AGA Arg	AAC Asn	CTC Leu 455	GGC Gly	TTC Phe	TGG Trp	GGC Gly	GGG Gly 460	CTG Leu	GAG Glu	TCC Ser	TCA Ser	13	92
CAG Gln 465	CGG Arg	GGC Gly	AGT Ser	Val	GTG Val 470	CCC Pro	CAG Gln	GAG Glu	CAG Gln	GAA Glu 475	CAT His	GCC Ala	ATG Met	TAGT	GGGCGC	14	44
CCTG	CCCG	TC I	TCCC	TCCT	G CI	'CTGG	GGTC	GGA	ACTG	GAG	TGCA	.GGGA	AC A	TGGA	GGAGG	15	04
AAGG	GAAG	AG C	TTTA	TTTT	G TA	AAAA	ATAA	AGA	TGAG	CGG	CA					15	46
(2)	INFO	тамя	TON	FOR	SEO.	TD N	0.10										

INFORMATION FOR SEQ ID NO:10:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1851 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double

 - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (ix) FEATURE:

 - (A) NAME/KEY: CDS
 (B) LOCATION: 1..1797
 (D) OTHER INFORMATION: /standard_name= "Betal-3"
- (ix) FEATURE:

 - (A) NAME/KEY: 3'UTR (B) LOCATION: 1795..1851
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:10:

ATG Met 1	GTC Val	CAG Gln	AAG Lys	ACC Thr 5	AGC Ser	ATG Met	TCC Ser	CGG Arg	GGC Gly 10	CCT Pro	TAC Tyr	CCA Pro	CCC Pro	TCC Ser 15	CAG Gln	48
GAG Glu	ATC Ile	CCC Pro	ATG Met 20	GGA Gly	GTC Val	TTC Phe	GAC Asp	CCC Pro 25	AGC Ser	CCG Pro	CAG Gln	GGC Gly	AAA Lys 30	TAC Tyr	AGC Ser	. 96
AAG Lys	AGG Arg	AAA Lys 35	GGG Gly	CGA Arg	TTC Phe	AAA Lys	CGG Arg 40	TCA Ser	GAT Asp	GGG Gly	AGC Ser	ACG Thr 45	TCC Ser	TCG Ser	GAT Asp	144
ACC Thr	ACA Thr 50	TCC Ser	AAC Asn	AGC Ser	TTT Phe	GTC Val 55	CGC Arg	CAG Gln	Gly	TCA Ser	GCG Ala 60	GAG Glu	TCC Ser	TAC Tyr	ACC Thr	192
AGC Ser 65	CGT Arg	CCA Pro	TCA Ser	GAC Asp	TCT Ser 70	GAT Asp	GTA Val	TCT Ser	CTG Leu	GAG Glu 75	GAG Glu	GAC Asp	CGG Arg	GAA Glu	GCC Ala 80	2.40
TTA Leu	AGG Arg	AAG Lys	GAA Glu	GCA Ala 85	GAG Glu	CGC Arg	CAG Gln	GCA Ala	TTA Leu 90	GCG Ala	CAG Gln	CTC Leu	GAG Glu	AAG Lys 95	GCC Ala	288
AAG Lys	ACC Thr	AAG Lys	CCA Pro 100	GTG Val	GCA Ala	TTT Phe	GCT Ala	GTG Val 105	CGG Arg	ACA Thr	AAT Asn	GTT Val	GGC Gly 110	TAC Tyr	AAT Asn	336
CCG Pro	TCT Ser	CCA Pro 115	GGG Gly	GAT Asp	GAG Glu	GTG Val	CCT Pro 120	GTG Val	CAG Gln	GGA Gly	GTG Val	GCC Ala 125	ATC Ile	ACC Thr	TTC Phe	384
GAG Glu	CCC Pro 130	Lys	GAC Asp	TTC Phe	CTG Leu	CAC His 135	Ile	AAG Lys	GAG Glu	AAA Lys	TAC Tyr 140	Asn	AAT Asn	GAC Asp	TGG Trp	432
TGG Trp 145	ATC Ile	GGG Gly	CGG Arg	CTG Leu	GTG Val 150	Lys	GAG Glu	GGC Gly	TGT Cys	GAG Glu 155	GTT Val	GGC	TTC Phe	ATT	CCC Pro 160	480
AGC Ser	CCC Pro	GTC Val	AAA Lys	CTG Leu 165	GAC Asp	AGC Ser	CTT	CGC Arg	CTG Leu 170	reu	CAG Gln	GAA Glu	CAG Gln	AAG Lys 175	CTG Leu	528
CGC Arg	CAG Gln	AAC Asn	CGC Arg 180	Leu	GGC	TCC	AGC Ser	AAA Lys 185	Ser	GGC	GAT Asp	AAC Asn	TCC Ser 190	002	TCC	576
AGT Ser	CTG Leu	GGA Gly 195	Asp	GTG Val	GTG Val	ACI Thr	GGC Gly 200	Ini	CGC Arg	CGC Arg	CCC Pro	ACA Thr 205		CCT	GCC Ala	624
Ser	· Ala	Lys	CAG Gln	Lys	Glr	Lys	s Sei	ACA Thr	GAC Glu	CAT His	GTG Val	PIC	CCC Pro	TAT Tyr	GAC Asp	673

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GTG Val 225	val	CCI Pro	TCC Ser	ATG Met	AGG Arg 230	Pro	ATC	ATC	CTG Leu	GTG Val 235	Gly	CCG Pro	TCG Ser	CTC	AAG Lys 240	720
GGC Gly	TAC Tyr	GAG Glu	GTT Val	ACA Thr 245	GAC Asp	ATG Met	ATG Met	CAG Gln	AAA Lys 250	Ala	TTA Leu	TTT Phe	GAC Asp	TTC Phe 255	TTG Leu	768
AAG Lys	CAT His	CGG Arg	TTT Phe 260	GAT Asp	GGC Gly	AGG Arg	ATC Ile	TCC Ser 265	Ile	ACT Thr	CGT Arg	GTG Val	ACG Thr 270	Ala	GAT Asp	816
ATT	TCC	CTG Leu 275	GCT Ala	AAG Lys	CGC	TCA Ser	GTT Val 280	CTC Leu	AAC Asn	AAC Asn	CCC Pro	AGC Ser 285	AAA Lys	CAC His	ATC Ile	864
ATC Ile	ATT Ile 290	GAG Glu	CGC	TCC Ser	AAC Asn	ACA Thr 295	CGC Arg	TCC Ser	AGC Ser	CTG Leu	GCT Ala 300	GAG Glu	GTG Val	CAG Gln	AGT Ser	912
GAA Glu 305	ATC Ile	GAG Glu	CGA Arg	ATC Ile	TTC Phe 310	GAG Glu	CTG Leu	GCC Ala	CGG Arg	ACC Thr 315	CTT Leu	CAG Gln	TTG Leu	GTC Val	GCT Ala 320	960
CTG Leu	GAT Asp	GCT Ala	GAC Asp	ACC Thr 325	ATC Ile	AAT Asn	CAC His	CCA Pro	GCC Ala 330	CAG Gln	CTG Leu	TCC Ser	AAG Lys	ACC Thr 335	TCG Ser	1008
CTG Leu	GCC Ala	CCC Pro	ATC Ile 340	ATT Ile	GTT Val	TAC Tyr	ATC Ile	AAG Lys 345	ATC Ile	ACC Thr	TCT Ser	CCC Pro	AAG Lys 350	GTA Val	CTT Leu	1056
CAA Gln	AGG Arg	CTC Leu 355	ATC Ile	AAG Lys	TCC Ser	CGA Arg	GGA Gly 360	AAG Lys	TCT Ser	CAG Gln	TCC Ser	AAA Lys 365	CAC His	CTC Leu	AAT Asn	1104
GTC Val	CAA Gln 370	ATA Ile	GCG Ala	GCC Ala	TCG Ser	GAA Glu 375	AAG Lys	CTG Leu	GCA Ala	CAG Gln	TGC Cys 380	CCC Pro	CCT Pro	GAA Glu	ATG Met	1152
TTT Phe 385	GAC Asp	ATC Ile	ATC Ile	CTG Leu	GAT Asp 390	GAG Glu	AAC Asn	CAA Gln	TTG Leu	GAG Glu 395	GAT Asp	GCC Ala	TGC Cys	GAG Glu	CAT His 400	1200
CTG Leu	GCG Ala	GAG Glu	TAC Tyr	TTG Leu 405	GAA Glu	GCC Ala	TAT Tyr	TGG Trp	AAG Lys 410	GCC Ala	ACA Thr	CAC His	CCG Pro	CCC Pro 415	AGC Ser	1248
AGC Ser	ACG Thr	CCA Pro	CCC Pro 420	AAT Asn	CCG Pro	CTG Leu	Leu	AAC Asn 425	CGC Arg	ACC Thr	ATG Met	Ala	ACC Thr 430	GCA Ala	GCC Ala	1296
CTG Leu	Ala	GCC Ala 435	AGC Ser	CCT Pro	GCC Ala	Pro	GTC Val 440	TCC Ser	AAC Asn	CTC Leu	CAG Gln	GGA Gly 445	CCC Pro	TAC Tyr	CTT Leu	1344

GCT A la	TCC Ser 450	GGG Gly	GAC Asp	CAG Gln	CCA Pro	CTG Leu 455	GAA Glu	CGG Arg	GCC Ala	ACC Thr	GGG Gly 460	GAG Glu	CAC His	GCC Ala	AGC Ser	1392
ATG Met 465	CAC His	GAG Glu	TAC Tyr	CCA Pro	GGG Gly 470	GAG Glu	CTG Leu	GGC Gly	CAG Gln	CCC Pro 475	CCA Pro	GGC Gly	CTT Leu	TAC Tyr	CCC Pro 480	1440
AGC Ser	AGC Ser	CAC His	CCA Pro	CCA Pro 485	GGC Gly	CGG Arg	GCA Ala	GGC Gly	ACG Thr 490	CTA Leu	CGG Arg	GCA Ala	CTG Leu	TCC Ser 495	CGC Arg	1488
CAA Gln	GAC Asp	ACT Thr	TTT Phe 500	GAT Asp	GCC Ala	GAC Asp	ACC Thr	CCC Pro 505	GGC Gly	AGC Ser	CGA Arg	AAC Asn	TCT Ser 510	GCC Ala	TAC Tyr	1536
ACG Thr	GAG Glu	CTG Leu 515	GGA Gly	GAC Asp	TCA Ser	TGT Cys	GTG Val 520	GAC Asp	ATG Met	GAG Glu	ACT Thr	GAC Asp 525	CCC Pro	TCA Ser	GAG Glu	1584
GGG Gly	CCA Pro 530	GGG Gly	CTT Leu	GGA Gly	GAC Asp	CCT Pro 535	GCA Ala	GGG Gly	GGC Gly	GGC Gly	ACG Thr 540	CCC Pro	CCA Pro	GCC Ala	CGA Arg	1632
CAG Gln 545	GGA Gly	TCC Ser	TGG Trp	GAG Glu	GAC Asp 550	GAG Glu	GAA Glu	GAA Glu	GAC Asp	TAT Tyr 555	GAG Glu	GAA Glu	GAG Glu	CTG Leu	ACC Thr 560	1680
GAC Asp	AAC Asn	CGG Arg	AAC Asn	CGG Arg 565	GGC Gly	CGG Arg	AAT Asn	AAG Lys	GCC Ala 570	CGC Arg	TAC Tyr	TGC Cys	GCT Ala	GAG Glu 575	GGT Gly	1728
GGG	GGT Gly	CCA Pro	GTT Val 580	Leu	GGG Gly	CGC Arg	AAC Asn	AAG Lys 585	AAT Asn	GAG Glu	CTG Leu	GAG Glu	GGC Gly 590	ııρ	GGA Gly	1776
CGA Arg	GGC Gly	GTC Val 595	TAC	ATT	CGC Arg	TGA	GAGG	CAG	GGGC	CACA	.CG G	CGGG	AGGA	A		1824
GGG	CTCT	GAG	CCCA	.GGGG	AG G	GGAG	GG									1851

(2) INFORMATION FOR SEQ ID NO:11:

- (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 3600 base pairs

 - (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (ix) FEATURE:
 - (A) NAME/KEY: CDS

 - (B) LOCATION: 35..3310
 (D) OTHER INFORMATION: /standard_name= "Alpha-2"

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	(i:		EATUI (A) 1 (B) 1	NAME,	KEY:	5′t	JTR .34									
	(i)	,	EATUR (A) 1 (B) I	IAME/	KEY:	3'E 330	TTR 083	600								
	(xi) SE	QUEN	ICE I	DESCR	IPTI	ON:	SEQ	ID N	0:11	. :					
GCG	GGGG	AGG	GGGC	ATTO	SAT C	TTCG	ATCG	ic ga	AG A	TG G let A	CT C	CT G	GC T	GC C ys I 5	TG eu	52
CTG Leu	GCC Ala	Leu	ACT Thr	Leu	ACA Thr	CTT Leu	TTC Phe	CAA Gln 15	Ser	TTG	CTC Leu	: ATC	GGC Gly 20	Pro	TCG Ser	100
TCG Ser	GAG Glu	GAG Glu 25	PIO	TTC Phe	CCT Pro	TCG Ser	GCC Ala 30	Val	ACT Thr	ATC Ile	AAA Lys	TCA Ser 35	TGG	GTG Val	GAT Asp	148
AAG Lys	ATG Met 40	GIN	GAA Glu	GAC Asp	CTT Leu	GTC Val 45	ACA Thr	CTG Leu	GCA Ala	AAA Lys	ACA Thr 50	GCA Ala	AGT Ser	GGA Gly	GTC Val	196
AAT Asn 55	CAG Gln	CTT Leu	GTT Val	GAT Asp	ATT Ile 60	TAT Tyr	GAG Glu	AAA Lys	TAT Tyr	CAA Gln 65	GAT Asp	TTG Leu	TAT Tyr	ACT Thr	GTG Val 70	244
GAA Glu	CCA Pro	AAT Asn	AAT Asn	GCA Ala 75	CGC Arg	CAG Gln	CTG Leu	GTA Val	GAA Glu 80	ATT Ile	GCA Ala	GCC Ala	AGG Arg	GAT Asp 85	ATT Ile	292
GAG Glu	AAA Lys	CTT Leu	CTG Leu 90	AGC Ser	AAC Asn	AGA Arg	TCT Ser	AAA Lys 95	GCC Ala	CTG Leu	GTG Val	AGC Ser	CTG Leu 100	GCA Ala	TTG Leu	340
GAA Glu	GCG Ala	GAG Glu 105	AAA Lys	GTT Val	CAA Gln	GCA Ala	GCT Ala 110	CAC His	CAG Gln	TGG Trp	AGA Arg	GAA Glu 115	GAT Asp	TTT Phe	GCA Ala	388
AGC Ser	AAT Asn 120	GAA Glu	GTT Val	GTC Val	TAC Tyr	TAC Tyr 125	AAT Asn	GCA Ala	AAG Lys	GAT Asp	GAT Asp 130	CTC Leu	GAT Asp	CCT Pro	GAG Glu	436
AAA Lys 135	AAT Asn	GAC Asp	AGT Ser	GAG Glu	CCA Pro 140	GGC Gly	AGC Ser	CAG Gln	AGG Arg	ATA Ile 145	AAA Lys	CCT Pro	GTT Val	TTC Phe	ATT Ile 150	484
GAA Glu	GAT Asp	GCT Ala	AAT Asn	TTT Phe 155	GGA Gly	CGA Arg	CAA Gln	ATA Ile	TCT Ser 160	TAT Tyr	CAG Gln	CAC His	GCA Ala	GCA Ala 165	GTC Val	532
CAT His	ATT Ile	CCT Pro	ACT Thr	GAC Asp	ATC Ile	TAT Tyr	GAG Glu	GGC Gly	TCA Ser	ACA Thr	ATT Ile	GTG Val	TTA Leu	AAT Asn	GAA Glu	580

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			170					175					180			
CTC Leu	AAC Asn	TGG Trp 185	ACA Thr	AGT Ser	GCC Ala	Leu	GAT Asp 190	GAA Glu	GTT Val	TTC Phe	AAA Lys	AAG Lys 195	AAT Asn	CGC Arg	GAG Glu	628
GAA Glu	GAC Asp 200	CCT Pro	TCA Ser	TTA Leu	TTG Leu	TGG Trp 205	CAG Gln	GTT Val	TTT Phe	GGC Gly	AGT Ser 210	GCC Ala	ACT Thr	GGC Gly	CTA Leu	676
GCT Ala 215	CGA Arg	TAT Tyr	TAT Tyr	CCA Pro	GCT Ala 220	TCA Ser	CCA Pro	TGG Trp	GTT Val	GAT Asp 225	AAT Asn	AGT Ser	AGA Arg	ACT Thr	CCA Pro 230	724
AAT Asn	AAG Lys	ATT Ile	GAC Asp	CTT Leu 235	TAT Tyr	GAT Asp	GTA Val	CGC Arg	AGA Arg 240	AGA Arg	CCA Pro	TGG Trp	TAC Tyr	ATC Ile 245	CAA Gln	772
GGA Gly	GCT Ala	GCA Ala	TCT Ser 250	CCT Pro	AAA Lys	GAC Asp	ATG Met	CTT Leu 255	ATT Ile	CTG Leu	GTG Val	GAT Asp	GTG Val 260	AGT Ser	GGA Gly	820
AGT Ser	GTT Val	AGT Ser 265	GGA Gly	TTG Leu	ACA Thr	CTT Leu	AAA Lys 270	CTG Leu	ATC Ile	CGA Arg	ACA Thr	TCT Ser 275	GTC Val	TCC Ser	GAA Glu	868
ATG Met	TTA Leu 280	Glu	ACC Thr	CTC Leu	TCA Ser	GAT Asp 285	GAT Asp	GAT Asp	TTC Phe	GTG Val	AAT Asn 290		GCT Ala	TCA Ser	TTT Phe	916
AAC Asn 295	AGC Ser		GCT Ala	CAG Gln	GAT Asp 300	vaı	AGC Ser	TGT Cys	TTT Phe	CAG Gln 305		CTT Leu	GTC Val	CAA Gln	GCA Ala 310	964
		AGA Arg	AAT Asn	AAA Lys 315	Lys	GTG Val	TTG Leu	AAA Lys	GAC Asp 320	, ATC	GTG Val	AAT Asn	TAA '	ATC Ile 325	ACA Thr	1012
GCC Ala	AAA Lys	GGA Gly	ATT / Ile	Thr	GAT Asp	TAT Tyr	AAG Lys	AAG Lys	. 613	TTT Phe	AGT Ser	r TTI	GCT Ala 340	TTI Phe	GAA Glu	1060
CA(G CTO	G CTT	CAA 1 12A 1		AAT Asi	GT7 Val	TCC Ser	MIG	GCA J Ala	AAA Asi	TG(AAS ASI 35!		ATT	ATT :	1108
AT(G CTI	A TTO		GAT ASI	GG/	A GG/ 7 Gly 36!	GI	A GAC	AG Ar	A GC	C CA a Gl: 37	G GAO n Gl	TA E	TT:	r AAC e Asn	115
AA Ly 37	A TA		T AAI n Ly	A GA' s As	F AAI p Lys	s ry	A GT	A CG	r GT. g Va	A TT 1 Ph 38	C AG e Ar 5	G TT g Ph	T TC	A GT' r Va	T GGT 1 Gly 390	120
		C AA s As	T TA n Ty	T GA	G AG	A GG g Gl	A CC	T AT	T CA e Gl	G TG n Tr	G AT	G GC	C TG a Cy	T GA s Gl	A AAC u Asn	125

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				39	5				40	0				40	5	
AA Ly	A GG s Gl	т та у ту	T TA' r Ty: 41	y.	T GA	A AT	r cc	T TC Se 41	r TT	r GG e Gl	T GC Y Ala	A AT	A AG e Ar	g Il	C AAT e Asn	1300
AC Th	T CA	G GA n Gl 42	2 -	r TT(G GA!	r GTT o Val	TTC L Let 430	7 61	A AGA Y Arg	A CCI	A ATO	GT: Val 435	l Le	A GC	A GGA a Gly	1348
	44)	- - ,.	, GII	· val	445	i iii	Tni	AST	ı Val	450	Leu)	ı As <u>ı</u>	Ala	A TTG a Leu	1396
455	5				460)	GIY	ını	Leu	465	Val	. Phe	Asr	ı Ile	A ACC Thr 470	1444
•			. 010	475	. Dys	1111	ASI	Leu	480	Asn	Gln	Leu	Ile	Let 485		1492
		1	490	nsp	val	261	neu	495	Asp	116	Lys	Arg	Leu 500	Thr	CCA Pro	1540
_		505	204	Cys	FIO	ASII	510	Tyr	Tyr	Phe	Ala	Ile 515	Asp	Pro	AAT Asn	1588
GGT Gly	TAT Tyr 520	GTT Val	TTA Leu	TTA Leu	CAT His	CCA Pro 525	AAT Asn	CTT Leu	CAG Gln	CCA Pro	AAG Lys 530	AAC Asn	CCC Pro	AAA Lys	TCT Ser	1636
535			Val	1111	540	GAT Asp	Pne	Leu	Asp	Ala 545	Glu	Leu	Glu	Asn	Asp 550	1684
	-1-		014	555	arg	AAT Asn	ьуѕ	Met	560	Asp	Gly	Glu	Ser	Gly 565	Glu	1732
AAA Lys	ACA Thr	TTC Phe	AGA Arg 570	ACT Thr	CTG Leu	GTT Val	AAA Lys	TCT Ser 575	CAA Gln	GAT Asp	GAG Glu	AGA Arg	TAT Tyr 580	ATT Ile	GAC Asp	1780
AAA Lys	GGA Gly	AAC Asn 585	AGG Arg	ACA Thr	TAC Tyr	ACA Thr	TGG Trp 590	ACA Thr	CCT Pro	GTC Val	AAT Asn	GGC Gly 595	ACA Thr	GAT Asp	TAC Tyr	1828
AGT Ser	TTG Leu 600	GCC Ala	TTG Leu	GTA Val	TTA Leu	CCA Pro 605	ACC Thr	TAC Tyr	AGT Ser	Phe	TAC Tyr 610	TAT Tyr	ATA Ile	AAA Lys	GCC Ala	1876
AAA Lys	CTA Leu	GAA Glu	GAG Glu	ACA . Thr	ATA Ile	ACT (CAG Gln	GCC Ala	AGA ' Arg !	TCA : Ser :	AAA Lys :	AAG Lys	GGC	AAA Lys	ATG Met	1924

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615					620					625					630		
AAG Lys	GAT Asp	TCG Ser	GAA Glu	ACC Thr 635	CTG Leu	AAG Lys	CCA Pro	GAT Asp	AAT Asn 640	TTT Phe	GAA Glu	GAA Glu	TCT Ser	GGC Gly 645	TAT Tyr	1972	2
ACA Thr	TTC Phe	ATA Ile	GCA Ala 650	CCA Pro	AGA Arg	GAT Asp	TAC Tyr	TGC Cys 655	AAT Asn	GAC Asp	CTG Leu	AAA Lys	ATA Ile 660	TCG Ser	GAT Asp	2020	D
AAT Asn	AAC Asn	ACT Thr 665	GAA Glu	TTT Phe	CTT Leu	TTA Leu	AAT Asn 670	TTC Phe	AAC Asn	GAG Glu	TTT Phe	ATT Ile 675	GAT Asp	AGA Arg	AAA Lys	2068	3
ACT Thr	CCA Pro 680	AAC Asn	AAC Asn	CCA Pro	TCA Ser	TGT Cys 685	AAC Asn	GCG Ala	GAT Asp	TTG Leu	ATT Ile 690	AAT Asn	AGA Arg	GTC Val	TTG Leu	2116	5
CTT Leu 695	GAT Asp	GCA Ala	GGC Gly	TTT Phe	ACA Thr 700	Asn	GAA Glu	CTT Leu	GTC Val	CAA Gln 705	AAT Asn	TAC Tyr	TGG Trp	AGT Ser	AAG Lys 710	2164	1
CAG Gln	AAA Lys	AAT Asn	ATC Ile	AAG Lys 715	GGA Gly	GTG Val	AAA Lys	GCA Ala	CGA Arg 720	TTT Phe	GTT Val	GTG Val	ACT Thr	GAT Asp 725	GGT Gly	2212	2
GGG Gly	ATT Ile	ACC Thr	AGA Arg 730	GTT Val	TAT Tyr	CCC Pro	AAA Lys	GAG Glu 735	GCT Ala	GGA Gly	GAA Glu	AAT Asn	TGG Trp 740	CAA Gln	GAA Glu	2260	D
AAC Asn	CCA Pro	GAG Glu 745	ACA Thr	TAT Tyr	GAG Glu	GAC Asp	AGC Ser 750	TTC Phe	TAT Tyr	AAA Lys	AGG Arg	AGC Ser 755	CTA Leu	GAT Asp	AAT Asn	230	В
GAT A sp	AAC Asn 760	TAT Tyr	GTT Val	TTC Phe	ACT Thr	GCT Ala 765	CCC Pro	TAC Tyr	TTT Phe	AAC Asn	AAA Lys 770	AGT Ser	GGA Gly	CCT Pro	GGT Gly	235	6
GCC Ala 775	TAT Tyr	GAA Glu	TCG Ser	GGC Gly	ATT Ile 780	ATG Met	GTA Val	AGC Ser	AAA Lys	GCT Ala 785	GTA Val	GAA Glu	ATA Ile	TAT Tyr	ATT Ile 790	240	4
Gln	GGG Gly	Lys	Leu	Leu 795	Lys	Pro	Ala	Val	Val 800	Gly	Ile	Lys	Ile	805	Val	245	2
AAT Asn	TCC Ser	TGG Trp	ATA Ile 810	GAG Glu	AAT Asn	TTC Phe	ACC Thr	AAA Lys 815	ACC Thr	TCA Ser	ATC Ile	AGA Arg	GAT Asp 820	CCG Pro	TGT Cys	250	0
GCT Ala	GGT Gly	CCA Pro 825	GTT Val	TGT Cys	GAC Asp	TGC Cys	AAA Lys 830	Arg	AAC Asn	AGT Ser	GAC Asp	GTA Val 835	ATG Met	GAT Asp	TGT Cys	254	8
GTG Val	ATT Ile	CTG Leu	GAT Asp	GAT Asp	GGT Gly	GGG Gly	TTT Phe	CTT Leu	CTG Leu	ATG Met	GCA Ala	AAT Asn	CAT His	GAT Asp	GAT Asp	259	6

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1	840					84	5				85	0				
TAT 1 Tyr 3 855	ACT Thr	AAT Asn	CAG Gln	AT:	r GG e Gl	7	A TT g Ph	T TT e Ph	T GG e Gl	A GA y Gl 86	n II	T GA' e Ası	r cc	C AG O Se	C TTG r Leu 870	2644
ATG A Met A	AGA Arg	CAC His	CTG Leu	GT1 Val 875		r AT	A TC. e Se:	A GT r Va	T TA 1 Ty 88	r Al	T TT	T AA(e Asr	C AAI	A TC: S Se: 88!	Tyr	2692
GAT 1 Asp 1	•		890		. Cya	, GI	1 PI	89	2 A VI	a Ala	a Pro) Lys	900 900	ı Gly	/ Ala	2740
GGA C Gly H	AT (CGC Arg 905	TCA Ser	GCA Ala	TAT Tyr	GTC Val	910) Sei	A GTZ r Val	A GCA L Ala	A GAC	ATA Ile 915	Leu	CAA Gln	ATT	2788
GGC T Gly T 9	20	. •			712 U	925	MIG	ıırţ	ser	. TIE	930	Gln	Gln	Phe	Leu	2836
TTG A Leu S 935					940	a. g	neu	Leu	GIU	945	Val	Glu	Met	Glu	Asp 950	2884
GAT G Asp A:	AC I sp P	TC :	ACG Thr	GCC Ala 955	TCC Ser	CTG Leu	TCC Ser	AAG Lys	CAG Gln 960	ser	TGC Cys	ATT Ile	ACT Thr	GAA Glu 965	CAA Gln	2932
ACC CA	AG T ln T	2	TTC Phe 970	TTC Phe	GAT Asp	AAC Asn	GAC Asp	AGT Ser 975	AAA Lys	TCA Ser	TTC Phe	AGT Ser	GGT Gly 980	GTA Val	TTA Leu	2980
GAC TO Asp Cy	ST G /s G 9	GA 1 ly 1 85	AAC Asn	TGT Cys	TCC Ser	AGA Arg	ATC Ile 990	TTT Phe	CAT His	GGA Gly	GAA Glu	AAG Lys 995	CTT Leu	ATG Met	AAC Asn	3028
ACC AA Thr As	C T	TA A	ATA :	TTC Phe	ATA Ile	ATG Met 1005	vai	GAG Glu	AGC Ser	AAA Lys	GGG Gly 1010	Thr	TGT Cys	CCA Pro	TGT Cys	3076
GAC AC Asp Th 1015	A CO	GA C rg L	TG (eu]	ueu	ATA Ile 1020	GTII	GCG Ala	GAG Glu	CAG Gln	ACT Thr 1025	Ser	GAC Asp	GGT Gly	Pro	AAT Asn 1030	3124
CCT TG Pro Cy	T GA S As	AC A sp M		TT : /al :	AAG Lys	CAA Gln	CCT Pro	AGA Arg	TAC Tyr 1040	Arg	AAA Lys	GGG (Pro .	GAT Asp 1045	GTC Val	3172
TGC TT	T GA e As	·	AC A sn A 050	AT (GTC ' Val 1	TTG Leu	GIU	GAT Asp 1055	Tyr	ACT Thr	GAC Asp	Cys (GGT (Gly (GGT (GTT Val	3220
TCT GGA	A TT y Le	'A Ai	AT C	ro s	CC (Ser 1	CTG ' Leu '	TGG Trp	TAT . Tyr	ATC . Ile	ATT (GGA :	ATC (CAG :	rrr (CTA Leu	3268

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1065

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CTA CTT TGG CTG GTA TCT GGC AGC ACA CAC CGG CTG TTA TGACCTTCTA Leu Leu Trp Leu Val Ser Gly Ser Thr His Arg Leu Leu 1080 1085 1090	3317
AAAACCAAAT CTGCATAGTT AAACTCCAGA CCCTGCCAAA ACATGAGCCC TGCCCTCAAT	3377
TACAGTAACG TAGGGTCAGC TATAAAATCA GACAAACATT AGCTGGGCCT GTTCCATGGC	3437
ATAACACTAA GGCGCAGACT CCTAAGGCAC CCACTGGCTG CATGTCAGGG TGTCAGATCC	3497
TTAAACGTGT GTGAATGCTG CATCATCTAT GTGTAACATC AAAGCAAAAT CCTATACGTG	3557
TCCTCTATTG GAAAATTTGG GCGTTTGTTG TTGCATTGTT GGT	3600
(2) INFORMATION FOR SEQ ID NO:12: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 323 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:	
CCCCCTGCCA GTGGCCAAAC AGAAGCAGAA GTCGGGTAAT GAAATGACTA ACTTAGCCTT	60
TGAACTAGAC CCCCTAGAGT TAGAGGAGGA AGAGGCTGAG CTTGGTGAGC AGAGTGGCTC	120
TGCCAAGACT AGTGTTAGCA GTGTCACCAC CCCGCCACCC CATGGCAAAC GCATCCCCTT	180
CTTTAAGAAG ACAGAGCATG TGCCCCCCTA TGACGTGGTG CCTTCCATGA GGCCCATCAT	240
CCTGGTGGGA CCGTCGCTCA AGGGCTACGA GGTTACAGAC ATGATGCAGA AAGCTTTATT	300
TGACTTCTTG AAGCATCGGT TTG	323
(2) INFORMATION FOR SEQ ID NO:13:	
 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 57 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear 	
(ii) MOLECULE TYPE: DNA (genomic)	
(xi) SEQUENĆE DESCRIPTION: SEQ ID NO:13:	
CCTATTGGTG TAGGTATACC AACAATTAAT TTAAGAAAAA GGAGACCCAA TATCCAG	57

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(2) INFORMATION FOR SEQ ID NO:14:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 180 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic)	
(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1132	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:	
TGG TCC TTT GCC TGC GCC TGT GCC GCC TTC ATC CTC CTC TTT CTC GGC Trp Ser Phe Ala Cys Ala Cys Ala Phe Ile Leu Leu Phe Leu Gly 1 5 10 15	48
GGT CTC GCC CTG CTG TTC TCC CTG CCT CGA ATG CCC CGG AAC CCA Gly Leu Ala Leu Leu Phe Ser Leu Pro Arg Met Pro Arg Asn Pro 25 30	96
TGG GAG TCC TGC ATG GAT GCT GAG CCC GAG CAC TAACCCTCCT GCGGCCCTAG Trp Glu ser Cys Met Asp Ala Glu Pro Glu His 35 40	149
CGACCCTCAG GCTTCTTCCC AGGAAGCGGG G	7.00
(2) INFORMATION FOR SEQ ID NO:15:	180
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 22 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: Other nucleic acid; (A) DESCRIPTION: Oligonucleotide	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:	
AATTCGGTAC GTACACTCGA GC	22
(2) INFORMATION FOR SEQ ID NO:16:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 18 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: Other nucleic acid;(A) DESCRIPTION: Oligonucleotide	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:	

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GCTCGAGTGT ACGTACCG

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(2) INFORMATION FOR SEQ ID NO:17:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 20 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: Other nucleic acid;(A) DESCRIPTION: Oligonucleotide	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:17:	
CCATGGTACC TTCGTTGACG	20
(2) INFORMATION FOR SEQ ID NO:18:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 24 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: Other nucleic acid; (A) DESCRIPTION: Oligonucleotide	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:18:	
AATTCGTCAA CGAAGGTACC ATGG	24
(2) INFORMATION FOR SEQ ID NO:19:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 2153 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
<pre>(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 531504 (D) OTHER INFORMATION: /standard_name= "Beta-3-1"</pre>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:19:	
CCGCCTCGGA CCCCCTGTCC CGGGGGAGGG GGAGAGCCCG CTACCCTGGT CT ATG Met 1	55
TOT TIT TOT GAO TOO AGT GOA ACO TTO CTG CTG AAC GAG GGT TOA GOO	103

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Se	r Ph	e Se	r As	p Se 5	r Se	r Ala	a Th	r Ph	e Le O	u Le	u As	n Gl		y Se	er Ala	L
GA As	C TC	:	C AC r Th 0	C AG	C CG	C CC	A TC Se:	г це	G GA u As	C TC. p Se:	A GA r As	p Va	C TC 1 Se	C CI	G GAG	151
GA:	G GA u As _l 3!	C CG P Ar	G GA g Gl	G AG' u Sei	r GCC	C CGC A Arg	AI	T GAI g Glu	A GT	A GAG	G AG u Se 4	r Gl	G GC n Al	T CA a Gl	.G CAG n Gln	199
CAC Gl: 50	G CT(Lei)	GA Gl	A AG	G GCO	AAC Lys	urs.	AAI Lys	A CCT	GT(G GCA l Ala	a Pho	T GC e Al	G GT a Va	G AG l Ar	G ACC g Thr 65	247
AA Asi	GTO Val	AG0	TAC	TGT Cys	, Gra	GTA Val	CTC Lev	GAT Asp	GAC Glu	1 GIU	TG(C CC	A GTO	C CA l Gl: 8	G GGC n Gly	295
TC1 Ser	GGA Gly	GT(AAC Asr 88		GAG Glu	GCC Ala	AAA Lys	GAT Asp	Pne	CTG Leu	CAC His	C ATT	AAI Lys	Gl ₁	G AAG 1 Lys	343
TAC	AGC Ser	AAT Asr 100		TGG Trp	TGG Trp	ATC Ile	GGG Gly 105	Arg	CTA Leu	GTG Val	AAA Lys	GAC Glu	ı Gly	GGG Gl	GAC Asp	391
ATC Ile	GCC Ala 115	TTC	ATC : Ile	CCC Pro	AGC Ser	CCC Pro 120	CAG Gln	CGC Arg	CTG Leu	GAG Glu	AGC Ser 125	Ile	CGG Arg	CTC Lev	AAA Lys	439
CAG Gln 130	GAG Glu	CAG Gln	AAG Lys	GCC Ala	AGG Arg 135	AGA Arg	TCT Ser	GGG Gly	AAC Asn	CCT Pro 140	TCC Ser	AGC Ser	CTG Leu	AGT Ser	GAC Asp 145	487
ATT Ile	GGC Gly	AAC Asn	CGA Arg	CGC Arg 150	TCC Ser	CCT Pro	CCG Pro	CCA Pro	TCT Ser 155	CTA Leu	GCC Ala	AAG Lys	CAG Gln	AAG Lys		535
AAG Lys	CAG Gln	GCG Ala	GAA Glu 165	CAT His	GTT Val	CCC Pro	CCG Pro	TAT Tyr 170	GAC Asp	GTG Val	GTG Val	CCC Pro	TCC Ser 175	ATG Met	CGG Arg	583
CCT Pro	GTG Val	GTG Val 180	CTG Leu	GTG Val	GGA Gly	Pro	TCT Ser 185	CTG Leu	AAA Lys	GGT Gly	TAT Tyr	GAG Glu 190	GTC Val	ACA Thr	GAC Asp	631
ATG Met	ATG Met 195	CAG Gln	AAG Lys	GCT Ala	CTC Leu	TTC Phe 200	GAC Asp	TTC Phe	CTC Leu	AAA Lys	CAC His 205	AGA Arg	TTT Phe	GAT Asp	GGC Gly	679
AGG Arg 210	ATC Ile	TCC Ser	ATC Ile	ACC Thr	CGA Arg 215	GTC . Val	ACA Thr	GCC Ala	Asp	CTC Leu 220	TCC Ser	CTG Leu	GCA Ala	AAG Lys	CGA Arg 225	727
TCT	GTG	CTC	AAC	AAT	CCG	GGC 2	AAG .	AGG .	ACC	ATC .	ATT	GAG	CGC	TCC		775

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Ser	Val	Leu	Asn	Asn 230	Pro	Gly	Lys	Arg	Thr 235	Ile	Ile	Glu	Arg	Ser 240	Ser	
	CGC Arg															823
	CTG Leu															871
	CAC His 275															919
TTT Phe 290	GTC Val	AAA Lys	GTG Val	TCC Ser	TCA Ser 295	CCA Pro	AAG Lys	GTA Val	CTC Leu	CAG Gln 300	CGT Arg	CTC Leu	ATT Ile	CGC Arg	TCC Ser 305	967
CGG Arg	GGG Gly	AAG Lys	TCA Ser	CAG Gln 310	ATG Met	AAG Lys	CAC His	CTG Leu	ACC Thr 315	GTA Val	CAG Gln	ATG Met	ATG Met	GCA Ala 320	TAT Tyr	1015
GAT Asp	AAG Lys	CTG Leu	GTT Val 325	CAG Gln	TGC Cys	CCA Pro	CCG Pro	GAG Glu 330	TCA Ser	TTT Phe	GAT Asp	GTG Val	ATT Ile 335	CTG Leu	GAT Asp	1063
	AAC Asn															1111
GTT Val	TAC Tyr 355	TGG Trp	CGG Arg	GCC Ala	ACG Thr	CAC His 360	CAC His	CCA Pro	GCC Ala	CCT Pro	GGC Gly 365	CCC Pro	GGA Gly	CTT Leu	CTG Leu	1159
GGT Gly 370	CCT Pro	CCC Pro	AGT Ser	GCC Ala	ATC Ile 375	CCC Pro	GGA Gly	CTT Leu	CAG Gln	AAC Asn 380	CAG Gln	CAG Gln	CTG Leu	CTG Leu	GGG Gly 385	1207
GAG Glu	CGT Arg	GGC Gly	GAG Glu	GAG Glu 390	CAC His	TCC Ser	CCC Pro	CTT Leu	GAG Glu 395	CGG Arg	GAC Asp	AGC Ser	TTG Leu	ATG Met 400	CCC Pro	1255
TCT Ser	GAT Asp	GAG Glu	GCC Ala 405	AGC Ser	GAG Glu	AGC Ser	TCC Ser	CGC Arg 410	CAA Gln	GCC Ala	TGG Trp	ACA Thr	GGA Gly 415	TCT Ser	TCA Ser	1303
CAG Gln	CGT Arg	AGC Ser 420	TCC Ser	CGC Arg	CAC His	CTG Leu	GAG Glu 425	GAG Glu	GAC Asp	TAT Tyr	GCA Ala	GAT Asp 430	GCC Ala	TAC Tyr	CAG Gln	1351
GAC Asp	CTG Leu 435	TAC Tyr	CAG Gln	CCT Pro	CAC His	CGC Arg 440	CAA Gln	CAC His	ACC Thr	TCG Ser	GGG Gly 445	CTG Leu	CCT Pro	AGT Ser	GCT Ala	1399
AAC	GGG	CAT	GAC	CCC	CAA	GAC	CGG	CTT	CTA	GCC	CAG	GAC	TCA	GAA	CAC	1447

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Asn Gly His Asp Pro Gln Asp Arg Leu Leu Ala Gln Asp Ser Glu His 450 460	
407	
AAC CAC AGT GAC CGG AAC TGG CAG CGC AAC CGG CCT TGG CCC AAG GAT Asn His Ser Asp Arg Asn Trp Gln Arg Asn Arg Pro Trp Pro Lys Asp 470 475 480	1495
AGC TAC TGA CAG C CTCCTGCTGC CCTACCCTGG CAGGCACAGG Ser Tyr *	1543
CGCAGCTGGC TGGGGGGCCC ACTCCAGGCA GGGTGGCGTT AGACTGGCAT	1593
CAGGCTGGCA CTAGGCTCAG CCCCCAAAAC CCCCTGCCCA GCCCCAGCTT CAGGGCTGCC	1648
TGTGGTCCCA AGGTTCTGGG AGAAACAGGG GACCCCTCA CCTCCTGGGC AGTGACCCCT	1708
ACTAGGCTCC CATTCCAGGT ACTAGCTGTG TGTTCTGCAC CCCTGGCACC TTCCTCCT	1768
CCCACACAGG AAGCTGCCCC ACTGGGCAGT GCCCTCAGGC CAGGATCCCC TTAGCAGGGT	1828
CCTTCCCACC AGACTCAGGG AAGGGATGCC CCATTAAAGT GACAAAAGGG TGGGTGTGGG	
CACCATGGCA TGAGGAAGAA ACAAGGTCCC TGAGCAGGCA CAAGTCCTGA CAGTCAAGGG	1888
	1948
ACTGCTTTGG CATCCAGGGC CTCCAGTCAC CTCACTGCCA TACATTAGAA ATGAGACAAT	2008
TCAAAGCCCC CCCAGGGTGG CACACCCATC TGTTGCTGGG GTGTGGCAGC CACATCCAAG	2068
ACTGGAGCAG CAGGCTGGCC ACGCTTGGGC CAGAGAGAGC TCACAGCTGA AGCTCTTGGA	2128
GGGAAGGGCT CTCCTCACCC AATCG	2153
(2) INFORMATION FOR SEQ ID NO:20:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 2144 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: unknown	
(ii) MOLECULE TYPE: DNA (genomic)	
<pre>(ix) FEATURE:</pre>	
(ii) MOLECULE TYPE: cDNA	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:20:	
CGCCCCGGC GCCGCTCGTT CCCCCGACCC GGACTCCCCC ATGTATGACG ACTCCTACGT	60
GCCCGGGTTT GAGGACTCGG AGGCGGTTTC AGCCGACTCC TACACCAGCC GCCCATCTCT	120

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GGACTCAGAC	GTCTCCCTGG	AGGAGGACCG	GGAGAGTGCC	CGGCGTGAAG	TAGAGAGCCA	180
GGCTCAGCAG	CAGCTCGAAA	GGGCCAAGCA	CAAACCTGTG	GCATTTGCGG	TGAGGACCAA	240
TGTCAGCTAC	TGTGGCGTAC	TGGATGAGGA	GTGCCCAGTC	CAGGGCTCTG	GAGTCAACTT	300
TGAGGCCAAA	GATTTTCTGC	ACATTAAAGA	GAAGTACAGC	AATGACTGGT	GGATCGGGCG	360
GCTAGTGAAA	GAGGGCGGG	ACATCGCCTT	CATCCCCAGC	CCCCAGCGCC	TGGAGAGCAT	420
CCGGCTCAAA	CAGGAGCAGA	AGGCCAGGAG	ATCTGGGAAC	CCTTCCAGCC	TGAGTGACAT	480
TGGCAACCGA	CGCTCCCCTC	CGCCATCTCT	AGCCAAGCAG	AAGCAAAAGC	AGGCGGAACA	540
TGTTCCCCCG	TATGACGTGG	TGCCCTCCAT	GCGGCCTGTG	GTGCTGGTGG	GACCCTCTCT	600
GAAAGGTTAT	GAGGTCACAG	ACATGATGCA	GAAGGCTCTC	TTCGACTTCC	TCAAACACAG	660
ATTTGATGGC	AGGATCTCCA	TCACCCGAGT	CACAGCCGAC	CTCTCCCTGG	CAAAGCGATC	720
TGTGCTCAAC	AATCCGGGCA	AGAGGACCAT	CATTGAGCGC	TCCTCTGCCC	GCTCCAGCAT	780
TGCGGAAGTG	CAGAGTGAGA	TCGAGCGCAT	ATTTGAGCTG	GCCAAATCCC	TGCAGCTAGT	840
AGTGTTGGAC	GCTGACACCA	TCAACCACCC	AGCACAGCTG	GCCAAGACCT	CGCTGGCCCC	900
CATCATCGTC	TTTGTCAAAG	TGTCCTCACC	AAAGGTACTC	CAGCGTCTCA	TTCGCTCCCG	960
GGGGAAGTCA	CAGATGAAGC	ACCTGACCGT	ACAGATGATG	GCATATGATA	AGCTGGTTCA	1020
GTGCCCACCG	GAGTCATTTG	ATGTGATTCT	GGATGAGAAC	CAGCTGGAGG	ATGCCTGTGA	1080
GCACCTGGCT	GAGTACCTGG	AGGTTTACTG	GCGGGCCACG	CACCACCCAG	CCCCTGGCCC	1140
CGGACTTCTG	GGTCCTCCCA	GTGCCATCCC	CGGACTTCAG	AACCAGCAGC	TGCTGGGGGA	1200
GCGTGGCGAG	GAGCACTCCC	CCCTTGAGCG	GGACAGCTTG	ATGCCCTCTG	ATGAGGCCAG	1260
CGAGAGCTCC	CGCCAAGCCT	GGACAGGATC	TTCACAGCGT	AGCTCCCGCC	ACCTGGAGGA	1320
GGACTATGCA	GATGCCTACC	AGGACCTGTA	CCAGCCTCAC	CGCCAACACA	CCTCGGGGCT	1380
GCCTAGTGCT	AACGGGCATG	ACCCCCAAGA	CCGGCTTCTA	GCCCAGGACT	CAGAACACAA	1440
CCACAGTGAC	CGGAACTGGC	AGCGCAACCG	GCCTTGGCCC	AAGGATAGCT	ACTGACAGCC	1500
TCCTGCTGCC	CTACCCTGGC	AGGCACAGGC	GCAGCTGGCT	GGGGGCCCA	CTCCAGGCAG	1560
GGTGGCGTTA	GACTGGCATC	AGGCTGGCAC	TAGGCTCAGC	CCCCAAAACC	CCCTGCCCAG	1620
CCCCAGCTTC	AGGGCTGCCT	GTGGTCCCAA	GGTTCTGGGA	GAAACAGGGG	ACCCCCTCAC	1680
CTCCTGGGCA	GTGACCCCTA	CTAGGCTCCC	ATTCCAGGTA	CTAGCTGTGT	GTTCTGCACC	1740
CCTGGCACCT	TCCTCTCCTC	CCACACAGGA	AGCTGCCCCA	CTGGGCAGTG	CCCTCAGGCC	1800

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AGGATCCCCT TAGCAGGGTC CTTCCCACCA GACTCAGGGA AGGGATGCCC CATTAAAGTG	1860
ACAAAAGGGT GGGTGTGGGC ACCATGGCAT GAGGAAGAAA CAAGGTCCCT GAGCAGGCAC	1920
AAGTCCTGAC AGTCAAGGGA CTGCTTTGGC ATCCAGGGCC TCCAGTCACC TCACTGCCAT	1980
ACATTAGAAA TGAGACAATT CAAAGCCCCC CCAGGGTGGC ACACCCATCT GTTGCTGGGG	2040
TGTGGCAGCC ACATCCAAGA CTGGAGCAGC AGGCTGGCCA CGCTTGGGCC AGAGAGAGCT	2100
CACAGCTGAA GCTCTTGGAG GGAAGGGCTC TCCTCACCCA ATCG	2144
(2) INFORMATION FOR SEQ ID NO:21:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 28 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: single (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: Other nucleic acid;(A) DESCRIPTION: Oligonucleotide	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:21:	
CTCAGTACCA TCTCTGATAC CAGCCCCA	28
(2) INFORMATION FOR SEQ ID NO:22:	
(i) SEQUENCE CHARACTERISTICS:	
(A) LENGTH: 7808 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
<pre>(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 2377769 (D) OTHER INFORMATION: /standard_name= "Alpha-1A-1"</pre>	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:22:	
GATGTCCCGA GCTGCTATCC CCGGCTCGGC CCGGGCAGCC GCCTTCTGAG CCCCCGACCC	60
GAGGCGCCGA GCCGCCGCCG CCCGATGGGC TGGGCCGTGG AGCGTCTCCG CAGTCGTAGC	120
TCCAGCCGCC GCGCTCCCAG CCCCGGCAGC CTCAGCATCA GCGGCGGCGG CGGCGGCGGC	180
GGCGTCTTCC GCATCGTTCG CCGCAGCGTA ACCCGGAGCC CTTTGCTCTT TGCAGA	236
ATG GCC CGC TTC GGA GAC GAG ATG CCG GCC CGC TAC GGG GGA GGA GGC	284

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Met 1	Ala	Arg	Phe	Gly 5	Asp	Glu	Met	Pro	Ala 10	Arg	Tyr	Gly	Gly	Gly 15	Gly		
TCC Ser	GGG Gly	GCA Ala	GCC Ala 20	GCC Ala	GGG Gly	GTG Val	GTC Val	GTG Val 25	GGC Gly	AGC Ser	GGA Gly	GGC Gly	GGG Gly 30	CGA Arg	GGA Gly	-	332
GCC Ala	GGG Gly	GGC Gly 35	AGC Ser	CGG Arg	CAG Gln	GGC Gly	GGG Gly 40	CAG Gln	CCC Pro	GGG Gly	GCG Ala	CAA Gln 45	AGG Arg	ATG Met	TAC Tyr		380
AAG Lys	CAG Gln 50	TCA Ser	ATG Met	GCG Ala	CAG Gln	AGA Arg 55	GCG Ala	CGG Arg	ACC Thr	ATG Met	GCA Ala 60	CTC Leu	TAC Tyr	AAC Asn	CCC Pro		428
ATC Ile 65	CCC Pro	GTC Val	CGA Arg	CAG Gln	AAC Asn 70	TGC Cys	CTC Leu	ACG Thr	GTT Val	AAC Asn 75	CGG Arg	TCT Ser	CTC Leu	TTC Phe	CTC Leu 80		476
TTC Phe	AGC Ser	GAA Glu	GAC Asp	AAC Asn 85	GTG Val	GTG Val	AGA Arg	AAA Lys	TAC Tyr 90	GCC Ala	AAA Lys	AAG Lys	ATC Ile	ACC Thr 95	GAA Glu		524
TGG Trp	CCT Pro	CCC Pro	TTT Phe 100	GAA Glu	TAT Tyr	ATG Met	ATT Ile	TTA Leu 105	GCC Ala	ACC Thr	ATC Ile	ATA Ile	GCG Ala 110	AAT Asn	TGC Cys		572
ATC Ile	GTC Val	CTC Leu 115	GCA Ala	CTG Leu	GAG Glu	CAG Gln	CAT His 120	CTG Leu	CCT Pro	GAT Asp	GAT Asp	GAC Asp 125	AAG Lys	ACC Thr	CCG Pro		620
ATG Met	TCT Ser 130	GAA Glu	CGG Arg	CTG Leu	GAT Asp	GAC Asp 135	ACA Thr	GAA Glu	CCA Pro	TAC Tyr	TTC Phe 140	ATT Ile	GGA Gly	ATT Ile	TTT Phe		668
TGT Cys 145	TTC Phe	GAG Glu	GCT Ala	GGA Gly	ATT Ile 150	AAA Lys	ATC Ile	ATT Ile	GCC Ala	CTT Leu 155	GGG Gly	TTT Phe	GCC Ala	TTC Phe	CAC His 160		716
						AAT Asn											764
						GCG Ala											812
ACG Thr	CTG Leu	AGG Arg 195	GCA Ala	GTT Val	CGA Arg	GTG Val	CTG Leu 200	CGG Arg	CCG Pro	CTC Leu	AAG Lys	CTG Leu 205	GTG Val	TCT Ser	GGA Gly		860
						GTC Val 215											908
CCT	TTG	CTG	CAG	ATC	GGC	CTC	CTC	CTA	TTT	TTT	GCA	ATC	CTT	ATT	TTT		956

Pro 225	Leu	Leu	Gln	Ile	Gly 230	Leu	Leu	Leu	Phe	Phe 235	Ala	Ile	Leu	Ile	Phe 240		
GCA Ala	ATC	ATA Ile	GGG Gly	TTA Leu 245	Glu	TTT Phe	TAT Tyr	ATG Met	GGA Gly 250	AAA Lys	TTT Phe	CAT His	ACC Thr	ACC Thr 255	TGC Cys	:	1004
TTT	GAA Glu	GAG Glu	GGG Gly 260	Thr	GAT Asp	GAC Asp	ATT Ile	CAG Gln 265	GGT Gly	GAG Glu	TCT Ser	CCG Pro	GCT Ala 270	CCA Pro	TGT Cys	:	1052
GGG	ACA Thr	GAA Glu 275	GAG Glu	CCC Pro	GCC Ala	CGC Arg	ACC Thr 280	TGC Cys	CCC Pro	AAT Asn	GGG Gly	ACC Thr 285	AAA Lys	TGT Cys	CAG Gln	3	1100
CCC Pro	TAC Tyr 290	TGG Trp	GAA Glu	GGG Gly	CCC Pro	AAC Asn 295	AAC Asn	GGG Gly	ATC Ile	ACT Thr	CAG Gln 300	TTC Phe	GAC Asp	AAC Asn	ATC Ile	3	1148
CTG Leu 305	TTT Phe	GCA Ala	GTG Val	CTG Leu	ACT Thr 310	GTT Val	TTC Phe	CAG Gln	TGC Cys	ATA Ile 315	ACC Thr	ATG Met	GAA Glu	GGG Gly	TGG Trp 320	1	1196
ACT Thr	GAT Asp	CTC Leu	CTC Leu	TAC Tyr 325	AAT Asn	AGC Ser	AAC Asn	GAT Asp	GCC Ala 330	TCA Ser	GGG Gly	AAC Asn	ACT Thr	TGG Trp 335	AAC Asn	1	L244
TGG Trp	TTG Leu	TAC Tyr	TTC Phe 340	ATC Ile	CCC Pro	CTC Leu	ATC Ile	ATC Ile 345	ATC Ile	GGC Gly	TCC Ser	TTT Phe	TTT Phe 350	ATG Met	CTG Leu	3	1292
AAC Asn	CTT Leu	GTG Val 355	CTG Leu	GGT Gly	GTG Val	CTG Leu	TCA Ser 360	GGG Gly	GAG Glu	TTT Phe	GCC Ala	AAA Lys 365	GAA Glu	AGG Arg	GAA Glu	1	1340
CGG Arg	GTG Val 370	GAG Glu	AAC Asn	CGG Arg	CGG Arg	GCT Ala 375	TTT Phe	CTG Leu	AAG Lys	CTG Leu	AGG Arg 380	CGG Arg	CAA Gln	CAA Gln	CAG Gln	1	.388
ATT Ile 385	GAA Glu	CGT Arg	GAG Glu	CTC Leu	AAT Asn 390	GGG Gly	TAC Tyr	ATG Met	GAA Glu	TGG Trp 395	ATC Ile	TCA Ser	AAA Lys	GCA Ala	GAA Glu 400	1	.436
GAG Glu	GTG Val	ATC Ile	CTC Leu	GCC Ala 405	GAG Glu	GAT Asp	GAA Glu	ACT Thr	GAC Asp 410	GGG Gly	GAG Glu	CAG Gln	AGG Arg	CAT His 415	CCC Pro	1	484
TTT Phe	GAT Asp	GGA Gly	GCT Ala 420	CTG Leu	CGG Arg	AGA Arg	ACC Thr	ACC Thr 425	ATA Ile	AAG Lys	AAA Lys	AGC Ser	AAG Lys 430	ACA Thr	GAT Asp	1	.532
TTG Leu	CTC Leu	AAC Asn 435	CCC Pro	GAA Glu	GAG Glu	Ala	GAG Glu 440	GAT Asp	CAG Gln	CTG Leu	GCT Ala	GAT Asp 445	ATA Ile	GCC Ala	TCT Ser	1	.580
GTG	GGT	TCT	CCC	TTC	GCC	CGA	GCC	AGC	TTA	AAA	AGT	GCC	AAG	CTG	GAG	1	628

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Val	Gly 450	Ser	Pro	Phe	Ala	Arg 455	Ala	Ser	Ile	Lys	Ser 460	Ala	Lys	Leu	Glu	
AAC Asn 465	TCG Ser	ACC Thr	TTT Phe	TTT Phe	CAC His 470	AAA Lys	AAG Lys	GAG Glu	AGG Arg	AGG Arg 475	ATG Met	CGT Arg	TTC Phe	TAC Tyr	ATC Ile 480	1676
CGC Arg	CGC Arg	ATG Met	GTC Val	AAA Lys 485	ACT Thr	CAG Gln	GCC Ala	TTC Phe	TAC Tyr 490	TGG Trp	ACT Thr	GTA Val	CTC Leu	AGT Ser 495	TTG Leu	1724
GTA Val	GCT Ala	CTC Leu	AAC Asn 500	ACG Thr	CTG Leu	TGT Cys	GTT Val	GCT Ala 505	ATT Ile	GTT Val	CAC His	TAC Tyr	AAC Asn 510	CAG Gln	CCC Pro	1772
GAG Glu	TGG Trp	CTC Leu 515	TCC Ser	GAC Asp	TTC Phe	CTT	TAC Tyr 520	TAT Tyr	GCA Ala	GAA Glu	TTC Phe	ATT Ile 525	TTC Phe	TTA Leu	GGA Gly	1820
CTC Leu	TTT Phe 530	ATG Met	TCC Ser	GAA Glu	ATG Met	TTT Phe 535	ATA Ile	AAA Lys	ATG Met	TAC Tyr	GGG Gly 540	CTT Leu	GGG Gly	ACG Thr	CGG Arg	1868
CCT Pro 545	TAC Tyr	TTC Phe	CAC His	TCT Ser	TCC Ser 550	TTC Phe	AAC Asn	TGC Cys	TTT Phe	GAC Asp 555	TGT Cys	GGG Gly	GTT Val	ATC Ile	ATT Ile 560	1916
					GTC Val											1964
					TTA Leu											2012
					GCA Ala											2060
					ATC Ile											2108
_					CTT Leu 630											2156
					ACT Thr											2204
					TTT Phe											2252
GTC	ATG	TAC	GAC	GGG	ATC	AAG	TCT	CAG	GGG	GGC	GTG	CAG	GGC	GGC	ATG	2300

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Va]	. Met	Ty:	As ₁	o Gly	/ Ile	E Lys	Ser 680	Glr	ı Gly	/ Gly	/ Val	Gl: 685		y Gl	y Met		
GT0 Val	Phe 690	ser	C ATO	TAT	TTC Phe	ATT E Ile 695	. Val	CTC Leu	ACG Thr	CTC Lev	TT1 Phe 700	e Gly	AA E raa v	TAC Ty	C ACC Thr	23	348
CTC Leu 705	neu	AAT Asn	GT(TTC Phe	TTG Leu 710	ALA	ATC Ile	GCT Ala	GTG Val	GAC Asp 715	Asn	CTO Lev	GCC Ala	AA(Asr	GCC Ala 720	23	96
CAG Gln	GAG Glu	CTC Leu	ACC Thr	AAG Lys 725	val	GAG Glu	GCG Ala	GAC Asp	GAG Glu 730	Gln	GAG Glu	GAA Glu	GAA Glu	GAA Glu 735	GCA Ala	24	44
GCG Ala	AAC Asn	CAG Gln	AAA Lys 740	Leu	GCC Ala	CTA Leu	CAG Gln	AAA Lys 745	Ala	AAG Lys	GAG Glu	GTG Val	GCA Ala 750	Glu	GTG Val	24	92
AGT Ser	CCT Pro	CTG Leu 755	Sei	GCG Ala	GCC Ala	AAC Asn	ATG Met 760	TCT Ser	ATA Ile	GCT Ala	GTG Val	AAA Lys 765	GAG Glu	CAA Gln	CAG Gln	25	40
AAG Lys	AAT Asn 770	CAA Gln	AAG Lys	CCA Pro	GCC Ala	AAG Lys 775	TCC Ser	GTG Val	TGG Trp	GAG Glu	CAG Gln 780	CGG Arg	ACC Thr	AGT Ser	GAG Glu	25	88
ATG Met 785	CGA Arg	AAG Lys	CAG Gln	AAC Asn	TTG Leu 790	CTG Leu	GCC Ala	AGC Ser	CGG Arg	GAG Glu 795	GCC Ala	CTG Leu	TAT Tyr	AAC Asn	GAA Glu 800	26:	36
ATG Met	GAC Asp	CCG Pro	GAC Asp	GAG Glu 805	CGC Arg	TGG Trp	AAG Lys	GCT Ala	GCC Ala 810	TAC Tyr	ACG Thr	CGG Arg	CAC His	CTG Leu 815	CGG Arg	268	34
CCA Pro	GAC Asp	ATG Met	AAG Lys 820	ACG Thr	CAC His	TTG Leu	GAC Asp	CGG Arg 825	CCG Pro	CTG Leu	GTG Val	GTG Val	GAC Asp 830	CCG Pro	CAG Gln	273	32
GAG Glu	AAC Asn	CGC Arg 835	AAC Asn	AAC Asn	AAC Asn	ACC Thr	AAC Asn 840	AAG Lys	AGC Ser	CGG Arg	GCG Ala	GCC Ala 845	GAG Glu	CCC Pro	ACC Thr	278	80
GTG Val	GAC Asp 850	CAG Gln	CGC Arg	CTC Leu	GGC Gly	CAG Gln 855	CAG Gln	CGC Arg	GCC Ala	GAG Glu	GAC Asp 860	TTC Phe	CTC Leu	AGG Arg	AAA Lys	282	8
CAG Gln 865	GCC Ala	CGC Arg	TAC Tyr	CAC His	GAT Asp 870	CGG Arg	GCC Ala	CGG Arg	Asp	CCC Pro 875	AGC Ser	GGC Gly	TCG Ser	GCG Ala	GGC Gly 880	287	6
CTG Leu	GAC Asp	GCA Ala	CGG Arg	AGG Arg 885	CCC Pro	TGG Trp	GCG Ala	Gly	AGC Ser 890	CAG Gln	GAG Glu	GCC Ala	GAG Glu	CTG Leu 895	AGC Ser	292	4
CGG	GAG	GGA	CCC	TAC	GGC	CGC	GAG	TCG	GAC	CAC	CAC	GCC	CGG	GAG	GGC	297	2

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Arg	Glu	Gly	Pro 900	Tyr	Gly	Arg	Glu	Ser 905	Asp	His	His	Ala	Arg 910	Glu	Gly	
AGC Ser	CTG Leu	GAG Glu 915	CAA Gln	CCC Pro	GGG Gly	TTC Phe	TGG Trp 920	GAG Glu	GGC Gly	GAG Glu	GCC Ala	GAG Glu 925	CGA Arg	GGC Gly	AAG Lys	302 0
GCC Ala	GGG Gly 930	Asp	CCC Pro	CAC His	CGG Arg	AGG Arg 935	CAC His	GTG Val	CAC His	CGG Arg	CAG Gln 940	GGG Gly	GGC Gly	AGC Ser	AGG Arg	3068
GAG Glu 945	AGC Ser	CGC Arg	AGC Ser	GGG Gly	TCC Ser 950	CCG Pro	CGC Arg	ACG Thr	GGC Gly	GCG Ala 955	GAC Asp	GGG Gly	GÁG Glu	CAT His	CGA Arg 960	3116
CGT Arg	CAT His	CGC Arg	GCG Ala	CAC His 965	CGC Arg	AGG Arg	CCC Pro	GGG Gly	GAG Glu 970	GAG Glu	GGT Gly	CCG Pro	GAG Glu	GAC Asp 975	AAG Lys	3164
GCG Ala	GAG Glu	CGG Arg	AGG Arg 980	GCG Ala	CGG Arg	CAC His	CGC Arg	GAG Glu 985	GGC Gly	AGC Ser	CGG Arg	CCG Pro	GCC Ala 990	CGG Arg	GGC Gly	3212
GGC Gly	GAG Glu	GGC Gly 995	GAG Glu	GGC Gly	GAG Glu	GGC Gly	CCC Pro 100	Asp	GGG Gly	GGC Gly	GAG Glu	CGC Arg 100	Arg	AGA Arg	AGG Arg	3260
CAC His	CGG Arg 1010	His	GGC Gly	GCT Ala	CCA Pro	GCC Ala 101	Thr	TAC Tyr	GAG Glu	GGG Gly	GAC Asp 102	Ala	CGG Arg	AGG Arg	GAG Glu	3308
GAC Asp 102	Lys	GAG Glu	CGG Arg	AGG Arg	CAT His 103	Arg	AGG Arg	AGG Arg	AAA Lys	GAG Glu 103	Asn	CAG Gln	GGC Gly	TCC	GGG Gly 1040	3356
GTC Val	CCT Pro	GTG Val	TCG Ser	GGC Gly 104	Pro	AAC Asn	CTG Leu	TCA Ser	ACC Thr 105	Thr	CGG Arg	CCA Pro	ATC Ile	CAG Gln 105	GIII	3404
GAC Asp	CTG Leu	GGC Gly	CGC Arg 106	Gln	GAC Asp	CCA Pro	CCC	CTG Leu 106	Ala	GAG Glu	GAT Asp	ATT	GAC Asp 107	ASII	ATG Met	3452
AAG Lys	AAC Asn	AAC Asn 107	Lys	CTG Leu	GCC Ala	ACC Thr	GCG Ala 108	Glu	TCG Ser	GCC Ala	GCT Ala	CCC Pro 108	HIP	GGC	AGC Ser	3500
CTT Leu	GGC Gly 109	His	GCC Ala	GGC Gly	CTG Leu	CCC Pro	Glr	AGC Ser	CCA Pro	GCC Ala	Lys 110	met	GGA Gly	AAC Asn	AGC Ser	3548
ACC Thr 110	Asp	CCC Pro	GGC Gly	CCC Pro	ATG Met	Lev	GCC Ala	TATO	C CCI Pro	GCC Ala 111	Met	GCC Ala	ACC Thr	AAC Asr	CCC Pro 1120	3596
CAG	AAC	GCC	GCC	AGC	CGC	CGG	ACC	3 CC	C AAC	. AAC	ccc	GGG	AA S	CCF	TCC	3644

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Gl	n As	n Al	a Al	.a Se 11	r Ar	g Ar	g Th	r Pr	O As	n As .30	n Pr	o Gl	y As	sn Pi	ro Se 135	r	
AA As:	T CC n Pr	C GG o Gl		C CC O Pr 40	C AA(o Ly:	G AC	C CCC	C GA	u As	T AG	C CT r Le	T AT u Il	e Va	C AC	CC AA	C n	3692
Pro	C AG	C GG r Gl 11	C AC y Th 55	C CA	G ACC	C AAT	1 261	A GC Ala	а шу	G AC's Th	T GC	C AG a Ar	д Гу	A CC	C GAC	C P	3740
CA(C ACC	C AC. r Th: 70	A GT	G GA(l As)	C ATO	C CCC Pro 117	PIC	GCC Ala	TGG Cy:	C CCI s Pro	A CCC Pro	o Pro	C CT	C AA u As	C CAC	3	3788
ACC Thr 118	GTO Val	GT Va	A CAI	A GTO	AAC Asn 119	· • s	AAC Asn	GCC Ala	C AAG R Asi	C CCA n Pro 119) Asp	C CCI	A CTO	G CC u Pr	A AAA o Lys 120	3	3836
AAA Lys	GAG Glu	GAZ Glu	A GAO	AAG Lys 120	AAG Lys	GAG Glu	GAG Glu	GAG Glu	GAZ Glu 121	ı Asp	GAC Asp	C CGT	GGG	G GAN	A GAC u Asp 15		3884
		•	122	0	-10	110	-yr	122	5	Met	Pne	: Ile	Let 123	ı Sei 80	C ACG		3932
		123	5	3	9	Lcu	124	D	Tyr	TIE	Leu	Asn 124	Leu 5	Arg	TAC Tyr		3980
TTT Phe	GAG Glu 125	ATG Met 0	TGC Cys	ATC Ile	CTC Leu	ATG Met 1255	Val	ATT Ile	GCC Ala	ATG Met	AGC Ser 126	Ser	ATC	GCC Ala	CTG Leu		4028
GCC Ala 1265	GCC Ala	GAG Glu	GAC Asp	CCT Pro	GTG Val 1270	GIII	CCC Pro	AAC Asn	GCA Ala	CCT Pro 1275	Arg	AAC Asn	AAC Asn	GTG Val	CTG Leu 1280)	4076
CGA Arg	TAC Tyr	TTT Phe	GAC Asp	TAC Tyr 1285	val	TTT Phe	ACA Thr	GGC Gly	GTC Val 129	Phe	ACC Thr	TTT Phe	GAG Glu	ATG Met 129	Val		4124
ATC Ile	AAG Lys	ATG Met	ATT Ile 1300	GAC Asp	CTG Leu	GGG Gly	ren	GTC Val 1305	Leu	CAT His	CAG Gln	GGT Gly	GCC Ala 131	Tyr	TTC Phe		4172
CGT Arg		CTC Leu 1315		AAT Asn	ATT Ile	neu.	GAC Asp 1320	TTC Phe	ATA Ile	GTG Val	GTC Val	AGT Ser 1325	Gly	GCC Ala	CTG Leu		4220
	GCC Ala 1330		GCC Ala	TTC Phe	T T T	GGC 1 Gly 1 1335	AAT :	AGC Ser	AAA Lys	GTA .	AAA Lys 1340	Asp	ATC Ile	AAC Asn	ACG Thr		4268
TT.	AAA	TCC	CTC	CGA	GTC (CTC (CGG (GTG (CTA	CGA	CCT (CTT	AAA	ACC	ATC		4316

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Ile Lys Ser 1345	Leu Arg Val		al Leu Arg 135		Thr Ile 1360	
AAG CGG CTG Lys Arg Leu						4364
CTT AAA AAC Leu Lys Asn		Ile Leu Il			Met Phe	4412
ATC TTC GCC Ile Phe Ala 1395	Val Val Ala					4460
TGC ACT GAC Cys Thr Asp 1410						4508
CTC CTC TAC Leu Leu Tyr 1425		Glu Val Ly		Asp Arg Glu		4556
AAG TAT GAA Lys Tyr Glu						4604
TTC ACC GTG Phe Thr Val		Glu Gly Tr			His Ser	4652
GTG GAC GCC Val Asp Ala 1475	Thr Phe Glu					4700
GAG ATG TCC Glu Met Ser 1490						4748
TTT GTC AAT Phe Val Asn 1505	ATC TTT GTG Ile Phe Val 151	Ala Leu Il	TC ATC ATC le Ile Ile 151	Thr Phe Gln	GAG CAA Glu Gln 1520	4796
GGG GAC AAG Gly Asp Lys						4844
TGC ATT GAT Cys Ile Asp	TTC GCC ATC Phe Ala Ile 1540	Ser Ala Ly	AG CCG CTG ys Pro Leu 545	ACC CGA CAC Thr Arg His 155	Met Pro	4892
CAG AAC AAG Gln Asn Lys 1555	Gln Ser Phe	CAG TAC CO Gln Tyr Ar 1560	GC ATG TGG rg Met Trp	CAG TTC GTG Gln Phe Val 1565	GTG TCT Val Ser	4940
CCG CCT TTC	GAG TAC ACG	ATC ATG GO	CC ATG ATC	GCC CTC AAC	ACC ATC	4988

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Pro	Pro 157	Phe	Glu	туг	Thr	Ile 157	Met	: Ala	a Met	: Ile	Ala 158	a Lei 30	ı Ası	ı Thi	r Ile	
GTG Val 158	neu	ATG Met	ATG Met	AAG Lys	TTC Phe 159	Tyr	GGG	GCT Ala	TCT Ser	GTT Val	. Ala	TAT Tyr	GAA Glu	A AAT	GCC Ala 1600	5036
CTG Leu	CGG Arg	GTG Val	TTC Phe	AAC Asn 160	тте	GTC Val	TTC Phe	ACC	TCC Ser 161	Leu	TTC Phe	TCI Ser	CTC Lev	GAZ Glu 161	TGT Cys	5084
GTG Val	CTG Leu	AAA Lys	GTC Val 162	Mec	GCT Ala	TTT Phe	GGG Gly	ATT Ile 162	Leu	AAT Asn	TAT	TTC Phe	CGC Arg	Asp	GCC Ala	5132
TGG Trp	AAC Asn	ATC Ile 163	PHE	GAC Asp	TTT Phe	GTG Val	ACT Thr 164	Val	CTG Leu	GGC Gly	AGC Ser	ATC Ile 164	Thr	GAT Asp	ATC	5180
CTC Leu	GTG Val 165	TILL	GAG Glu	TTT	GGG Gly	AAT Asn 165	Pro	AAT Asn	AAC Asn	TTC Phe	ATC Ile 166	Asn	CTG Leu	AGC Ser	TTT Phe	5228
CTC Leu 1665	ALG	CTC Leu	TTC Phe	CGA Arg	GCT Ala 1670	Ala	CGG Arg	CTC Leu	ATC Ile	AAA Lys 167	Leu	CTC	CGT Arg	CAG Gln	GGT Gly 1680	5276
TAC Tyr	ACC Thr	ATC Ile	CGC Arg	ATT Ile 1685	CTT Leu	CTC Leu	TGG Trp	ACC Thr	TTT Phe 169	Val	CAG Gln	TCC Ser	TTC Phe	AAG Lys 169	Ala	5324
CTG Leu	CCT Pro	TAT Tyr	GTC Val 1700	Cys	CTG Leu	CTG Leu	ATC Ile	GCC Ala 170	Met	CTC Leu	TTC Phe	TTC Phe	ATC Ile 171	Tyr	GCC Ala	5372
ATC Ile	ATT Ile	GGG Gly 1715	met	CAG Gln	GTG Val	TTT Phe	GGT Gly 1720	Asn	ATT Ile	GGC Gly	ATC Ile	GAC Asp 172	Val	GAG Glu	GAC Asp	5420
GAG Glu	GAC Asp 1730	ser	GAT Asp	GAA Glu	GAT Asp	GAG Glu 1735	Phe	CAA Gln	ATC Ile	ACT Thr	GAG Glu 1740	His	AAT Asn	AAC Asn	TTC Phe	5468
CGG Arg 1745	Inr	TTC Phe	TTC Phe	GIn	GCC Ala 1750	Leu	ATG Met	CTT Leu	CTC Leu	TTC Phe 1755	Arg	AGT Ser	GCC Ala	ACC Thr	GGG Gly 1760	5516
GAA (GCT Ala	TGG Trp	HIS	AAC Asn 1765	тте	ATG Met	CTT Leu	TCC Ser	TGC Cys 1770	Leu	AGC Ser	GGG Gly	AAA Lys	CCG Pro 1775	Cys	5564
GAT A	AAG Lys	ASI	TCT Ser 1780	GTA	ATC Ile:	CTG Leu	Thr	CGA Arg 1785	Glu	TGT Cys	GGC Gly	AAT Asn	GAA Glu 1790	Phe	GCT Ala	5612
TAT :	TTT	TAC	TTT (GTT '	TCC '	TTC .	ATC	TTC	CTC	TGC	TCG	TTT	CTG	ATG	CTG	5660

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Tyr	Phe	Tyr 1795		Val	Ser	Phe	Ile 1800		Leu	Cys	Ser	Phe 1805		Met	Leu	
		Phe					Met					TAC Tyr				5708
	Ser					Pro					Glu	TAC Tyr				5756
					Pro					Arg		CCT Pro			Asp	5804
				Leu					Pro			GGT Gly		Gly		5852
			Ala					Lys				CGG Arg 1885	Met			5900
		Ala					Val					ACC Thr				5948
CTG Leu 1905	Ile	CGC Arg	ACA Thr	GCC Ala	CTG Leu 1910	Asp	ATC Ile	AAG Lys	ATT Ile	GCC Ala 1915	Lys	GGA Gly	GGA Gly	GCC Ala	GAC Asp 1920	5996
					Ala					Glu		ATG Met			Trp	6044
CCC Pro	AAT Asn	CTG Leu	TCC Ser 1940	Gln	AAG Lys	ACG Thr	CTA Leu	GAC Asp 194	Leu	CTG Leu	GTC Val	ACA Thr	CCT Pro 1950	His	AAG Lys	6092
TCC Ser	ACG Thr	GAC Asp 195	Leu	ACC Thr	GTG Val	GGG Gly	AAG Lys 1960	Ile	TAC Tyr	GCA Ala	GCC Ala	ATG Met 1965	Met	ATC Ile	ATG Met	6140
GAG Glu	TAC Tyr 1970	Tyr	CGG Arg	CAG Gln	AGC Ser	AAG Lys 197!	Ala	AAG Lys	AAG Lys	CTG Leu	CAG Gln 198	GCC Ala	ATG Met	CGC Arg	GAG Glu	6188
GAG Glu 198	Gln	GAC Asp	CGG Arg	ACA Thr	CCC Pro 1990	Leu	ATG Met	TTC Phe	CAG Gln	CGC Arg 199	Met	GAG Glu	CCC Pro	CCG Pro	TCC Ser 2000	6236
CCA Pro	ACG Thr	CAG Gln	GAA Glu	GGG Gly 200!	Gly	CCT Pro	GGC Gly	CAG Gln	AAC Asn 201	Ala	CTC Leu	CCC Pro	TCC Ser	ACC Thr 201	Gln	6284
CTG	GAC	CCA	GGA	GGA	GCC	CTG	ATG	GCT	CAC	GAA	AGC	GGC	CTC	AAG	GAG	6332

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Leu	Asp	Pro	Gly 202	Gly 0	Ala	Leu	Met	Ala 202	His 5	Glu	Ser	Gly	Leu 203		Glu	
AGC Ser	CCG	TCC Ser 203	Trp	GTG Val	ACC Thr	CAG Gln	CGT Arg 204	Ala	CAG Gln	GAG Glu	ATG Met	Phe 204	Gln	AAG Lys	ACG Thr	6380
GGC Gly	ACA Thr 205	Trp	AGT Ser	CCG Pro	GAA Glu	CAA Gln 205	Gly	CCC Pro	CCT Pro	ACC	GAC Asp 206	Met	CCC Pro	AAC Asn	AGC Ser	6428
CAG Gln 206	PIO	AAC Asn	TCT Ser	CAG Gln	TCC Ser 207	Val	GAG Glu	ATG Met	CGA Arg	GAG Glu 207	Met	GGC Gly	AGA Arg	GAT Asp	GGC Gly 2080	6476
TAC Tyr	TCC Ser	GAC Asp	AGC Ser	GAG Glu 208	Hls	TAC Tyr	CTC Leu	CCC Pro	ATG Met 209	Glu	GGC Gly	CAG Gln	GGC Gly	CGG Arg 209		6524
GCC Ala	TCC Ser	ATG Met	CCC Pro 210	Arg	CTC Leu	CCT Pro	GCA Ala	GAG Glu 210	Asn	CAG Gln	AGG Arg	AGA Arg	AGG Arg 211	Gly	CGG Arg	6572
CCA Pro	CGT Arg	GGG Gly 211	AAT Asn 5	AAC Asn	CTC Leu	AGT Ser	ACC Thr 2120	Ile	TCA Ser	GAC Asp	ACC Thr	AGC Ser 212	Pro	ATG Met	AAG Lys	6620
CGT Arg	TCA Ser 213	Ата	TCC Ser	GTG Val	CTG Leu	GGC Gly 2135	Pro	AAG Lys	GCC Ala	CGA Arg	CGC Arg 2140	Leu	GAC Asp	GAT Asp	TAC Tyr	6668
TCG Ser 2145	Leu	GAG Glu	CGG Arg	GTC Val	CCG Pro 2150	Pro	GAG Glu	GAG Glu	AAC Asn	CAG Gln 2155	Arg	CAC His	CAC His	CAG Gln	CGG Arg 2160	6716
CGC Arg	CGC Arg	GAC Asp	CGC Arg	AGC Ser 2165	His	CGC Arg	GCC Ala	TCT Ser	GAG Glu 2170	Arg	TCC Ser	CTG Leu	GGC Gly	CGC Arg 2175	Tyr	6764
ACC Thr	GAT Asp	GTG Val	GAC Asp 2180	Thr	GGC Gly	TTG Leu	GGG Gly	ACA Thr 2185	Asp	CTG Leu	AGC Ser	ATG Met	ACC Thr 2190	Thr	CAA Gln	6812
TCC Ser	GGG Gly	GAC Asp 2195	CTG Leu	CCG Pro	TCG Ser	Lys	GAG Glu 2200	Arg	GAC Asp	CAG Gln	GAG Glu	CGG Arg 2205	Gly	CGG Arg	CCC Pro	6860
AAG Lys	GAT Asp 2210	Arg	AAG Lys	CAT His	Arg	CAG Gln 2215	CAC His	CAC His	CAC His	CAC His	CAC His 2220	His	CAC His	CAC His	CAC His	6908
CAT His 2225	Pro	CCG Pro	CCC Pro	Pro	GAC Asp 2230	Lys .	GAC Asp	CGC Arg	Tyr	GCC Ala 2235	Gln	GAA Glu	CGG Arg	CCG Pro	GAC Asp 2240	6956
CAC	GGC	CGG	GCA	CGG	GCT	CGG (GAC	CAG	CGC	TGG	TCC	CGC	TCG	ccc	AGC	7004

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Ні	s Gly	Arg	Ala	Arg 224		Arg	Asp	Gln	Arg 225		Ser	Arg	Ser	Pro 225			
	G GGC			His					Gln					Val		705	52
	A AGC y Ser		Ala					Gly					Arg			710	00
	C CGC G Arg 229	Gln					Pro					Pro				714	8 :
Ту	T TCC r Ser 05					Lys					Gly					719	6
	G CAG n Gln				Gln					Val					Arg	724	. 4
	G GCC a Ala			Gly					Pro					Glu		729	2
	G GCC u Ala		Asp					Gly					Gly			734	. O
	C AGG O Arg 237	Met					Pro					Ser				738	8
Ar	G GCC g Ala 85					Gly					Ala					743	.6
G1 Va	G TCC	GAG Glu	GGG Gly	CCC Pro 240	Pro	GGT Gly	CCC Pro	CGG Arg	CAC His 241	His	GGC Gly	TAC Tyr	TAC Tyr	CGG Arg 241	Gly	748	4
TC Se	C GAC	TAC Tyr	GAC Asp 242	Glu	GCC Ala	GAT Asp	GGC Gly	CCG Pro 242	Gly	AGC Ser	GGG Gly	GGC Gly	GGC Gly 2430	Glu	GAG Glu	753	
GC Al	C ATG	GCC Ala 243	Gly	GCC Ala	TAC Tyr	GAC Asp	GCG Ala 244	Pro	CCC Pro	CCC Pro	GTA Val	CGA Arg 244	His	GCG Ala	TCC Ser	758	30
	G GGC r Gly 245	Ala					Pro					Ala				762	28
GC	C TGC	GCC	TCG	CCT	TCT	CGG	CAC	GGC	CGG	CGA	CTC	CCC	AAC	GGC	TAC	767	76

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Ala Cys Ala Ser Pro Ser Arg His Gly Arg Arg Leu Pro Asn Gly Tyr 2465 2470 2475 2480	
TAC CCG GCG CAC GGA CTG GCC AGG CCC CGC GGG CCG GGC TCC AGG AAG Tyr Pro Ala His Gly Leu Ala Arg Pro Arg Gly Pro Gly Ser Arg Lys 2485 2490 2495	7724
GGC CTG CAC GAA CCC TAC AGC GAG AGT GAC GAT GAT TGG TGC TAAGCCCGGC Gly Leu His Glu Pro Tyr Ser Glu Ser Asp Asp Asp Trp Cys 2505 2510	7776
CGAGGTGGCG CCCCCACGCA CC	7808
(2) INFORMATION FOR SEQ ID NO:23:	
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 7791 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(ix) FEATURE: (A) NAME/KEY: CDS	
(B) LOCATION: 2377037(D) OTHER INFORMATION: /standard_name= "Alpha-1A-2"	
,	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:23:	
GATGTCCCGA GCTGCTATCC CCGGCTCGGC CCGGGCAGCC GCCTTCTGAG CCCCCGACCC	60
GAGGCGCCGA GCCGCCGCCG CCCGATGGGC TGGGCCGTGG AGCGTCTCCG CAGTCGTAGC	120
TCCAGCCGCC GCGCTCCCAG CCCCGGCAGC CTCAGCATCA GCGGCGGCGG CGGCGGCGGC	180
GGCGTCTTCC GCATCGTTCG CCGCAGCGTA ACCCGGAGCC CTTTGCTCTT TGCAGA	236
ATG GCC CGC TTC GGA GAC GAG ATG CCG GCC CGC TAC GGG GGA GGC Met Ala Arg Phe Gly Asp Glu Met Pro Ala Arg Tyr Gly Gly Gly 1 5 10	284
TCC GGG GCA GCC GCG GTG GTC GTG GGC AGC GGA GGC GGG CGA GGA Ser Gly Ala Ala Gly Val Val Val Ser Gly Gly Gly Arg Gly 20 25 30	332
GCC GGG GGC AGC CGG CAG GGC GGG CAG CCC GGG GCG CAA AGG ATG TAC Ala Gly Gly Ser Arg Gln Gly Gln Pro Gly Ala Gln Arg Met Tyr 35 40 45	380
AAG CAG TCA ATG GCG CAG AGA GCG CGG ACC ATG GCA CTC TAC AAC CCC Lys Gln Ser Met Ala Gln Arg Ala Arg Thr Met Ala Leu Tyr Asn Pro 50 55 60	428

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	CCC Pro															476
	AGC Ser															. 524
	CCT Pro															572
	GTC Val															620
	TCT Ser 130															668
	TTC Phe															716
	GGC Gly															764
	CTA Leu															812
	CTG Leu															860
	CCA Pro 210															908
CCT Pro 225	TTG Leu	CTG Leu	CAG Gln	ATC Ile	GGC Gly 230	CTC Leu	CTC Leu	CTA Leu	TTT Phe	TTT Phe 235	GCA Ala	ATC Ile	CTT Leu	ATT Ile	TTT Phe 240	956
GCA Ala	ATC Ile	ATA Ile	GGG Gly	TTA Leu 245	GAA Glu	TTT Phe	TAT Tyr	ATG Met	GGA Gly 250	AAA Lys	TTT Phe	CAT His	ACC Thr	ACC Thr 255	TGC Cys	1004
TTT Phe	GAA Glu	GAG Glu	GGG Gly 260	ACA Thr	GAT Asp	GAC Asp	ATT Ile	CAG Gln 265	GGT Gly	GAG Glu	TCT Ser	CCG Pro	GCT Ala 270	CCA Pro	TGT Cys	1052
GGG Gly	ACA Thr	GAA Glu 275	GAG Glu	CCC Pro	GCC Ala	CGC Arg	ACC Thr 280	TGC Cys	CCC Pro	AAT Asn	GGG Gly	ACC Thr 285	AAA Lys	TGT Cys	CAG Gln	1100

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CC Pr	C TA o Ty 29		G GA p Gl	A GG u Gl	G CCC	C AAG Asi 29	1 W21	GGG GGI	G ATO	C ACT	T CAG	n Ph	C GA	C AZ p As	AC AT	C 1148
CT Lev 30	G TT u Ph	T GC e Al	A GT a Va	G CT	G ACT u Thi 310	. val	TTC Phe	CAC Glr	TGC Cys	C ATA 5 Ile 315	e Thi	C AT	G GA t Gl	A GG u Gl	G TG y Trj 320	Þ
				32!	5	. Ser	ASI	ASE	330	ser	. GIÀ	/ As:	n Th	r Tr 33		1
		- 3	34	0		, nea	. 116	345	ite	GIY	Ser	Phe	9 Ph	e Me	G CTG t Lev	İ
		35	5	- 01,	vai	Deu	360	GIY	GIU	Pne	Ala	365	Gli G	ı Ar	G GAA g Glu	
_	370)			ni g	375	Pne	Leu	ьуs	Leu	Arg 380	Arg	, Glr	Gli	A CAG	
385			, 021		390	GIY	IYL	Met	GIU	395	IIe	Ser	Lys	Ala	GAA Glu 400	1436
			Leu	405	GIU	Asp	GIU	Thr	Asp 410	Gly	Glu	Gln	Arg	His 415		1484
		1	420	Deu	CGG Arg	Arg	inr	425	ııe	гàг	Lys	Ser	Lys 430	Thr	Asp	1532
		435	110	GIU	GAG Glu	Ald	440	Asp	GIN	Leu	Ala	Asp 445	Ile	Ala	Ser	1580
GTG Val	GGT Gly 450	TCT Ser	CCC Pro	TTC Phe	GCC Ala	CGA Arg 455	GCC Ala	AGC Ser	ATT . Ile	ьуs	AGT Ser 460	GCC Ala	AAG Lys	CTG Leu	GAG Glu	1628
AAC Asn 465	TCG Ser	ACC Thr	TTT Phe	TTT Phe	CAC His 470	AAA . Lys .	AAG (Lys (GAG . Glu .	Arg .	AGG Arg i 475	ATG Met	CGT Arg	TTC Phe	TAC Tyr	ATC Ile 480	1676
CGC Arg	CGC Arg	ATG Met	GTC Val	AAA Lys 485	ACT (CAG (Gln)	GCC : Ala 1	Phe '	TAC ! Tyr ! 490	TGG /	ACT (GTA Val	CTC Leu	AGT Ser 495	TTG Leu	1724
GTA Val	GCT Ala	CTC Leu	AAC Asn 500	ACG Thr	CTG : Leu (TGT (Cys \	var z	GCT A Ala : 505	ATT (Ile \	GTT (Val I	CAC '	TAC Tyr	AAC Asn 510	CAG Gln	CCC Pro	1772

GAG Glu	TGG Trp	CTC Leu 515	TCC Ser	GAC Asp	TTC Phe	CTT Leu	TAC Tyr 520	TAT Tyr	GCA Ala	GAA Glu	TTC Phe	ATT Ile 525	TTC Phe	TTA Leu	GGA Gly	1820
CTC Leu	TTT Phe 530	ATG Met	TCC Ser	GAA Glu	ATG Met	TTT Phe 535	ATA Ile	AAA Lys	ATG Met	TAC Tyr	GGG Gly 540	CTT Leu	GGG Gly	ACG Thr	CGG Arg	1868
CCT Pro 545	TAC Tyr	TTC Phe	CAC His	TCT Ser	TCC Ser 550	TTC Phe	AAC Asn	TGC Cys	TTT Phe	GAC Asp 555	TGT Cys	GGG Gly	GTT Val	ATC Ile	ATT Ile 560	1916
GGG Gly	AGC Ser	ATC Ile	TTC Phe	GAG Glu 565	GTC Val	ATC Ile	TGG Trp	GCT Ala	GTC Val 570	ATA Ile	AAA Lys	CCT Pro	GGC Gly	ACA Thr 575	TCC Ser	1964
TTT Phe	GGA Gly	ATC Ile	AGC Ser 580	GTG Val	TTA Leu	CGA Arg	GCC Ala	CTC Leu 585	AGG Arg	TTA Leu	TTG Leu	CGT Arg	ATT Ile 590	TTC Phe	AAA Lys	2012
Val	Thr	Lys 595	Tyr	Trp	GCA Ala	Ser	Leu 600	Arg	Asn	Leu	Val	Val 605	Ser	Leu	Leu	2060
AAC Asn	TCC Ser 610	ATG Met	AAG Lys	TCC Ser	ATC Ile	ATC Ile 615	AGC Ser	CTG Leu	TTG Leu	TTT Phe	CTC Leu 620	CTT Leu	TTC Phe	CTG Leu	TTC Phe	2108
Ile 625	Val	Val	Phe	Ala	CTT Leu 630	Leu	Gly	Met	Gln	Leu 635	Phe	Gly	Gly	GIn	Phe 640	2156
Asn	Phe	Asp	Glu	Gly 645	ACT Thr	Pro	Pro	Thr	Asn 650	Phe	Asp	Thr	Phe	655	Ala	2204
Ala	Ile	Met	Thr 660	Val	TTT Phe	Gln	Ile	Leu 665	Thr	Gly	Glu	Asp	670	Asn	GIU	2252
Val	Met	Tyr 675	Asp	Gly	Ile	Lys	Ser 680	Gln	Gly	Gly	Val	Gln 685	Gly	GIA		2300
GTG Val	TTC Phe 690	Ser	ATC Ile	TAT	TTC Phe	ATT Ile 695	GTA Val	CTG Leu	ACG Thr	CTC Leu	TTT Phe 700	GGG	AAC Asn	TAC Tyr	ACC Thr	2348
CTC Leu 705	Leu	AAT Asn	GTG Val	TTC Phe	TTG Leu 710	Ala	ATC Ile	GCT Ala	GTG Val	GAC Asp 715	Asn	CTG Leu	GCC Ala	AAC Asn	GCC Ala 720	2396
CAG Gln	GAG Glu	CTC Leu	ACC Thr	AAG Lys 725	Val	GAG Glu	GCG Ala	GAC Asp	GAG Glu 730	Gln	GAG Glu	GAA Glu	GAA Glu	GAA Glu 735	GCA Ala	2444

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GCG Ala	AAC Asn	CAG Gln	AAA Lys 740	Leu	GCC Ala	CTA Leu	CAG Gln	AAA Lys 745	Ala	AAG Lys	GA0 Glu	GTG Val	GCA Ala 750	Glu	GTG Val	2492
AGT Ser	CCT Pro	CTG Leu 755	TCC	GCG Ala	GCC Ala	AAC Asn	ATG Met 760	TCT Ser	ATA Ile	GCT Ala	GTG Val	AAA Lys 765	GAG Glu	CAA Gln	CAG Gln	2540
AAG Lys	AAT Asn 770	GIN	AAG Lys	CCA Pro	GCC Ala	AAG Lys 775	TCC Ser	GTG Val	TGG Trp	GAG Glu	CAG Gln 780	CGG Arg	ACC Thr	AGT Ser	GAG Glu	2588
ATG Met 785	CGA Arg	AAG Lys	CAG Gln	AAC Asn	TTG Leu 790	CTG Leu	GCC Ala	AGC Ser	CGG Arg	GAG Glu 795	GCC Ala	CTG Leu	TAT Tyr	AAC Asn	GAA Glu 800	2636
ATG Met	GAC Asp	CCG Pro	GAC Asp	GAG Glu 805	CGC Arg	TGG Trp	AAG Lys	GCT Ala	GCC Ala 810	TAC	ACG Thr	CGG Arg	CAC His	CTG Leu 815	CGG Arg	2684
CCA Pro	GAC Asp	ATG Met	AAG Lys 820	ACG Thr	CAC His	TTG Leu	GAC Asp	CGG Arg 825	CCG Pro	CTG Leu	GTG Val	GTG Val	GAC Asp 830	CCG Pro	CAG Gln	2732
GIU	ASN	835	Asn	Asn	Asn	Thr	Asn 840	Lys	Ser	Arg	Ala	GCC Ala 845	Glu	Pro	Thr	2780
GTG Val	GAC Asp 850	CAG Gln	CGC Arg	CTC Leu	GGC Gly	CAG Gln 855	CAG Gln	CGC Arg	GCC Ala	GAG Glu	GAC Asp 860	TTC Phe	CTC Leu	AGG Arg	AAA Lys	2828
865	Ala	Arg	Tyr	His	870	Arg	Ala	Arg	Asp	Pro 875	Ser	GGC Gly	Ser	Ala	880	2876
CTG Leu	GAC Asp	GCA Ala	CGG Arg	AGG Arg 885	CCC Pro	TGG Trp	GCG Ala	GGA Gly	AGC Ser 890	CAG Gln	GAG Glu	GCC Ala	GAG Glu	CTG Leu 895	AGC Ser	2924
CGG Arg	GAG Glu	GGA Gly	CCC Pro 900	TAC Tyr	GGC Gly	CGC Arg	GAG Glu	TCG Ser 905	GAC Asp	CAC His	CAC His	GCC Ala	CGG Arg 910	GAG Glu	GGC Gly	2972
AGC Ser	CTG Leu	GAG Glu 915	CAA Gln	CCC Pro	GGG Gly	Phe	TGG Trp 920	GAG Glu	GGC Gly	GAG Glu	GCC Ala	GAG Glu 925	CGA Arg	GGC Gly	AAG Lys	3020
GCC Ala	GGG Gly 930	GAC Asp	CCC Pro	CAC His	Arg .	AGG Arg 935	CAC His	GTG Val	CAC His	Arg	CAG Gln 940	GGG Gly	GGC Gly	AGC Ser	AGG Arg	3068
GAG Glu 945	AGC Ser	CGC Arg	AGC Ser	Gly	TCC Ser 950	CCG Pro .	CGC . Arg '	ACG Thr	Gly .	GCG Ala 955	GAC Asp	GGG Gly	GAG Glu	His	CGA Arg 960	3116

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	CAT His															316	54
	GAG Glu															. 321	L2
	GAG Glu							Asp					Arg			326	50
	CGG Arg 1010	His					Thr					Ala				330	8(
	AAG Lys 5					Arg					Asn					335	6
	CCT Pro				Pro					Thr					Gln	340)4
	CTG Leu			Gln					Ala					Asn		345	32
	AAC Asn		Lys					Glu					His			350	00
	GGC Gly 109	His					Gln					Met				354	18
ACC Thr 110	GAC Asp 5	CCC Pro	GGC Gly	CCC Pro	ATG Met 1110	Leu	GCC Ala	ATC Ile	CCT Pro	GCC Ala 111	Met	GCC Ala	ACC Thr	AAC Asn	CCC Pro 1120	359	96
	AAC Asn				Arg					Asn					Ser	364	14
AAT Asn	CCC Pro	GGC Gly	CCC Pro 114	Pro	AAG Lys	ACC Thr	CCC Pro	GAG Glu 114	Asn	AGC Ser	CTT Leu	ATC Ile	GTC Val 115	Thr	AAC Asn	369	}2
CCC Pro	AGC Ser	GGC Gly 115	Thr	CAG Gln	ACC Thr	AAT Asn	TCA Ser 116	Ala	AAG Lys	ACT Thr	GCC Ala	AGG Arg 116	Lys	CCC Pro	GAC Asp	374	10
CAC His	ACC Thr 117	Thr	GTG Val	GAC Asp	ATC Ile	CCC Pro 117	Pro	GCC Ala	TGC Cys	CCA Pro	CCC Pro 118	Pro	CTC Leu	AAC Asn	CAC His	378	38

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ACC (Thr V	GTC Val	GTA Val	CAA Gln	GTG Val	AAC Asn 119	rys	AAC Asn	GCC	AAC Asr	CCA Pro) Asp	C CCA	A CTO	G CCA	A AAA D Lys 1200	;	3836
AAA (GAG Glu	GAA Glu	GAG Glu	AAG Lys 120	ьys	GAG Glu	GAG Glu	GAG Glu	GAA Glu 121	. Asp	GAC Asp	C CGI	Gly GGG	G GAA / Glu 121	Asp	:	3884
GGC C	Pro	AAG Lys	CCA Pro 122	met	CCT Pro	CCC Pro	TAT Tyr	AGC Ser 122	Ser	Met	TTC Phe	ATC Ile	CTG Leu 123	Ser	ACG Thr	3	3932
ACC A	AAC Asn	CCC Pro 123	ьeu	CGC Arg	CGC Arg	CTG Leu	TGC Cys 124	His	TAC Tyr	ATC Ile	CTG Leu	AAC Asn 124	Leu	CGC Arg	TAC Tyr	3	3980
TTT G Phe G	SAG Slu .250	MEC	TGC Cys	ATC Ile	CTC Leu	ATG Met 125	Val	ATT Ile	GCC Ala	ATG Met	AGC Ser 126	Ser	ATC	GCC Ala	CTG Leu	4	028
GCC G Ala A 1265	CC la	GAG Glu	GAC Asp	CCT Pro	GTG Val 1270	GIN	CCC Pro	AAC Asn	GCA Ala	CCT Pro 127	Arg	AAC Asn	AAC Asn	GTG Val	CTG Leu 1280	4	076
CGA T	Ϋ́	Pne	Asp	1285	Val	Phe	Thr	Gly	Val 129	Phe D	Thr	Phe	Glu	Met 129	Val 5	4	124
ATC A	ys	Met	1300	Asp	ren	GIÀ	Leu	Val 130	Leu	His	Gln	Gly	Ala 131	Tyr	Phe	4	172
CGT G	Sp.	CTC Leu 1315	rrb	AAT Asn	ATT Ile	CTC Leu	GAC Asp 1320	Phe	ATA Ile	GTG Val	GTC Val	AGT Ser 1325	Gly	GCC Ala	CTG Leu	4:	220
GTA GO Val Al	CC la 330	Pne .	GCC Ala	TTC Phe	Thr	GGC Gly 1335	AAT Asn	AGC Ser	AAA Lys	GGA Gly	AAA Lys 1340	Asp	ATC Ile	AAC Asn	ACG Thr	4:	268
ATT AN Ile Ly 1345	AA '	TCC Ser	CTC Leu	Arg	GTC Val 1350	CTC Leu	CGG Arg	GTG Val	CTA Leu	CGA Arg 1355	Pro	CTT Leu	AAA Lys	ACC Thr	ATC Ile 1360	43	316
AAG CO	GG (rg]	CTG (Leu :	Pro .	AAG Lys : 1365	CTC . Leu	AAG Lys .	GCT Ala	Val	TTT Phe 1370	Asp	TGT Cys	GTG Val	Val	AAC Asn 1375	Ser	43	364
CTT AA Leu Ly	AA 1 /s 1	ASD Y	GTC ' Val 1 1380	Phe	AAC . Asn	ATC	Leu :	ATC Ile 1385	Val	TAC Tyr	ATG Met	Leu	TTC Phe 1390	Met	TTC Phe	. 44	112
ATC TI Ile Ph	ne A	SCC (Ala N L395	STG (/al \	GTG (/al /	GCT (Ala '	Val (CAG (Sln 1	CTC Leu	TTC . Phe	AAG Lys	Gly	AAA Lys 1405	Phe	TTC Phe	CAC His	44	160

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•	TGC Cys	ACT Thr 1410	Asp	GAG Glu	TCC Ser	AAA Lys	GAG Glu 1415	Phe	GAG Glu	AAA Lys	Asp	TGT Cys 1420	Arg	GGC Gly	AAA Lys	TAC Tyr	45	80
	CTC Leu 1425	Leu	TAC Tyr	GAG Glu	AAG Lys	AAT Asn 1430	Glu	GTG Val	AAG Lys	GCG Ala	CGA Arg 1435	Asp	CGG Arg	GAG Glu	TTP	AAG Lys 1440	45	56
	AAG Lys	TAT Tyr	GAA Glu	TTC Phe	CAT His 144	TAC Tyr 5	GAC Asp	AAT Asn	GTG Val	CTG Leu 1450	Trp	GCT Ala	CTG Leu	CTG Leu	ACC Thr 1455	neu	46	04
	TTC Phe	ACC Thr	GTG Val	TCC Ser 146	Thr	GGA Gly	GAA Glu	GGC Gly	TGG Trp 146	Pro	CAG Gln	GTC Val	CTC Leu	AAG Lys 1470	UTO	TCG Ser	46	52
	GTG Val	GAC Asp	GCC Ala 147	Thr	TTT Phe	GAG Glu	AAC Asn	CAG Gln 1480	GTA	CCC Pro	AGC Ser	CCC Pro	GGG Gly 1485	TAT	CGC Arg	ATG Met	47	00
	GAG Glu	ATG Met 149	Ser	ATT	TTC Phe	TAC Tyr	GTC Val 149	Val	TAC Tyr	TTT Phe	GTG Val	GTG Val 150	Pile	CCC Pro	TTC Phe	TTC Phe	47	748
	TTT Phe 150	Val	AAT Asn	ATC Ile	TTT Phe	GTG Val 151	Ala	TTG Leu	ATC Ile	ATC Ile	ATC Ile 151	THI	TTC Phe	CAG Gln	GAG Glu	CAA Gln 1520	47	796
	GGG Gly	GAC Asp	AAG Lys	ATG Met	ATG Met 152	GAG Glu 5	GAA Glu	TAC Tyr	AGC Ser	CTG Leu 153	GIU	AAA Lys	AAT Asn	GAG Glu	AGG Arg 153		48	344
	TGC Cys	ATT Ile	GAT Asp	TTC Phe 154	Ala	ATC	AGC Ser	GCC Ala	AAG Lys 154	Pro	CTG Leu	ACC Thr	CGA Arg	CAC His 155	MEC	CCG Pro	41	892
	CAG Gln	AAC Asi	AA0 1 Lys 15	Gln	AGC Ser	TTC Phe	CAG Gln	TAC Tyr 156	Arg	ATG Met	TGG Trp	CAG Gln	TTC Phe 156	Val	GTG Val	TCT Ser	4:	940
	CCG	CCT Pro	o Phe	GAG Glu	TAC	ACG Thr	ATC Ile 157	Mec	GCC Ala	ATG Met	ATC Ile	GCC Ala 158		AAC Asn	ACC Thr	ATC Ile	4	988
	GTG Val	. Le	r ATO	G ATO	AAC Lys	TTC Phe 159	Tyr	GGG Gly	GCT Ala	TCT a Ser	GTI Val		TAT Tyr	GAP Glu	AAT ASN	GCC Ala 1600	5	036
	CT(Let	G CG	G GT g Va	G TTO	C AAG ASI 160	n Ile	GTC Val	TTC Phe	ACC Thi	TCC r Sei 161	nec	TTO Phe	TCI Ser	CTC Lev	GAA Glu 161	TGT Cys .5	5	084
	GT(Va	G CT Le	G AA u Ly	A GT s Va 16	1 Me	G GCT t Ala	r TTT	r GGG e Gly	3 AT' 7 Il 16:	e ne	AA E 1 Asi	r TAT	r TTO	C CGG Arg 16:	J L	GCC Ala	5	3132

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	1635	T Tul	1640		lle Thr Asp 1645	Ile
1650		1655	FIO ASII A	AC TTC ATC ASS Phe 11660	sn Leu Ser	Phe
1665		1670	arg bed 1.	TC AAA CTT C le Lys Leu L 1675	eu Arg Gln	Gly 1680
	10	685	16	TT GTG CAG T ne Val Gln S 90	er Phe Lys 1699	Ala
	1700	, - Lou Deu 1	1705	G CTC TTC T t Leu Phe P	he Ile Tyr 1710	Ala
1	.715	1	720		sp Val Glu 725	Asp
1730		1735	ne GIN II	C ACT GAG CA e Thr Glu Hi 1740	s Asn Asn	Phe
1745		1750	er neu ne	C TTC CGG AG u Phe Arg Se 1755	r Ala Thr	Gly 1760
GAA GCT T	GG CAC AA rp His As: 17	176	TT TCC TGG eu Ser Cys 17	C CTC AGC GG E Leu Ser Gl 70	G AAA CCG : y Lys Pro (rgt 5564 Cys
GAT AAG AA ASP Lys Aa	AC TCT GGG sn Ser Gli 1780	C ATC CTG AC y Ile Leu Th	T CGA GAO T Arg Glu 1785	G TGT GGC AA 1 Cys Gly As:	T GAA TTT (n Glu Phe <i>I</i> 1790	SCT 5612 Ma
	AC TTT GTT /r Phe Val /95	. DCT FIFE II	C TTC CTC e Phe Leu 00	TGC TCG TT Cys Ser Pho	e Leu Met I	ETG 5660 eu
AAT CTC TT Asn Leu Ph 1810	CT GTC GCC ie Val Ala	GTC ATC AT Val Ile Me 1815	G GAC AAC t Asp Asn	TTT GAG TAG Phe Glu Tyr 1820	C CTC ACC C Leu Thr A	GA 5708 rg
GAC TCC TC Asp Ser Se 1825	C ATC CTG r Ile Leu	GGC CCC CA Gly Pro Hi 1830	C CAC CTG s His Leu	GAT GAG TAC Asp Glu Tyr 1835	. Val Arg V	TC 5756 al 840
TGG GCC GA Trp Ala Gl	G TAT GAC u Tyr Asp 184	+10 MIG AL	T TGG GGC a Trp Gly 185	CGC ATG CCT Arg Met Pro		

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Met Tyr Gln M	ATG CTG AGA CA Met Leu Arg Hi 1860	C ATG TCT CO s Met Ser Pr 1865	CG CCC CTG GG ro Pro Leu Gl	GT CTG GGG ly Leu Gly 1870	AAG 5852 Lys
AAG TGT CCG G Lys Cys Pro A 1875	GCC AGA GTG GC Ala Arg Val Al	T TAC AAG CO a Tyr Lys Ai 1880	rg Leu Leu Ar	GG ATG GAC rg Met Asp 885	CTG 5900 Leu
CCC GTC GCA G Pro Val Ala A 1890	GAT GAC AAC AC Asp Asp Asn Th 18	r Val His Ph	TC AAT TCC AC ne Asn Ser Th 1900	CC CTC ATG	GCT 5948 Ala
CTG ATC CGC A Leu Ile Arg 7 1905	ACA GCC CTG GA Thr Ala Leu As 1910	C ATC AAG AT p lle Lys II	TT GCC AAG GO le Ala Lys GJ 1915	GA GGA GCC ly Gly Ala	GAC 5996 Asp 1920
AAA CAG CAG A Lys Gln Gln N	ATG GAC GCT GA Met Asp Ala Gl 1925	u Leu Arg Ly	AG GAG ATG AT ys Glu Met Me 930	IG GCG ATT et Ala Ile 1935	Trp
Pro Asn Leu S	TCC CAG AAG AC Ser Gln Lys Th 1940	G CTA GAC C r Leu Asp Le 1945	TG CTG GTC ACeu Leu Val Ti	CA CCT CAC hr Pro His 1950	AAG 6092 Lys
TCC ACG GAC C Ser Thr Asp I 1955	CTC ACC GTG GG Leu Thr Val Gl	G AAG ATC TA y Lys Ile To 1960	yr Ala Ala Me	TG ATG ATC et Met Ile 965	ATG 6140 Met
GAG TAC TAC (Glu Tyr Tyr A 1970	CGG CAG AGC AA Arg Gln Ser Ly 19	G GCC AAG A s Ala Lys L 75	AG CTG CAG GO ys Leu Gln Al 1980	CC ATG CGC la Met Arg	GAG 6188 Glu
GAG CAG GAC (Glu Gln Asp / 1985	CGG ACA CCC CT Arg Thr Pro Le 1990	C ATG TTC C. u Met Phe G	AG CGC ATG G ln Arg Met G 1995	AG CCC CCG lu Pro Pro	TCC 6236 Ser 2000
CCA ACG CAG (Pro Thr Gln (GAA GGG GGA CC Glu Gly Gly Pr 2005	o Gly Gln A	AC GCC CTC Co sn Ala Leu P: 010	CC TCC ACC ro Ser Thr 2015	Gin
Leu Asp Pro (GGA GGA GCC CT Gly Gly Ala Le 2020	G ATG GCT C u Met Ala H 2025	AC GAA AGC G is Glu Ser G	GC CTC AAG ly Leu Lys 2030	GAG 6332 Glu
AGC CCG TCC ' Ser Pro Ser ' 2035	TGG GTG ACC CA	G CGT GCC C n Arg Ala G 2040	ln Glu Met P	TC CAG AAG he Gln Lys 045	ACG 6380 Thr
GGC ACA TGG : Gly Thr Trp : 2050	AGT CCG GAA CA Ser Pro Glu Gl 20	A GGC CCC C n Gly Pro P 055	CT ACC GAC A ro Thr Asp M 2060	TG CCC AAC let Pro Asn	AGC 6428 Ser
CAG CCT AAC Gln Pro Asn 2065	TCT CAG TCC G Ser Gln Ser Va 2070	rg GAG ATG C al Glu Met A	GA GAG ATG G arg Glu Met G 2075	GC AGA GAT ly Arg Asp	GGC 6476 Gly 2080

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TAC	TCC	GAC	AGC	GAG	CAC	TAC	CTC	ccc	C ATC	GAA	4 GGC	CAG	GGC	CGC	GCT		6524
•				208	5	, TAL	ьеп	PIC	209	00 10	i GIA	Gln	Gly	209	g Ala 95		0524
GCC Ala	TCC Ser	ATG Met	Pro 210	5	CTC	Pro	GCA Ala	GA0 Glu 210	ASI	CAG Gln	AGG Arg	AGA Arg	AGG Arg 211	Gly	C CGG		6572
CCA Pro	CGT Arg	GGG Gly 211	AAT Asn 5	AAC Asn	CTC Leu	AGT Ser	ACC Thr 212	116	TCA Ser	GAC Asp	ACC Thr	AGC Ser 212	Pro	ATG Met	AAG Lys		6620
CGT Arg	TCA Ser 213	GCC Ala	TCC	GTG Val	CTG Leu	GGC Gly 213	PIO	AAG Lys	GCC Ala	CGA Arg	CGC Arg 214	Leu	GAC Asp	GAT Asp	TAC Tyr		6668
TCG Ser 2145	CTG Leu	GAG Glu	CGG Arg	GTC Val	CCG Pro 215	CCC Pro	GAG Glu	GAG Glu	AAC Asn	CAG Gln 215	Arg	CAC His	CAC His	CAG Gln	CGG Arg 2160)	6716
CGC Arg	CGC Arg	GAC Asp	CGC Arg	AGC Ser 2165		CGC Arg	GCC Ala	TCT Ser	GAG Glu 2170	Arg	TCC Ser	CTG Leu	GGC Gly	CGC Arg 217	Tyr		6764
ACC Thr	GAT Asp	GTG Val	GAC Asp 2180		GGC Gly	TTG Leu	GGG Gly	ACA Thr 218	Asp	CTG Leu	AGC Ser	ATG Met	ACC Thr 2190	Thr	CAA Gln		6812
TCC Ser		GAC Asp 2195		CCG Pro	TCG Ser	Lys	GAG Glu 2200	Arg	GAC Asp	CAG Gln	GLu	CGG Arg 2205	Gly	CGG Arg	CCC Pro	1	6860
AAG (Lys ;	GAT Asp 2210	CGG Arg	AAG Lys :	CAT His	n g	CAG Gln 2215	CAC His	CAC His	CAC His	Hls	CAC His 2220	His .	CAC His	CAC His	CAC His	(5908
CAT (His) 2225	CCC (CCG Pro	CCC (Pro)		GAC Asp 2230	rys .	GAC (CGC Arg	Tyr	GCC Ala 2235	Gln (GAA Glu	CGG Arg	${ t Pro}$	GAC Asp 2240	E	5956
CAC (GGC (CGG (2	CGG (Arg) 2245	GCT (Ala)	CGG (Arg)	GAC (Asp (SIN	CGC Arg 2250	TGG '	TCC (Ser)	CGC ! Arg !	Ser :	CCC Pro 2255	Ser	7	004
GAG G Glu G	GC (y	GAG (Glu F 2260	CAC A	ATG (Met 1	GCG (Ala H	ils A	CGG Arg 2265	CAG ' Gln	TAGT'	rccg:	ra ao	GTGG:	AAGC	С	7	054
CAGCC	CCCI	C A	ACATO	TGGT	ACC	CAGCA	CTC	CGC	GGCG	GG (CCGCC	GCCI	AG C	rccc	CCAGA	. 7	114
cccc																	174
CGGGG																	234
CCAGC	GGCC	C TC	GGAG	GTAC	CCA	GGCC	CCA	CGG	CCGAC	CC 1	CTGG	CCGG	A GA	TCG	SCCGC		294

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CCACGGGGG CCACAGCAGC GGCCGCTCGC CCAGGATGGA GAGGCGGGTC CCAGGCCCGG	7354
CCCGGAGCGA GTCCCCCAGG GCCTGTCGAC ACGGCGGGGC CCGGTGGCCG GCATCTGGCC	7414
CGCACGTGTC CGAGGGGCCC CCGGGTCCCC GGCACCATGG CTACTACCGG GGCTCCGACT	7474
ACGACGAGGC CGATGGCCCG GGCAGCGGGG GCGGCGAGGA GGCCATGGCC GGGGCCTACG	7534
ACGCGCCACC CCCCGTACGA CACGCGTCCT CGGGCGCCAC CGGGCGCTCG CCCAGGACTC	7594
CCCGGGCCTC GGGCCCGGCC TGCGCCTCGC CTTCTCGGCA CGGCCGGCGA CTCCCCAACG	7654
GCTACTACCC GGCGCACGGA CTGGCCAGGC CCCGCGGGCC GGGCTCCAGG AAGGGCCTGC	7714
ACGAACCCTA CAGCGAGAGT GACGATGATT GGTGCTAAGC CCGGGCGAGG TGGCGCCCGC	7774
CCGGCCCCC ACGCACC	7791
(2) INFORMATION FOR SEQ ID NO:24:	

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 7032 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (ix) FEATURE:
 - (A) NAME/KEY: CDS

- (B) LOCATION: 166..6921
- (D) OTHER INFORMATION: /standard_name= "Alpha-1E-1"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:24:

GCTGCTGCTG CCTCTCCGAA GAGCTCGCGG AGCTCCCCAG	AGGCGGTGGT CCCCGTGCTT 60
GTCTGGATGC GGCTCTGAGT CTCCGTGTGT CTTTCTGCTT	GTTGCTGTGT GCGGGTGTTC 120
GGCCGCGATC ACCTTTGTGT GTCTTCTGTC TGTTTAAACC	TCAGG ATG GCT CGC 174 Met Ala Arg 1
TTC GGG GAG GCG GTG GTC GCC AGG CCA GGG TCC Phe Gly Glu Ala Val Val Ala Arg Pro Gly Ser 5	GGC GAT GGA GAC TCG 222 Gly Asp Gly Asp Ser 15
GAC CAG AGC AGG AAC CGG CAA GGA ACC CCC GTG Asp Gln Ser Arg Asn Arg Gln Gly Thr Pro Val 20 25 30	Pro Ala Ser Gly Gin
GCG GCC GCC TAC AAG CAG ACG AAA GCA CAG AGG Ala Ala Ala Tyr Lys Gln Thr Lys Ala Gln Arg 40	G GCG CGG ACT ATG GCT 318 G Ala Arg Thr Met Ala 50

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TTO	TAC	AAC Ası	CCC Pro	o TT€	CCC Pro	GTC Val	CGG Arg	Gln 60	ı Asn	TGT Cys	TTC Phe	ACC Thr	GTC Val	l Ası	AGA Arg	366
TCC	CTC Leu	TTC Phe	TTE	TTC Phe	GGA Gly	GAA Glu	GAT Asp 75	Asn	: ATT	GTC Val	AGG Arg	AAA Lys 80	Туз	GCC Ala	AAG Lys	414
AAG Lys	CTC Leu 85	TTE	GAT Asp	TGG Trp	CCG Pro	CCA Pro 90	TTT Phe	GAG Glu	TAC	ATG Met	ATC Ile 95	Leu	GCC Ala	ACC Thr	ATC	462
ATT Ile 100	-ALA	AAC Asn	TGC Cys	ATC	GTC Val 105	Leu	GCC Ala	CTG Leu	GAG Glu	CAG Gln 110	His	CTT Leu	CCT	GAG Glu	GAT Asp 115	510
GAC Asp	AAG Lys	ACC Thr	Pro	Met 120	TCC Ser	CGA Arg	AGA Arg	CTG Leu	GAG Glu 125	AAG Lys	ACA Thr	GAA Glu	CCT Pro	TAT Tyr 130	TTC Phe	558
ATT Ile	GGG	ATC Ile	TTT Phe 135	Cys	TTT Phe	GAA Glu	GCT Ala	GGG Gly 140	ATC Ile	AAA Lys	ATT Ile	GTG Val	GCC Ala 145	CTG Leu	GGG Gly	606
TTC Phe	ATC Ile	TTC Phe 150	CAT His	AAG Lys	GGC Gly	TCT Ser	TAC Tyr 155	CTC Leu	CGC Arg	AAT Asn	GGC Gly	TGG Trp 160	AAT Asn	GTC Val	ATG Met	654
GAC Asp	TTC Phe 165	ATC Ile	GTG Val	GTC Val	CTC Leu	AGT Ser 170	GGC Gly	ATC Ile	CTG Leu	GCC Ala	ACT Thr 175	GCA Ala	GGA Gly	ACC Thr	CAC His	702
TTC Phe 180	AAT Asn	ACT Thr	CAC His	GTG Val	GAC Asp 185	CTG Leu	AGG Arg	ACC Thr	CTC Leu	CGG Arg 190	GCT Ala	GTG Val	CGT Arg	GTC Val	CTG Leu 195	750
CGG Arg	CCT Pro	TTG Leu	AAG Lys	CTC Leu 200	GTG Val	TCA Ser	GGG Gly	ATA Ile	CCT Pro 205	AGC Ser	CTG Leu	CAG Gln	ATT Ile	GTG Val 210	TTG Leu	798
AAG Lys	TCC Ser	ATC Ile	ATG Met 215	AAG Lys	GCC Ala	ATG Met	Val	CCT Pro 220	CTT Leu	CTG Leu	CAG Gln	ATT Ile	GGC Gly 225	CTT Leu	CTG Leu	846
CTC Leu	TTC Phe	TTT Phe 230	GCC Ala	ATC Ile	CTG Leu	Met	TTT Phe 235	GCT Ala	ATC Ile	ATT Ile	GGT Gly	TTG Leu 240	GAG Glu	TTC Phe	TAC Tyr	894
AGT Ser	GGC Gly 245	AAG Lys	TTA Leu	CAT His	CGA Arg	GCG Ala 250	TGC Cys	TTC Phe	ATG . Met .	Asn	AAT Asn 255	TCA Ser	GGT Gly	ATT Ile	CTA Leu	942
GAA Glu 260	GGA Gly	TTT Phe	GAC Asp	Pro	CCT Pro: 265	CAC His	CCA Pro	TGT Cys	Gly '	GTG Val 270	CAG Gln	GGC Gly	TGC Cys	Pro	GCT Ala 275	990

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GGT Gly	TAT Tyr	GAA Glu	TGC Cys	AAG Lys 280	GAC Asp	TGG Trp	ATC Ile	GGC Gly	CCC Pro 285	AAT Asn	GAT Asp	GGG Gly	ATC Ile	ACC Thr 290	CAG Gln	1038
TTT Phe	GAT Asp	AAC Asn	ATC Ile 295	CTT Leu	TTT Phe	GCT Ala	GTG Val	CTG Leu 300	ACT Thr	GTC Val	TTC Phe	CAG Gln	TGC Cys 305	ATC Ile	ACC Thr	1086
ATG Met	GAA Glu	GGG Gly 310	TGG Trp	ACC Thr	ACT Thr	GTG Val	CTG Leu 315	TAC Tyr	AAT Asn	ACC Thr	AAT Asn	GAT Asp 320	GCC Ala	TTA Leu	GGA Gly	1134
GCC Ala	ACC Thr 325	TGG Trp	AAT Asn	TGG Trp	CTG Leu	TAC Tyr 330	TTC Phe	ATC Ile	CCC Pro	CTC Leu	ATC Ile 335	ATC Ile	ATT Ile	GGA Gly	TCC Ser	1182
TTC Phe 340	TTT Phe	GTT Val	CTC Leu	AAC Asn	CTA Leu 345	GTC Val	CTG Leu	GGA Gly	GTG Val	CTT Leu 350	TCC Ser	GGG Gly	GAA Glu	TTT Phe	GCC Ala 355	1230
AAA Lys	GAG Glu	AGA Arg	GAG Glu	AGA Arg 360	GTG Val	GAG Glu	AAC Asn	CGA Arg	AGG Arg 365	GCT Ala	TTC Phe	ATG Met	AAG Lys	CTG Leu 370	CGG Arg	1278
CGC Arg	CAG Gln	CAG Gln	CAG Gln 375	ATT Ile	GAG Glu	CGT Arg	GAG Glu	CTG Leu 380	AAT Asn	GGC Gly	TAC Tyr	CGT Arg	GCC Ala 385	TGG Trp	ATA Ile	1326
GAC Asp	AAA Lys	GCA Ala 390	Glu	GAA Glu	GTC Val	ATG Met	CTC Leu 395	GCT Ala	GAA Glu	GAA Glu	AAT Asn	AAA Lys 400	AAT Asn	GCT Ala	GGA Gly	1374
ACA Thr	TCC Ser 405	GCC Ala	TTA Leu	GAA Glu	GTG Val	CTT Leu 410	CGA Arg	AGG Arg	GCA Ala	ACC Thr	ATC Ile 415	AAG Lys	AGG Arg	AGC Ser	CGG Arg	1422
ACA Thr 420	GAG Glu	GCC Ala	ATG Met	ACT Thr	CGA Arg 425	GAC Asp	TCC Ser	AGT Ser	GAT Asp	GAG Glu 430	CAC	TGT Cys	GTT Val	GAT Asp	ATC Ile 435	1470
TCC Ser	TCT Ser	GTG Val	GGC Gly	ACA Thr 440	Pro	CTG Leu	GCC Ala	CGA Arg	GCC Ala 445	AGT Ser	ATC Ile	AAA Lys	AGT Ser	GCA Ala 450	AAG Lys	1518
GTA Val	GAC Asp	GGG Gly	GTC Val 455	Ser	TAT Tyr	TTC Phe	CGG Arg	CAC His	ьys	GAA Glu	AGG Arg	CTT	CTG Leu 465	CGC Arg	ATC Ile	1566
TCC Ser	ATI	CGC Arg	, His	ATG Met	GTT Val	AAA Lys	TCC Ser 475	GIN	GTG Val	TTT Phe	TAC	TGG Trp 480	116	GTG Val	CTG Leu	1614
AGC Ser	CTI Lev 485	ı Val	GCA Ala	CTC Lev	AAC Asn	ACT Thr 490	Ala	TGI Cys	GTG Val	GCC Ala	ATT 11e 495	vai	CAT His	CAC His	AAC Asn	1662

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CAG		ראי	ם ידים	7 OT	7 700											
Gln 500	Pro	o Gli	n Trj) Let	1 Thi 505	. uts	Let	ı Lei	TAC Ty:	TA: Ty: 510	: Ala	A GAI a Gli	A TI u Ph	T CI e Le	G TTT u Phe 515	1710
CTG Leu	GG/ Gly	A CTO	TTO Phe	C CTC E Leu 520	r net	GAG Glu	ATO Met	TCC Ser	C CTC Lev 525	ı Lys	ATO	TA:	r GG r Gl	C AT Y Me 53	G GGG t Gly 0	1758
	****	, Det	535	. Phe	nis	ser	Ser	540	Asn	ı Cys	Phe	e Asr	9 Pho 54	e Gl 5	G GTC y Val	1806
ACA Thr	GTG Val	GG(Gl ₃ 55(T ATC	TTT Phe	GAA Glu	GTG Val 555	val	TGG	GCA Ala	ATO	TTC Phe 560	Arg	A CC	r GGT	1854
ACG Thr	TCI Ser 565		GGA Gly	ATC Ile	AGT Ser	GTC Val 570	TTG Leu	CGA Arg	GCC Ala	CTC Leu	CGG Arg 575	Leu	CT/	A AGI	A ATA	1902
580				Lys	585	11p	ATA	ser	ьeu	Arg 590	Asn	Leu	Val	. Val	TCC Ser 595	1950
				600	פעב	261	TTE	116	605	Leu	Leu	Phe	Leu	610		1998
CTC Leu	TTC Phe	ATC Ile	GTT Val 615	GTC Val	TTT Phe	GCT Ala	CTC Leu	CTA Leu 620	GGA Gly	ATG Met	CAG Gln	TTA Leu	TTT Phe 625	GGA Gly	GGC	2046
AGG Arg	TTT Phe	AAC Asn 630	TTT Phe	AAT Asn	GAT Asp	GGG Gly	ACT Thr 635	CCT Pro	TCG Ser	GCA Ala	AAT Asn	TTT Phe 640	GAT Asp	ACC Thr	TTC Phe	2094
Pro .	GCA Ala 645	GCC Ala	ATC Ile	ATG Met	ACT Thr	GTG Val 650	TTC Phe	CAG Gln	ATC Ile	CTG Leu	ACG Thr 655	GGT Gly	GAG Glu	GAC Asp	TGG Trp	2142
AAT (Asn (660	GAG Glu	GTG Val	ATG Met	TAC Tyr	AAT Asn 665	GGG Gly	ATC Ile	CGC Arg	TCC Ser	CAG Gln 670	GGT Gly	GGG Gly	GTC Val	AGC Ser	TCA Ser 675	2190
GGC :	ATG Met	TGG Trp	TCT Ser	GCC Ala 680	ATC Ile	TAC Tyr	TTC Phe	Ile	GTG Val 685	CTC Leu	ACC Thr	TTG Leu	TTT Phe	GGC Gly 690	AAC Asn	2238
TAC I	ACG Thr	Leu	CTG Leu 695	AAT Asn	GTG ' Val :	TTC Phe	Leu .	GCT . Ala 700	ATC Ile	GCT Ala	GTG Val	Asp	AAT Asn 705	CTC Leu	GCC Ala	2286
AAC (Asn A		CAG Gln 710	GAA Glu	CTG : Leu '	ACC :	Lys 1	GAT (Asp (GAA (Glu (CAG (GAG (Glu (Glu	GAA Glu 720	GAG Glu	GCC Ala	TTC Phe	2334

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	CAG Gln 725															. 23	82
	CCC Pro															24	30
	TCC Ser															24	78
	TCC Ser															25	26
	CTC Leu															25	74
	CCC Pro 805															26	22
	CTG Leu															26	70
TCC Ser	CTC Leu	AAG Lys	GGG Gly	GAT Asp 840	GGA Gly	GGG Gly	GAC Asp	CGA Arg	TCC Ser 845	AGT Ser	GCC Ala	CTG Leu	GAC Asp	AAC Asn 850	CAG Gln	27	18
AGG Arg	ACC Thr	CCT Pro	TTG Leu 855	TCC Ser	CTG Leu	GGC Gly	CAG Gln	CGG Arg 860	GAG Glu	CCA Pro	CCA Pro	TGG Trp	CTG Leu 865	GCC Ala	AGG Arg	27	66
CCC Pro	TGT Cys	CAT His 870	GGA Gly	AAC Asn	TGT Cys	GAC Asp	CCG Pro 875	ACT Thr	CAG Gln	CAG Gln	GAG Glu	GCA Ala 880	GGG Gly	GGA Gly	GGA Gly	28	14
GAG Glu	GCT Ala 885	GTG Val	GTG Val	ACC Thr	TTT Phe	GAG Glu 890	GAC Asp	CGG Arg	GCC Ala	AGG Arg	CAC His 895	AGG Arg	CAG Gln	AGC Ser	CAA Gln	28	62
CGG Arg 900	CGC Arg	AGC Ser	CGG Arg	CAT His	CGC Arg 905	CGC Arg	GTC Val	AGG Arg	ACA Thr	GAA Glu 910	GGC Gly	AAG Lys	GAG Glu	TCC Ser	TCT Ser 915	29	10
TCA Ser	GCC Ala	TCC Ser	CGG Arg	AGC Ser 920	AGG Arg	TCT Ser	GCC Ala	AGC Ser	CAG Gln 925	GAA Glu	CGC Arg	AGT Ser	CTG Leu	GAT Asp 930	Glu	29	58
GCC Ala	ATG Met	CCC Pro	ACT Thr 935	GAA Glu	GGG Gly	GAG Glu	AAG Lys	GAC Asp 940	CAT His	GAG Glu	CTC Leu	AGG Arg	GGC Gly 945	AAC Asn	CAT His	30	06

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GGT Gly	GCC Ala	AAG Lys 950	GIU	CCA Pro	ACC Thr	ATC Ile	CAA Gln 955	GIt	A GAG	AGA Arg	A GCC J Ala	C CAG Gln 960	Asp	TTA Leu	AGG Arg	3054
AGG Arg	ACC Thr 965	ASI	AGT Ser	CTG Leu	ATG Met	GTG Val 970	Ser	AGA Arg	GGC Gly	TCC Ser	GGG Gly 975		GCA Ala	GGA Gly	GGC	3102
CTT Leu 980	asp	GAG Glu	GCT Ala	GAC Asp	ACC Thr 985	Pro	CTA Leu	GTC Val	CTG Leu	CCC Pro	His	CCT Pro	GAG Glu	CTG Leu	GAA Glu 995	3150
vuı	Gry	цуѕ	nis	100	vai O	ьeu	Thr	GIu	Gln 100	Glu 5	Pro	GAA Glu	Gly	Ser 101	Ser 0	3198
GAG Glu	CAG Gln	GCC Ala	CTG Leu 101	rea	GGG	AAT Asn	GTG Val	CAG Gln 102	Leu	GAC Asp	ATG Met	GGC	CGG Arg 102	Val	ATC Ile	3246
361	GIII	103	0	PIO	Asp	Leu	Ser 1035	Cys 5	Ile	Thr	Ala	AAC Asn 1040	Thr	Asp	Lys	3294
AIG	104	5	GIU	ser	Thr	1050	Val	Thr	Val	Ala	Ile 105	-	Asp	Val	Asp	3342
1060)	val	АБР	ser	106	vaı	Val	His	Ile	Ser 107	Asn O	AAG Lys	Thr	Asp	Gly 1075	3390
GIU	AIA	ser	Pro	1080) Lys	Glu	Ala	Glu	Ile 1085	Arg	Glu	GAT Asp	Glu	Glu 1090	Glu	3438
GTG Val	GAG Glu	AAG Lys	AAG Lys 1095	Lys	CAG Gln	AAG Lys	Lys	GAG Glu 1100	Lys	CGT Arg	GAG Glu	ACA Thr	GGC Gly 1105	Lys	GCC Ala	3486
ATG Met	GTG Val	CCC Pro 1110	His	AGC Ser	TCA Ser	Met	TTC Phe 1115	Ile	TTC Phe	AGC Ser	ACC Thr	ACC Thr 1120	Asn	CCG Pro	ATC Ile	3534
Arg	AGG Arg 1125	Ala	TGC Cys	CAC His	TAC Tyr	ATC Ile 1130	GTG . Val .	AAC Asn	CTG Leu	CGC Arg	TAC Tyr 1135	TTT Phe	GAG Glu	ATG Met	TGC Cys	3582
ATC Ile 1140	Leu	CTG Leu	GTG Val	Ile .	GCA Ala 1145	GCC . Ala	AGC . Ser :	AGC Ser	Ile	GCC Ala 1150	Leu	GCG Ala	GCA Ala	Glu .	GAC Asp 1155	3630
CCC Pro	GTC Val	CTG Leu	Thr .	AAC Asn 1160	Ser	GAG (Glu)	CGC : Arg :	Asn	AAA Lys 1165	Val	CTG Leu	AGG ' Arg '	Tyr	TTT Phe 1170	GAC Asp	3678

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TAT GTG TTC A								3726
	1175		1180		1185			
GAC CAA GGC TASP Gln Gly I	Leu Ile Leu	CAG GAT Gln Asp 1195	Gly Ser	TAC TTC Tyr Phe	CGA GAC Arg Asp 1200	TTG 1		3774
AAC ATC CTG (Asn Ile Leu) 1205	Asp Phe Val	GTG GTC Val Val 1210	GTT GGC Val Gly	GCA TTG Ala Leu 1215	Val Ala	TTT (GCT Ala	3822
CTG GCG AAC C Leu Ala Asn J 1220	GCT TTG GGA Ala Leu Gly 1225	Thr Asn	AAA GGA Lys Gly	CGG GAC Arg Asp 1230	ATC AAG Ile Lys	Thr 1	ATC Ile 1235	3870
AAG TCT CTG (Lys Ser Leu)	CGG GTG CTC Arg Val Leu 1240	CGA GTT Arg Val	CTA AGG Leu Arg 1245	Pro Leu	AAA ACC Lys Thr	ATC I Ile I 1250	AAG Lys	3918
CGC TTG CCC A	AAG CTC AAG Lys Leu Lys 1255	GCC GTC Ala Val	TTC GAC Phe Asp 1260	TGC GTA Cys Val	GTG ACC Val Thr 1265	Ser 1		3966
AAG AAT GTC : Lys Asn Val 1 1270	Phe Asn Ile		Val Tyr					4014
TTT GCT GTC A Phe Ala Val : 1285	ATC GCA GTT Ile Ala Val	CAG CTC Gln Leu 1290	TTC AAG Phe Lys	GGA AAG Gly Lys 1295	Phe Phe	TAT T	TGC Cys	4062
ACG GAC AGT : Thr Asp Ser : 1300	TCC AAG GAC Ser Lys Asp 130!	Thr Glu	AAG GAG Lys Glu	TGC ATA Cys Ile 1310	GGC AAC Gly Asn	Tyr '	GTA Val 1315	4110
GAT CAC GAG ASP His Glu	AAA AAC AAG Lys Asn Lys 1320	ATG GAG Met Glu	GTG AAG Val Lys 132	Gly Arg	GAA TGG Glu Trp	AAG (Lys 2 1330	Arg	4158
CAT GAA TTC (His Glu Phe	CAC TAC GAC His Tyr Asp 1335	AAC ATT Asn Ile	ATC TGG Ile Trp 1340	GCC CTG Ala Leu	CTG ACC Leu Thr 1345	Leu :	TTC Phe	4206
ACC GTC TCC Thr Val Ser	Thr Gly Glu	GGA TGG Gly Trp 135	Pro Gln	GTT CTG Val Leu	CAG CAC Gln His 1360	TCT (GTA Val	4254
GAT GTG ACA Asp Val Thr 1365	GAG GAA GAC Glu Glu Asp	CGA GGC Arg Gly 1370	CCA AGC Pro Ser	CGC AGC Arg Ser 137	Asn Arg	ATG Met	GAG Glu	4302
ATG TCT ATC Met Ser Ile 1380	TTT TAT GTA Phe Tyr Val 138	Val Tyr	TTT GTG Phe Val	GTC TTC Val Phe 1390	CCC TTC Pro Phe	Phe	TTT Phe 1395	4350

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GTC Val	AAT Asn	ATC Ile	TTI Phe	GTG Val	ATa	CTC Leu	ATC	Ile	ATC Ile	Thr	TTC Phe	CAG	GAG Glu	CAA Glr 141	GGG Gly		4398
GAT Asp	' AAG	ATG Met	ATG Met	GIU	GAG Glu	TGC Cys	AGC Ser	CTG Leu 142	Glu	AAG Lys	AAT Asn	GAG Glu	AGG Arg 142	Ala	TGC Cys		4446
ATC	GAC Asp	Phe 143	Ara	ATC	AGC Ser	GCC Ala	AAA Lys 143	Pro	CTC Leu	ACC Thr	CGC Arg	TAC Tyr 144	Met	CCG Pro	CAG Gln		4494
AAC Asn	AGA Arg 144	HIS	ACC	TTC Phe	CAG Gln	TAC Tyr 145	Arg	GTG Val	TGG Trp	CAC His	TTT Phe 145	Val	GTG Val	TCT Ser	CCG Pro		4542
TCC Ser 146	FIIE	GAG Glu	TAC Tyr	ACC Thr	ATT Ile 146	ATG Met 5	GCC Ala	ATG Met	ATC Ile	GCC Ala 147	Leu	AAT Asn	ACT Thr	GTT Val	GTG Val 1475		4590
CTG Leu	ATG Met	ATG Met	AAG Lys	TAT Tyr 1480	туr	TCT Ser	GCT Ala	CCC Pro	TGT Cys 148	Thr	TAT Tyr	GAG Glu	CTG Leu	GCC Ala 149	Leu		4638
AAG Lys	TAC Tyr	CTG Leu	AAT Asn 149	тте	GCC Ala	TTC Phe	ACC Thr	ATG Met 150	Val	TTT Phe	TCC Ser	CTG Leu	GAA Glu 1505	Cys	GTC Val		4686
CTG Leu	AAG Lys	GTC Val 151	TTE	GCT Ala	TTT Phe	GGC Gly	TTT Phe 1515	Leu	AAC Asn	TAT Tyr	TTC Phe	CGA Arg 1520	Asp	ACC Thr	TGG Trp		4734
AAT Asn	ATC Ile 1529	Pne	GAC Asp	TTC Phe	ATC Ile	ACC Thr 1530	Val	ATT Ile	GGC Gly	AGT Ser	ATC Ile 1535	Thr	GAA Glu	ATT Ile	ATC Ile		4782
CTG Leu 1540	1111	GAC Asp	AGC Ser	AAG Lys	CTG Leu 1545	GTG Val	AAC Asn	ACC Thr	AGT Ser	GGC Gly 1550	Phe	AAT Asn	ATG Met	AGC Ser	TTT Phe 1555		4830
CTG Leu	AAG Lys	CTC Leu	TTC Phe	CGA Arg 1560	Ala	GCC Ala	CGC Arg	CTC Leu	ATA Ile 1565	Lys	CTC Leu	CTG Leu	Arg	CAG Gln 1570	Gly		4878
TAT Tyr	ACC Thr	ATA Ile	CGC Arg 1575	Ile	TTG Leu	CTG Leu	Trp	ACC Thr 1580	Phe	GTG Val	CAG Gln	Ser	TTT Phe 1585	AAG Lys	GCC Ala	4	1926
CTC Leu	Pro	TAT Tyr 1590	vai	TGC Cys	CTT Leu	TTA . Leu	ATT Ile 1595	GCC Ala	ATG Met	CTT Leu	Phe :	TTC Phe 1600	Ile	TAT Tyr	GCC Ala	4	1974
тте	ATT Ile 1605	GIA	ATG Met	CAG (Gln	Val	TTT Phe 1610	GGA . Gly .	AAC Asn	ATA Ile	Lys	TTA (Leu) 1615	GAC Asp	GAG (Glu (GAG Glu	AGT Ser	5	5022

CAC ATO His Ile 1620	AAC Asn	CGG Arg	CAC His	AAC Asn 1625	Asn	TTC Phe	CGG Arg	AGT Ser	TTC Phe 1630	Phe	GGG Gly	TCC Ser	CTA Leu	ATG Met 1635	5070
CTA CTO	TTC Phe	AGG Arg	AGT Ser 1640	Ala	ACA Thr	GGT Gly	GAG Glu	GCC Ala 1645	\mathtt{Trp}	CAG Gln	GAG Glu	ATT Ile	ATG Met 1650	Leu	5118
TCA TGG Ser Cys	CTT Leu	GGG Gly 1655	Glu	AAG Lys	GGC Gly	TGT Cys	GAG Glu 1660	Pro	GAC Asp	ACC Thr	ACC Thr	GCA Ala 1669	Pro	TCA Ser	5166
GGG CAG	AAC Asn 167	Glu	AAT Asn	GAA Glu	CGC Arg	TGC Cys 1675	Gly	ACC Thr	GAT Asp	CTG Leu	GCC Ala 1680	Tyr	GTG Val	TAC Tyr	5214
TTT GTG Phe Val	l Ser	TTC Phe	ATC Ile	TTC Phe	TTC Phe 1690	Cys	TCC Ser	TTC Phe	TTG Leu	ATG Met 1695	Leu	AAC Asn	CTG Leu	TTT Phe	5262
GTG GCC Val Ala 1700	C GTC a Val	ATC Ile	ATG Met	GAC Asp 1705	Asn	TTT Phe	GAG Glu	TAC Tyr	CTG Leu 1710	Thr	CGG Arg	GAC Asp	TCC Ser	TCC Ser 1715	5310
ATC CT	G GGG	CCT Pro	CAC His 1720	His	TTG Leu	GAC Asp	GAG Glu	TTT Phe 1725	Val	CGC Arg	GTC Val	TGG Trp	GCA Ala 1730	Glu	5358
TAT GA Tyr As	C CGA	GCA Ala 173	Ala	TGT Cys	GGC Gly	CGC Arg	ATC Ile 1740	His	TAC Tyr	ACT Thr	GAG Glu	ATG Met 174	Tyr	GAA Glu	5406
ATG CT Met Le	ACT u Thr 175	Leu	ATG Met	TCA Ser	CCT Pro	CCG Pro 175	Leu	GGC Gly	CTC Leu	GGC Gly	AAG Lys 176	Arg	TGT Cys	CCC Pro	5454
TCC AA Ser Ly 17	s Val	GCA Ala	TAT Tyr	AAG Lys	AGG Arg 177	Leu	GTC Val	CTG Leu	ATG Met	AAC Asn 177	Met	CCA Pro	GTA Val	GCT Ala	5502
GAG GA Glu As 1780	C ATG p Met	ACG Thr	GTC Val	CAC His 178	Phe	ACC Thr	TCC Ser	ACA Thr	CTT Leu 179	Met	GCT Ala	CTG Leu	ATC Ile	CGG Arg 1795	5550
ACA GC Thr Al	T CTG a Leú	GAC Asp	ATT Ile 180	Lys	ATT Ile	GCC Ala	AAA Lys	GGT Gly 180	Gly	GCA Ala	GAC Asp	AGG Arg	CAG Gln 181	GIn	5598
CTA GA Leu As	C TCA p Ser	GAG Glu 181	Leu	CAA Gln	AAG Lys	GAG Glu	ACC Thr 182	Leu	GCC Ala	ATC Ile	TGG Trp	CCT Pro 182	His	CTA Leu	5646
		101	- .												

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CT Le	G AC u Th	_ , ,	G GG l Gl	C AAI y Lys	A ATO	TATE Tyr	. AT	A GCA A Ala	A ATO	G ATO	3 ATC	e Met	GA(Asp	TAC Ty:	C TAT		5742
AA: Ly: 18:		G AG	T AAG	G GTO	AAG Lys 186	: rys	G CAC	AGG Arg	G CAC	G CAC n Glr 187	ı Let	GAG Glu	GAA Glu	CAC Gli	AAA Lys 1875		5790
			o met	188	30 30	Arg	Met	GIU	188	Ser 85	Ser	Leu	Pro	Glr 189	_		5838
		, ,,,,,,	189	5	L Lys	Ата	Leu	190	0 0	Leu	. Gln	Gln	Asp 190	Pro 5	GTT Val		5886
		191	10	GLY	AIG	ser	191	Tyr 5	Pro	Ser	Met	Ser 192	Pro 0	Leu	TCT		5934
	192	5		1116	GIII	193	0 Ala	Cys	мет	Asp	Pro 193	Ala 5	Asp	Asp	GGA Gly		5982
194	0	021	GAA Glu	AL 9	194	5 5	ren	val	Val	195	Asp 0	Pro	Ser	Ser	Met 1955		6030
5	9	Der	TTT Phe	196	0	TTE	Arg	Asp	Lys 196	Arg 5	Ser	Asn	Ser	Ser 197	Trp O	ı	6078
TTG Leu	GAG Glu	GAA Glu	TTC Phe 197	Set	ATG Met	GAG Glu	CGA Arg	AGC Ser 1980	Ser	GAA Glu	AAT Asn	ACC Thr	TAC Tyr 1985	Lys	TCC Ser		6126
CGT Arg	CGC Arg	CGG Arg 199	AGT Ser 0	TAC Tyr	CAC His	TCC Ser	TCC Ser 1995	rea	CGG Arg	CTG Leu	TCA Ser	GCC Ala 2000	His	CGC Arg	CTG Leu	•	5174
AAC Asn	TCT Ser 200	ASP	TCA Ser	GGC Gly	CAC His	AAG Lys 2010	Ser	GAC Asp	ACT Thr	CAC His	CCC Pro 2015	Ser	GGG Gly	GGC Gly	AGG Arg	6	5222
GAG Glu 2020	Arg	CGA Arg	CGA Arg	TCA Ser	AAA Lys 2025	GIU	CGA Arg	AAG Lys	CAT His	CTT Leu 2030	Leu	TCT Ser	CCT Pro	GAT Asp	GTC Val 2035	6	5270
TCC Ser	CGC Arg	TGC Cys	AAT Asn	TCA Ser 2040	GIU	GAG Glu	CGA Arg	GIA	ACC Thr 2045	Gln	GCT Ala	GAC Asp	\mathtt{Trp}	GAG Glu 2050	Ser	6	318
CCA Pro	GAG Glu	CGC Arg	CGT Arg 2055	GIN	TCC . Ser .	AGG Arg	ser	CCC . Pro : 2060	AGT Ser	GAG Glu	GGC Gly	Arg :	TCA Ser 2065	CAG Gln	ACG Thr	6	366

CCC Pro	AAC Asn	AGA Arg 2070	Gln	GGC Gly	ACA Thr	GGT Gly	TCC Ser 2075	Leu	AGT Ser	GAG Glu	AGC Ser	TCC Ser 2080	Ile	CCC Pro	TCT Ser	6414
GTC Val	TCT Ser 2085	Asp	ACC Thr	AGC Ser	ACC Thr	CCA Pro 2090	Arg	AGA Arg	AGT Ser	CGT Arg	CGG Arg 2095	Gln	CTC Leu	CCA Pro	CCC Pro	6462
GTC Val 210	CCG Pro	CCA Pro	AAG Lys	CCC Pro	CGG Arg 210	Pro	CTC Leu	CTT Leu	TCC Ser	TAC Tyr 2110	Ser	TCC Ser	CTG Leu	ATT Ile	CGA Arg 2115	6510
CAC His	GCG Ala	GGC Gly	AGC Ser	ATC Ile 2120	Ser	CCA Pro	CCT Pro	GCT Ala	GAT Asp 2125	Gly	AGC Ser	GAG Glu	GAG Glu	GGC Gly 2130	Ser	6558
CCG Pro	CTG Leu	ACC Thr	TCC Ser 2135	Gln	GCT Ala	CTG Leu	GAG Glu	AGC Ser 2140	Asn	AAT Asn	GCT Ala	TGG Trp	CTG Leu 2145	Thr	GAG Glu	6606
TCT Ser	TCC Ser	AAC Asn 2150	Ser	CCG Pro	CAC His	CCC Pro	CAG Gln 215	Gln	AGG Arg	CAA Gln	CAT His	GCC Ala 2160	Ser	CCA Pro	CAG Gln	6654
CGC Arg	TAC Tyr 216	Ile	TCC Ser	GAG Glu	CCC Pro	TAC Tyr 2170	Leu	GCC Ala	CTG Leu	CAC His	GAA Glu 217	Asp	TCC Ser	CAC His	GCC Ala	6702
TCA Ser 218	GAC Asp	TGT Cys	GTT Val	GAG Glu	GAG Glu 218	Glu	ACG Thr	CTC Leu	ACT Thr	TTC Phe 219	Glu	GCA Ala	GCC Ala	GTG Val	GCT Ala 2195	6750
ACT Thr	AGC Ser	CTG Leu	GGC Gly	CGT Arg 220	Ser	AAC Asn	ACC Thr	ATC Ile	GGC Gly 220	Ser	GCC Ala	CCA Pro	CCC Pro	CTG Leu 221	Arg	6798
CAT His	AGC Ser	TGG Trp	CAG Gln 221	Met	CCC Pro	AAC Asn	GGG Gly	CAC His 222	Tyr	CGG Arg	CGG Arg	CGG Arg	AGG Arg 222	Arg	GGG Gly	6846
GGG Gly	CCT Pro	GGG Gly 223	Pro	GGC Gly	ATG Met	ATG Met	TGT Cys 223	Gly	GCT Ala	GTC Val	AAC Asn	AAC Asn 224	Leu	CTA Leu	AGT Ser	6894
	ACG Thr 224	Glu					Cys	TAG.	AGGC	TGC	TCCC	CCCT	CC G	ATGC.	ATGCT	6948
CTT	CTCT	CAC .	ATGG.	AGAA	AA C	CAAG.	ACAG	A AT	TGGG	AAGC	CAG	TGCG	GCC	CCGC	GGGGAG	7008
GAA	GAGG	GAA .	AAGG.	AAGA	TG G	AAG										7032

- (2) INFORMATION FOR SEQ ID NO:25:
 - (i) SEQUENCE CHARACTERISTICS:
 (A) LENGTH: 7089 base pairs

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- (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (ix) FEATURE:

 - (A) NAME/KEY: CDS
 (B) LOCATION: 166..6978
 (D) OTHER INFORMATION: /standard_name= "Alpha-1E-3"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:25:

							_				J .						
GC'	TGCT	GCTG	CCT	CTCC	GAA	GAGC:	rcgc	GG A	GCTC	CCCA	G AG	GCGG'	TGGT	CCC	CGTGCI	T	60
															GGTGTI		120
						GTCTI										_	120
												ì	Met 1	Ala 1	arg		174
	3	5			- vu.	10	ALG	PIC	, GT ²	/ Ser	1:	y As <u>r</u> 5	Gly	/ Asp	TCG Ser	٠	222
GAC Asp 20	CAC Glr	AGC Ser	AGC Arg	AA(Asr	C CGC Arg 25	, GIII	GGA Gly	ACC Thr	Pro	GTG Val	Pro	G GCC	TCC Ser	GGG Gly	CAG Gln 35		270
GCG Ala	GCC Ala	GCC Ala	TAC	AAG Lys 40	, Q11.	ACG Thr	AAA Lys	GCA Ala	CAG Gln 45	Arg	GCG	G CGG	ACI Thr	ATG Met			318
TTG Leu	TAC	AAC Asn	CCC Pro 55		CCC Pro	GTC Val	CGG Arg	CAG Gln 60	AAC Asn	TGT Cys	TTC Phe	ACC Thr	GTC Val 65	AAC Asn	AGA Arg		366
TCC Ser	CTG Leu	TTC Phe 70	ATC Ile	TTC Phe	GGA Gly	GAA Glu	GAT Asp 75	AAC Asn	ATT Ile	GTC Val	AGG Arg	AAA Lys 80	TAT Tyr	GCC Ala	AAG Lys		414
AAG Lys	CTC Leu 85	ATC Ile	GAT Asp	TGG Trp	CCG Pro	CCA Pro 90	TTT Phe	GAG Glu	TAC Tyr	ATG Met	ATC Ile 95	CTG Leu	GCC Ala	ACC Thr	ATC Ile		462
ATT Ile 100	GCC Ala	AAC Asn	TGC Cys	ATC Ile	GTC Val 105	CTG Leu	GCC Ala	CTG Leu	GAG Glu	CAG Gln 110	CAT His	CTT Leu	CCT Pro	GAG Glu	GAT Asp 115		510
GAC Asp	AAG Lys	ACC Thr	CCC Pro	ATG Met 120	TCC Ser	CGA Arg	AGA Arg	CTG Leu	GAG Glu 125	AAG Lys	ACA Thr	GAA Glu	CCT Pro	TAT Tyr 130	TTC Phe		558
ATT	GGG	ATC	TTT	TGC	TTT	GAA	GCT	GGG	ATC	AAA	ATT	GTG	GCC	CTG	GGG		606

Ile	Gly	Ile	Phe 135	Cys	Phe	Glu	Ala	Gly 140	Ile	Lys	Ile	Val	Ala 145	Leu	Gly	
								CTC Leu								654
								ATC Ile								702
								ACC Thr						Val		750
								ATA Ile								798
		-						CCT Pro 220								846
								GCT Ala								894
								TTC Phe								942
GAA Glu 260	GGA Gly	TTT Phe	GAC Asp	CCC Pro	CCT Pro 265	CAC His	CCA Pro	TGT Cys	GGT Gly	GTG Val 270	CAG Gln	GGC Gly	TGC Cys	CCA Pro	GCT Ala 275	990
GGT Gly	TAT Tyr	GAA Glu	TGC Cys	AAG Lys 280	GAC Asp	TGG Trp	ATC Ile	GGC Gly	CCC Pro 285	AAT Asn	GAT Asp	GGG Gly	ATC Ile	ACC Thr 290	CAG Gln	1038
TTT Phe	GAT Asp	AAC Asn	ATC Ile 295	CTT Leu	TTT Phe	GCT Ala	GTG Val	CTG Leu 300	ACT Thr	GTC Val	TTC Phe	CAG Gln	TGC Cys 305	ATC Ile	ACC Thr	1086
ATG Met	GAA Glu	GGG Gly 310	TGG Trp	ACC Thr	ACT Thr	GTG Val	CTG Leu 315	TAC Tyr	AAT Asn	ACC Thr	AAT Asn	GAT Asp 320	GCC Ala	TTA Leu	GGA Gly	1134
GCC Ala	ACC Thr 325	TGG Trp	AAT Asn	TGG Trp	CTG Leu	TAC Tyr 330	TTC Phe	ATC Ile	CCC Pro	CTC Leu	ATC Ile 335	ATC Ile	ATT Ile	GGA Gly	TCC Ser	1182
TTC Phe 340	TTT Phe	GTT Val	CTC Leu	AAC Asn	CTA Leu 345	GTC Val	CTG Leu	GGA Gly	GTG Val	CTT Leu 350	TCC Ser	GGG Gly	GAA Glu	TTT Phe	GCC Ala 355	1230
AAA	GAG	AGA	GAG	AGA	GTG	GAG	AAC	CGA	AGG	GCT	TTC	ATG	AAG	CTG	CGG	1278

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Lys	Glu	Arg	g Glu	360	y Vai	l Glu	Asr	Arg	365	Ala	a Phe	e Met	Lys	370	ı Arg	
CGC Arg	CAG Gln	G CAC	G CAG A Glr 375	1 TT6	r GA0 ≥ Glu	G CGI 1 Arg	GAG	CTC Lev 380	ı Asr	GGC GGC	TAC Tyr	C CGI	GCC Ala 385	Tr	ATA Ile	1326
GAC Asp	AAA Lys	GCA Ala 390	a GIU	GAZ Glu	A GTO	ATG Met	Leu 395	Ala	GAA Glu	GAA Glu	AAT Asn	Lys 400	Asn	GC1 Ala	GGA Gly	1374
ACA Thr	Ser 405	MIC	TTA Leu	GAA Glu	GTG Val	CTT Leu 410	CGA Arg	AGG Arg	GCA Ala	ACC	Ile 415	Lys	AGG Arg	AGC	CGG Arg	1422
ACA Thr 420	GIU	GCC	ATG Met	ACT	CGA Arg 425	Asp	TCC Ser	AGT Ser	GAT Asp	GAG Glu 430	His	TGT Cys	GTT Val	GAT Asp	ATC Ile 435	1470
TCC Ser	TCT Ser	GTG Val	GGC	ACA Thr 440	PIO	CTG Leu	GCC Ala	CGA Arg	GCC Ala 445	AGT Ser	ATC Ile	AAA Lys	AGT Ser	GCA Ala 450	AAG Lys	1518
GTA Val	GAC Asp	GGG	GTC Val 455	TCT Ser	TAT	TTC Phe	CGG Arg	CAC His 460	AAG Lys	GAA Glu	AGG Arg	CTT Leu	CTG Leu 465	CGC Arg	ATC Ile	1566
TCC Ser	ATT Ile	CGC Arg 470	nis	ATG Met	GTT Val	AAA Lys	TCC Ser 475	CAG Gln	GTG Val	TTT Phe	TAC Tyr	TGG Trp 480	ATT Ile	GTG Val	CTG Leu	1614
AGC Ser	CTT Leu 485	GTG Val	GCA Ala	CTC Leu	AAC Asn	ACT Thr 490	GCC Ala	TGT Cys	GTG Val	GCC Ala	ATT Ile 495	GTC Val	CAT His	CAC His	AAC Asn	1662
CAG Gln 500	CCC Pro	CAG Gln	TGG Trp	CTC Leu	ACC Thr 505	CAC His	CTC Leu	CTC Leu	TAC Tyr	TAT Tyr 510	GCA Ala	GAA Glu	TTT Phe	CTG Leu	TTT Phe 515	1710
CTG Leu	GGA Gly	CTC Leu	TTC Phe	CTC Leu 520	TTG Leu	GAG Glu	ATG Met	TCC Ser	CTG Leu 525	AAG Lys	ATG Met	TAT Tyr	GGC Gly	ATG Met 530	GGG Gly	1758
CCT Pro	CGC Arg	CTT Leu	TAT Tyr 535	TTT Phe	CAC His	TCT Ser	TCA Ser	TTC Phe 540	AAC Asn	TGC Cys	TTT Phe	GAT Asp	TTT Phe 545	GGG Gly	GTC Val	1806
ACA Thr	GTG Val	GGC Gly 550	AGT Ser	ATC Ile	TTT Phe	GAA Glu	GTG Val 555	GTC Val	TGG Trp	GCA Ala	ATC Ile	TTC Phe 560	AGA Arg	CCT Pro	GGT Gly	1854
inr	TCT Ser 565	TTT Phe	GGA Gly	ATC Ile	AGT Ser	GTC Val 570	TTG Leu	CGA Arg	GCC Ala	CTC Leu	CGG Arg 575	CTT Leu	CTA Leu	AGA Arg	ATA Ile	1902
TTT	AAA	ATA	ACC	AAG	TAT	TGG	GCT	TCC	CTA	CGG .	AAT	TTG	GTG	GTC	TCC	1950

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Phe 580	Lys	Ile	Thr	Lys	Tyr 585	Trp	Ala	Ser	Leu	Arg 590	Asn	Leu	Val	Val	Ser 595		
TTG Leu	ATG Met	AGC Ser	TCA Ser	ATG Met 600	AAG Lys	TCT Ser	ATC Ile	ATC Ile	AGT Ser 605	TTG Leu	CTT Leu	TTC Phe	CTC Leu	CTC Leu 610	TTC Phe		1998
CTC Leu	TTC Phe	ATC Ile	GTT Val 615	GTC Val	TTT Phe	GCT Ala	CTC Leu	CTA Leu 620	GGA Gly	ATG Met	CAG Gln	TTA Leu	TTT Phe 625	GGA Gly	GGC Gly		2046
AGG Arg	TTT Phe	AAC Asn 630	TTT Phe	AAT Asn	GAT Asp	GGG Gly	ACT Thr 635	CCT Pro	TCG Ser	GCA Ala	AAT Asn	TTT Phe 640	GAT Asp	ACC Thr	TTC Phe		2094
CCT Pro	GCA Ala 645	GCC Ala	ATC Ile	ATG Met	ACT Thr	GTG Val 650	TTC Phe	CAG Gln	ATC Ile	CTG Leu	ACG Thr 655	GGT Gly	GAG Glu	GAC Asp	TGG Trp		2142
AAT Asn 660	GAG Glu	GTG Val	ATG Met	TAC Tyr	AAT Asn 665	GGG Gly	ATC Ile	CGC Arg	TCC Ser	CAG Gln 670	GGT Gly	GGG Gly	GTC Val	AGC Ser	TCA Ser 675		2190
GGC Gly	ATG Met	TGG Trp	TCT Ser	GCC Ala 680	ATC Ile	TAC Tyr	TTC Phe	ATT Ile	GTG Val 685	CTC Leu	ACC Thr	TTG Leu	TTT Phe	GGC Gly 690	AAC Asn		2238
TAC Tyr	ACG Thr	CTA Leu	CTG Leu 695	AAT Asn	GTG Val	TTC Phe	TTG Leu	GCT Ala 700	ATC Ile	GCT Ala	GTG Val	GAT Asp	AAT Asn 705	CTC	GCC Ala		2286
AAC Asn	GCC Ala	CAG Gln 710	GAA Glu	CTG Leu	ACC Thr	AAG Lys	GAT Asp 715	GAA Glu	CAG Gln	GAG Glu	GAA Glu	GAA Glu 720	GAG Glu	GCC Ala	TTC		2334
AAC Asn	CAG Gln 725	AAA Lys	CAT His	GCA Ala	CTG Leu	CAG Gln 730	AAG Lys	GCC Ala	AAG Lys	GAG Glu	GTC Val 735	AGC Ser	CCG Pro	ATG Met	TCT Ser		2382
GCA Ala 740	Pro	AAC Asn	ATG Met	CCT Pro	TCG Ser 745	ATC Ile	GAA Glu	AGA Arg	GAC Asp	AGA Arg 750	Arg	AGA Arg	AGA Arg	CAC His	CAC His 755		2430
ATG Met	TCG Ser	ATG Met	TGG	GAG Glu 760	Pro	CGC Arg	AGC Ser	AGC Ser	CAC His 765	Leu	AGG Arg	GAG Glu	CGG Arg	AGG Arg 770	CGC Arg		2478
CGG Arg	CAC	CAC His	ATG Met 775	Ser	GTG Val	TGG Trp	GAG Glu	CAG Gln 780	Arg	ACC Thr	AGC Ser	CAG Gln	CTG Leu 785	Arg	AAG Lys	•	2526
CAC	ATG Met	CAG Gln 790	Met	TCC	: AGC : Ser	CAG Gln	GAG Glu 795	ı Ala	CTC Leu	AAC Asn	AGA Arg	GAG Glu 800	GIU	GCG Ala	CCG Pro		2574
ACC	ATG	AAC	CCG	CTC	: AAC	CCC	CTC	: AAC	CCG	CTC	AGC	TCC	CTC	AAC	CCG		2622

Thr	Met 805	Asn	Pro	Lev	Asr	Pro 810	Leu	Asr.	Pro	Leu	Ser 815		Let	ı Asr	Pro		
CTC Leu 820	ASD	GCC Ala	CAC His	Pro	AGC Ser 825	· Leu	TAT Tyr	CGG Arg	CGA Arg	CCC Pro 830	Arg	GCC Ala	ATI Ile	GAG Glu	GGC Gly 835		2670
CTG Leu	GCC	CTG Leu	GGC	CTG Leu 840	Ala	CTG Leu	GAG Glu	AAG Lys	Phe 845	Glu	GAG Glu	GAG Glu	CGC	ATC Ile 850	AGC Ser		2718
CGT Arg	GGG	GGG Gly	TCC Ser 855	Leu	AAG Lys	GGG Gly	GAT Asp	GGA Gly 860	Gly	GAC Asp	CGA Arg	TCC Ser	AGT Ser 865	Ala	CTG Leu		2766
GAC Asp	AAC Asn	CAG Gln 870	Arg	ACC Thr	CCT Pro	TTG Leu	TCC Ser 875	CTG Leu	GGC	CAG Gln	CGG Arg	GAG Glu 880	CCA Pro	CCA Pro	TGG Trp		2814
CTG Leu	GCC Ala 885	AGG Arg	CCC Pro	TGT Cys	CAT His	GGA Gly 890	AAC Asn	TGT Cys	GAC Asp	CCG Pro	ACT Thr 895	CAG Gln	CAG Gln	GAG Glu	GCA Ala		2862
GGG Gly 900	GGA Gly	GGA Gly	GAG Glu	GCT Ala	GTG Val 905	GTG Val	ACC Thr	TTT Phe	GAG Glu	GAC Asp 910	CGG Arg	GCC Ala	AGG Arg	CAC His	AGG Arg 915		2910
CAG Gln	AGC Ser	CAA Gln	CGG Arg	CGC Arg 920	AGC Ser	CGG Arg	CAT His	CGC Arg	CGC Arg 925	GTC Val	AGG Arg	ACA Thr	GAA Glu	GGC Gly 930	AAG Lys		2958
GAG Glu	TCC Ser	TCT Ser	TCA Ser 935	GCC Ala	TCC Ser	CGG Arg	AGC Ser	AGG Arg 940	TCT Ser	GCC Ala	AGC Ser	CAG Gln	GAA Glu 945	CGC Arg	AGT Ser		3006
CTG Leu	GAT Asp	GAA Glu 950	GCC Ala	ATG Met	CCC Pro	ACT Thr	GAA Glu 955	GGG Gly	GAG Glu	AAG Lys	GAC Asp	CAT His 960	GAG Glu	CTC Leu	AGG Arg	•	3054
GGC Gly	AAC Asn 965	CAT His	GGT Gly	GCC Ala	AAG Lys	GAG Glu 970	CCA Pro	ACG Thr	ATC Ile	CAA Gln	GAA Glu 975	GAG Glu	AGA Arg	GCC Ala	CAG Gln		3102
GAT Asp 980	TTA Leu	AGG Arg	AGG Arg	ACC Thr	AAC Asn 985	AGT Ser	CTG Leu	ATG Met	GTG Val	TCC Ser 990	AGA Arg	GGC Gly	TCC Ser	GGG Gly	CTG Leu 995		3150
GCA Ala	GGA Gly	GGC Gly	CTT Leu	GAT Asp 1000	Glu	GCT Ala	GAC Asp	ACC Thr	CCC Pro 1005	Leu	GTC Val	CTG Leu	CCC Pro	CAT His 1010	Pro		3198
GAG Glu	CTG Leu	GAA Glu	GTG Val 1015	Gly	AAG Lys	CAC His	Val	GTG Val 1020	Leu	ACG Thr	GAG Glu	CAG Gln	GAG Glu 1025	Pro	GAA Glu		3246
GGC	AGC .	agt	GAG	CAG	GCC	CTG	CTG	GGG	AAT	GTG	CAG	CTA	GAC	ATG	GGC		3294

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Gly	Ser	Ser 1030		Gln	Ala	Leu	Leu 1035		Asn	Val	Gln	Leu 1040	Asp	Met	Gly	
CGG Arg	GTC Val 1045	Ile	AGC Ser	CAG Gln	AGC Ser	GAG Glu 1050	Pro	GAC Asp	CTC Leu	TCC Ser	TGC Cys 1055	ATC Ile	ACG Thr	GCC Ala	AAC Asn	3342
ACG Thr 1060	Asp	AAG Lys	GCC Ala	ACC Thr	ACC Thr 1065	Glu	AGC Ser	ACC Thr	AGC Ser	GTC Val 1070	Thr	GTC Val	GCC Ala	ATC Ile	CCC Pro 1075	3390
GAC Asp	GTG Val	GAC Asp	CCC Pro	TTG Leu 1080	Val	GAC Asp	TCA Ser	ACC Thr	GTG Val 1085	Val	CAC His	ATT Ile	AGC Ser	AAC Asn 1090	Lys	3438
ACG Thr	GAT Asp	GGG Gly	GAA Glu 1095	Ala	AGT Ser	CCC Pro	TTG Leu	AAG Lys 1100	Glu	GCA Ala	GAG Glu	ATC Ile	AGA Arg 110	GIU	GAT Asp	3486
GAG Glu	GAG Glu	GAG Glu 1110	Val	GAG Glu	AAG Lys	AAG Lys	AAG Lys 1115	Gln	AAG Lys	AAG Lys	GAG Glu	AAG Lys 1120	Arg	GAG Glu	ACA Thr	3534
GGC Gly	AAA Lys 112	Ala	ATG Met	GTG Val	CCC Pro	CAC His 1130	Ser	TCA Ser	ATG Met	TTC Phe	ATC Ile 113	TTC Phe	AGC Ser	ACC Thr	ACC Thr	3582
AAC Asn 1140	Pro	ATC Ile	CGG Arg	AGG Arg	GCC Ala 1145	Cys	CAC His	TAC Tyr	ATC Ile	GTG Val 1150	Asn	CTG Leu	CGC Arg	TAC Tyr	TTT Phe 1155	3630
GAG Glu	ATG Met	TGC Cys	ATC Ile	CTC Leu 116	Leu	GTG Val	ATT Ile	GCA Ala	GCC Ala 116	Ser	AGC Ser	ATC Ile	GCC Ala	CTG Leu 117	Ala	3678
GCA Ala	GAG Glu	GAC Asp	CCC Pro 117	Val	CTG Leu	ACC Thr	AAC Asn	TCG Ser 118	Glu	CGC Arg	AAC Asn	AAA Lys	GTC Val 118	_ ren	AGG Arg	3726
TAT	TTT Phe	GAC Asp 119	Tyr	GTG Val	TTC Phe	ACG Thr	GGC Gly 119	Val	TTC Phe	ACC Thr	TTT Phe	GAG Glu 120	met	GTT Val	ATA Ile	3774
AAG Lys	ATG Met 120	Ile	GAC Asp	CAA Gln	GGC Gly	TTG Leu 121	Ile	CTG Leu	CAG Gln	GAT Asp	GGG Gly 121	ser	TAC	TTC Phe	CGA Arg	3822
GAC Asp 122	Leu	TGG Trp	AAC Asn	ATC Ile	CTG Leu 122	Asp	TTT Phe	GTG Val	GTG Val	GTC Val 123	val	GGC	GCA Ala	TTG Leu	GTG Val 1235	3870
GCC Ala	TTT Phe	GCT Ala	CTG Leu	GCG Ala 124	Asn	GCT Ala	TTG Leu	GGA Gly	ACC Thr 124	ASD	AAA Lys	GGA Gly	CGG Arg	GAC Asp 125	ATC Ile	3918
AAG	ACC	ATC	AAG	TCI	CTG	CGG	GTG	CTC	CGA	GTT	CTA	AGG	CCF	CTG	AAA :	3966

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Lys	Th	r Il	e Ly 12	s Se: 55	r Lei	u Ar	g Va	l Le 12	u Arg	y Val	l Le	u Arg	9 Pro		u Lys	
ACC	ATC Ile	C AAG E Ly: 12	S AL	C TT(g Let	G CCC	C AAG b Lys	G CTC	u Ly:	G GC(s Ala	C GT(a Val	C TT	C GAC e Asr 128	Суя	C GT	A GTG l Val	4014
ACC	TCC Ser 128	- 26	G AA	G AAT S Asi	r GT(TTO Phe 129	Ası	T AT	A CTO	ATT	GT(Val	l Tyr	AAG Lys	CTC	TTC Phe	4062
130	0	. 116	S PIN	E ALS	130)5	Ala	a Val	l Gln	131	Phe 0	. Lys	Gly	Lys	TTC Phe 1315	4110
		Cyc	· +111	132	o ser	ser	. ràs	: Asp	132	Glu 5	Lys	Glu	Cys	Ile 133		4158
AAC Asn	TAT	GTA Val	GAT Asp 133	nis	GAG Glu	AAA Lys	AAC Asn	Lys 134	Met	GAG Glu	GTG Val	AAG Lys	GGC Gly 134	Arg	GAA Glu	4206
TGG Trp	AAG Lys	CGC Arg 135	uis	GAA Glu	TTC Phe	CAC His	TAC Tyr 135	Asp	AAC Asn	ATT Ile	ATC Ile	TGG Trp 136	Ala	CTG Leu	CTG Leu	4254
ACC Thr	CTC Leu 136		ACC	GTC Val	TCC Ser	ACA Thr 137	GIA	GAA Glu	GGA Gly	TGG Trp	CCT Pro 137	Gln	GTT Val	CTG Leu	CAG Gln	4302
CAC His 1380		GTA Val	GAT Asp	GTG Val	ACA Thr 138	GIU	GAA Glu	GAC Asp	CGA Arg	GGC Gly 1390	Pro	AGC Ser	CGC Arg	AGC Ser	AAC Asn 1395	4350
CGC Arg	ATG Met	GAG Glu	ATG Met	TCT Ser 1400	тте	TTT Phe	TAT Tyr	GTA Val	GTC Val 1405	Tyr	TTT Phe	GTG Val	GTC Val	TTC Phe 141	Pro	4398
TTC Phe	TTC Phe	TTT Phe	GTC Val 141	Asn	ATC Ile	TTT Phe	GTG Val	GCT Ala 1420	Leu	ATC Ile	ATC Ile	ATC Ile	ACC Thr 1425	Phe	CAG Gln	4446
GAG (Glu (CAA Gln	GGG Gly 1430	ASP	AAG Lys	ATG Met	ATG Met	GAG Glu 1435	Glu	TGC Cys	AGC Ser	CTG Leu	GAG Glu 1440	Lys	AAT Asn	GAG Glu	4494
-5.	GCG Ala 1445	-,5	ATC Ile	GAC Asp	PHE	GCC Ala 1450	TTE	AGC Ser	GCC Ala	Lys	CCT Pro 1455	CTC Leu	ACC Thr	CGC Arg	TAC Tyr	4542
ATG (Met 1 1460	CCG Pro	CAG Gln	AAC Asn	Arg	CAC His 1465	Inr	TTC Phe	CAG Gln	Tyr .	CGC Arg 1470	Val	TGG Trp	CAC His	TTT Phe	GTG Val 1475	4590
TG 7	CT	CCG	TCC	TTT	GAG	TAC	ACC	ATT	ATG	GCC :	ATG	ATC (GCC '	TTG	AAT	4638

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Val	Ser	Pro	Ser	Phe 1480		Tyr	Thr	Ile	Met 1485		Met	Ile	Ala	Leu 1490		
				Met					Ser		CCC Pro			Tyr		4686
			Lys					Ala			ATG Met		Phe			4734
		Val					Ala				TTG Leu 1535	Asn				4782
	Thr					Asp					ATT Ile					4830
					Asp					Asn	ACC Thr				Asn	4878
				Lys					Ala		CTC Leu			Leu		4926
			Tyr					Leu			ACC Thr		Val			4974
		Ala					Cys				GCC Ala 1615	Met				5022
ATT Ile 1620	Tyr	GCC Ala	ATC Ile	ATT Ile	GGG Gly 1625	Met	CAG Gln	GTA Val	TTT Phe	GGA Gly 1630	AAC Asn)	ATA Ile	AAA Lys	TTA Leu	GAC Asp 1635	5070
GAG Glu	GAG Glu	AGT Ser	CAC His	ATC Ile 1640	Asn	CGG Arg	CAC His	AAC Asn	AAC Asn 1645	Phe	CGG Arg	AGT Ser	TTC Phe	TTT Phe 1650	Gly	5118
TCC Ser	CTA Leu	ATG Met	CTA Leu 1655	Leu	TTC Phe	AGG Arg	AGT Ser	GCC Ala 1660	Thr	GGT Gly	GAG Glu	GCC Ala	TGG Trp 1669	Gln	GAG Glu	5166
ATT Ile	ATG Met	CTG Leu 1670	Ser	TGC Cys	CTT Leu	GGG Gly	GAG Glu 1675	Lys	GGC Gly	TGT Cys	GAG Glu	CCT Pro 1680	Asp	ACC Thr	ACC Thr	5214
GCA Ala	CCA Pro 168	Ser	GGG Gly	CAG Gln	AAC Asn	GAG Glu 1690	Asn	GAA Glu	CGC Arg	TGC Cys	GGC Gly 1695	Thr	GAT Asp	CTG Leu	GCC Ala	5262
TAC	GTG	TAC	TTT	GTC	TCC	TTC	ATC	TTC	TTC	TGC	TCC	TTC	TTG	ATG	CTC	5310

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Tyr Val Tyr 1700	Phe Val Ser	Phe Ile 1	Phe Phe (Cys Ser Phe	e Leu Met	Leu 1715
AAC CTG TTT Asn Leu Phe	GTG GCC GTC Val Ala Val 1720	ATC ATG (GAC AAC 1 Asp Asn I 1725	TTT GAG TAG Phe Glu Tyr	C CTG ACT Leu Thr 1730	Arg
GAC TCC TCC Asp Ser Ser	ATC CTG GGG Ile Leu Gly 1735	Pro His H	CAC TTG G His Leu A 1740	AC GAG TTT Asp Glu Phe	GTC CGC Val Arg 1745	GTC 5406 Val
TGG GCA GAA Trp Ala Glu 1750	Tyr Asp Arg	GCA GCA TALA ALA C	TGT GGC C	GC ATC CAT rg Ile His 176	Tyr Thr	GAG 5454 Glu
ATG TAT GAA Met Tyr Glu 1765	ATG CTG ACT Met Leu Thr	CTC ATG T Leu Met S 1770	CCA CCT C Ser Pro P	CG CTA GGC ro Leu Gly 1775	CTC GGC Leu Gly	AAG 5502 Lys
AGA TGT CCC Arg Cys Pro 1780	TCC AAA GTG Ser Lys Val 1785	Ala Tyr L	ys Arg L	TG GTC CTG eu Val Leu 790	ATG AAC Met Asn	ATG 5550 Met 1795
CCA GTA GCT Pro Val Ala	GAG GAC ATG Glu Asp Met 1800	ACG GTC C Thr Val H	AC TTC A is Phe T 1805	CC TCC ACA hr Ser Thr	CTT ATG Leu Met 1810	Ala
CTG ATC CGG	ACA GCT CTG Thr Ala Leu 1815	Asp Ile L	AA ATT G ys lle A 820	CC AAA GGT la Lys Gly	GGT GCA Gly Ala 1825	GAC 5646 Asp
AGG CAG CAG (Arg Gln Gln 1 1830	CTA GAC TCA Leu Asp Ser	GAG CTA C Glu Leu G 1835	AA AAG GA ln Lys G	AG ACC CTA lu Thr Leu 184	Ala Ile	TGG 5694 Trp
CCT CAC CTA ? Pro His Leu S 1845	ser Gin Lys	ATG CTG G Met Leu A 1850	AT CTG CT sp Leu Le	TT GTG CCC eu Val Pro 1855	ATG CCC : Met Pro :	AAA 5742 Lys
GCC TCT GAC C Ala Ser Asp I 1860	CTG ACT GTG Leu Thr Val 1865	Gly Lys I	le Tyr Al	CA GCA ATG la Ala Met 370	Met Ile 1	ATG 5790 Met 1875
GAC TAC TAT A Asp Tyr Tyr I	AAG CAG AGT ys Gln Ser 1880	AAG GTG AA Lys Val Ly	AG AAG CA ys Lys Gl 1885	AG AGG CAG .n Arg Gln	CAG CTG (Gln Leu (GAG 5838 Glu
GAA CAG AAA A Glu Gln Lys A 1	AT GCC CCC . Asn Ala Pro 1 895	Met Phe Gl	AG CGC AI ln Arg Me	G GAG CCT	TCA TCT (Ser Ser I 1905	CTG 5886 Leu
CCT CAG GAG A Pro Gln Glu I 1910	TC ATT GCT :	AAT GCC AA Asn Ala Ly 1915	AA GCC CT /s Ala Le	G CCT TAC u Pro Tyr 1920	Leu Gln G	CAG 5934 Sln
GAC CCC GTT T	CA GGC CTG	AGT GGC CG	GG AGT GG	A TAC CCT	TCG ATG A	AGT 5982

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Asp	Pro 1925		Ser	Gly	Leu	Ser 1930		Arg	Ser	Gly	Tyr 1935		Ser	Met	Ser	
CCA Pro 1940	CTC Leu)	TCT Ser	CCC Pro	CAG Gln	GAT Asp 1945	Ile	TTC Phe	CAG Gln	TTG Leu	GCT Ala 1950	Cys	ATG Met	GAC Asp	CCC Pro	GCC Ala 1955	6030
GAT Asp	GAC Asp	GGA Gly	CAG Gln	TTC Phe 1960	Gln	GAA Glu	CGG Arg	CAG Gln	TCT Ser 1965	Leu	GTG Val	GTG Val	ACA Thr	GAC Asp 1970	Pro	6078
AGC Ser	TCC Ser	ATG Met	AGA Arg 1975	Arg	TCA Ser	TTT Phe	TCC Ser	ACT Thr 1980	Ile	CGG Arg	GAT Asp	AAG Lys	CGT Arg 1985	Ser	AAT Asn	6126
TCC Ser	TCG Ser	TGG Trp 1990	Leu	GAG Glu	GAA Glu	TTC Phe	TCC Ser 1995	Met	GAG Glu	CGA Arg	AGC Ser	AGT Ser 2000	Glu	AAT Asn	ACC Thr	6174
TAC Tyr	AAG Lys 2005	Ser	CGT Arg	CGC Arg	CGG Arg	AGT Ser 2010	Tyr	CAC His	TCC Ser	TCC Ser	TTG Leu 2015	Arg	CTG Leu	TCA Ser	GCC Ala	6222
CAC His 2020	CGC Arg	CTG Leu	AAC Asn	TCT Ser	GAT Asp 2025	Ser	GGC Gly	CAC His	AAG Lys	TCT Ser 2030	Asp	ACT Thr	CAC His	CCC Pro	TCA Ser 2035	6270
GGG Gly	GGC Gly	AGG Arg	GAG Glu	CGG Arg 2040	Arg	CGA Arg	TCA Ser	AAA Lys	GAG Glu 2045	Arg	AAG Lys	CAT His	CTT Leu	CTC Leu 2050	Ser	6318
CCT Pro	GAT Asp	GTC Val	TCC Ser 2055	Arg	TGC Cys	AAT Asn	TCA Ser	GAA Glu 2060	Glu	CGA Arg	GGG Gly	ACC Thr	CAG Gln 206	Ala	GAC Asp	6366
TGG Trp	GAG Glu	TCC Ser 2070	Pro	GAG Glu	CGC Arg	CGT Arg	CAA Gln 2075	Ser	AGG Arg	TCA Ser	CCC Pro	AGT Ser 2080	Glu	GGC Gly	AGG Arg	6414
TCA Ser	CAG Gln 2085	Thr	CCC Pro	AAC Asn	AGA Arg	CAG Gln 2090	Gly	ACA Thr	GGT Gly	TCC Ser	CTA Leu 209!	Ser	GAG Glu	AGC Ser	TCC Ser	6462
ATC Ile 210	CCC Pro 0	TCT Ser	GTC Val	TCT Ser	GAC Asp 210	Thr	AGC Ser	ACC Thr	CCA Pro	AGA Arg 211	Arg	AGT Ser	CGT Arg	CGG Arg	CAG Gln 2115	6510
CTC Leu	CCA Pro	CCC Pro	GTC Val	CCG Pro 212	Pro	AAG Lys	CCC Pro	CGG Arg	CCC Pro 212	Leu	CTT Leu	TCC Ser	TAC Tyr	AGC Ser 213	Ser	6558
CTG Leu	ATT Ile	CGA Arg	CAC His 213	Ala	GGC Gly	AGC Ser	ATC Ile	TCT Ser 214	Pro	CCT Pro	GCT Ala	GAT Asp	GGA Gly 214	Ser	GAG Glu	6606
GAG	GGC	TCC	CCG	CTG	ACC	TCC	CAA	GCT	CTG	GAG	AGC	AAC	TAA	GCT	TGG	6654

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Glu	Gly	Ser 215	Pro 0	Leu	Thr	Ser	Gln 215	Ala 5	Leu	Glu	Ser	Asr. 216	Asr 0	Ala	Trp	
CTG Leu	ACC Thr 216	GAG Glu 5	TCT Ser	TCC Ser	AAC Asn	TCT Ser 217	PIO	CAC His	CCC	CAG Gln	CAG Gln 217	Arg	CAA Gln	CAT His	GCC Ala	6702
TCC Ser 2180	CCA Pro	CAG Gln	CGC Arg	TAC Tyr	ATC Ile 218	Ser	GAG Glu	CCC Pro	TAC Tyr	TTG Leu 219	Ala	CTG Leu	CAC His	GAA Glu	GAC Asp 2195	6750
TCC Ser	CAC His	GCC Ala	TCA Ser	GAC Asp 2200		GTT Val	GAG Glu	GAG Glu	GAG Glu 220	Inr	CTC Leu	ACT Thr	TTC Phe	GAA Glu 221	Ala	6798
GCC Ala	GTG Val	GCT Ala	ACT Thr 2215		CTG Leu	GGC Gly	CGT Arg	TCC Ser 2220	ASN	ACC Thr	ATC Ile	GGC Gly	TCA Ser 222	Ala	CCA Pro	6846
CCC Pro	CTG Leu	CGG Arg 2230	CAT His	AGC Ser	TGG Trp	CAG Gln	ATG Met 2235	PIO	AAC Asn	GGG Gly	CAC His	TAT Tyr 2240	Arg	CGG Arg	CGG Arg	6894
AGG Arg	CGC Arg 2245	GGG Gly	GGG Gly	CCT Pro	GGG Gly	CCA Pro 2250	GGC Gly	ATG Met	ATG Met	Cys	GGG Gly 2255	Ala	GTC Val	AAC Asn	AAC Asn	6942
CTG (Leu] 2260	CTA Leu	AGT Ser	GAC . Asp '		GAA Glu 2265	Giu	GAT Asp	GAC Asp	гуs	TGC Cys 2270		.GGCT	GC I	cccc	CCTCC	6995
SATG	CATG	CT C	TTCT	CTCA	TA C	GGAG	АААА	CCA	AGAC	AGA .	ATTG	GGAA	GC C	AGTG	CGGCC	7055
							GATG									7089
(2)]	(NFO	RMAT:	I NOI	FOR S	SEQ :	ID N	0:26	:								
	(i)	(A) (B) (C)	LEN	IGTH: PE: r RANDE	26: ucle	34 ba eic a SS: o	doubl	pair	s							
(ii)	MOLE	CULE	TYF	E: I	ANC	(genc	omic))							
(ix)	(A) (B)	TURE: NAM LOC	E/KE ATIO	N: 1	19	.0N-	/a+-		.a	·					

(D) OTHER INFORMATION: /standard_name= "Beta-2d"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:26:

ATG GTC CAA AGG GAC ATG TCC AAG TCT CCT CCC ACA CCG GCG GCG Met Val Gln Arg Asp Met Ser Lys Ser Pro Pro Thr Pro Ala Ala Ala 48

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1				5					10					15		
GTG Val	GCG Ala	CAG Gln	GAG Glu 20	ATC Ile	CAG Gln	ATG Met	GAA Glu	CTG Leu 25	CTA Leu	GAG Glu	AAC Asn	GTG Val	GCT Ala 30	CCC Pro	GCG Ala	96
					GCC Ala											144
AAA Lys	AAC Asn 50	AGA Arg	TTT Phe	AAA Lys	GGA Gly	TCT Ser 55	GAT Asp	GGA Gly	AGC Ser	ACG Thr	TCA Ser 60	TCT Ser	GAT Asp	ACT Thr	ACC Thr	192
TCA Ser 65	AAT Asn	AGT Ser	TTT Phe	GTT Val	CGC Arg 70	CAG Gln	GGT Gly	TCG Ser	GCA Ala	GAC Asp 75	TCC Ser	TAC Tyr	ACT Thr	AGC Ser	CGT Arg 80	240
CCA Pro	TCC Ser	GAT Asp	TCC Ser	GAT Asp 85	GTA Val	TCT Ser	CTG Leu	GAG Glu	GAG Glu 90	GAC Asp	CGG Arg	GAG Glu	GCA Ala	GTG Val 95	CGC Arg	288
					CAG Gln											336
AAG Lys	CCC Pro	GTT Val 115	GCA Ala	TTT Phe	GCG Ala	GTT Val	CGG Arg 120	ACA Thr	AAT Asn	GTC Val	AGC Ser	TAC Tyr 125	AGT Ser	GCG Ala	GCC Ala	384
CAT His	GAA Glu 130	GAT Asp	GAT Asp	GTT Val	CCA Pro	GTG Val 135	CCT Pro	GGC Gly	ATG Met	GCC Ala	ATC Ile 140	TCA Ser	TTC Phe	GAA Glu	GCA Ala	432
AAA Lys 145	GAT Asp	TTT Phe	CTG Leu	CAT His	GTT Val 150	AAG Lys	GAA Glu	AAA Lys	TTT Phe	AAC Asn 155	AAT Asn	GAC Asp	TGG Trp	TGG Trp	ATA Ile 160	480
GGG Gly	CGA Arg	TTG Leu	GTA Val	AAA Lys 165	GAA Glu	GGC Gly	TGT Cys	GAA Glu	ATC Ile 170	GGA Gly	TTC Phe	ATT Ile	CCA Pro	AGC Ser 175	CCA Pro	528
GTC Val	AAA Lys	CTA Leu	GAA Glu 180	AAC Asn	ATG Met	AGG Arg	CTG Leu	CAG Gln 185	CAT His	GAA Glu	CAG Gln	AGA Arg	GCC Ala 190	AAG Lys	CAA Gln	576
GGG Gly	AAA Lys	TTC Phe 195	TAC Tyr	TCC Ser	AGT Ser	AAA Lys	TCA Ser 200	GGA Gly	GGA Gly	AAT Asn	TCA Ser	TCA Ser 205	TCC Ser	AGT Ser	TTG Leu	624
GGT Gly	GAC Asp 210	ATA Ile	GTA Val	CCT Pro	AGT Ser	TCC Ser 215	AGA Arg	AAA Lys	TCA Ser	ACA Thr	CCT Pro 220	CCA Pro	TCA Ser	TCT Ser	GCT Ala	672
ATA Ile	GAC Asp	ATA Ile	GAT Asp	GCT Ala	ACT Thr	GGC Gly	TTA Leu	GAT Asp	GCA Ala	GAA Glu	GAA Glu	AAT Asn	GAT Asp	ATT Ile	CCA Pro	720

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225	5				23	0				23	5				24	0
GCA Ala	A AA	C CA n Hi	C CG s Ar	C TC g Se 24	T FI	T AA	A CC s Pr	C AG o Se	T GC r Al 25	a Asi	C AG n Se	T GT r Va	A AC 1 Th	G TC r Se 25	A CC	
		,	26	0	S AL	a we	L Pro	26:	e Pno 5	e Lys	s Ly	s Th	r Gl: 27	u Hi O	C ACT	
CCT Pro	Pro	TA' Ty: 27	- 2201	r gr	G GT. l Va	A CCI	TC(Ser 28(r mei	G CGI	A CCA	A GT	G GT0 l Va: 28!	l Lei	A GT 1 Va	G GGC l Gly	864
	290)	- -	, G.	y ly.	295	i val	Thi	. Asr	Met	300	t Glr	ı Lys	Al:	G CTG a Leu	
305				. <u>.</u>	310)	PHE	: GIU	ı GIŞ	315	Ile	e Ser	: Ile	Th	A AGG Arg 320	
				325	5 561	neu	MIG	. Lys	330	ser	Val	. Leu	Asn	Asr 335		1008
	-3-		340		116	GIU	Arg	345	Asn	Thr	Arg	Ser	Ser 350	Lev	GCG Ala	1056
	•41	355	. Sel	GIU	TIE	GIU	360	Ile	Phe	Glu	Leu	Ala 365	Arg	Thr	TTG Leu	1104
CAG Gln	TTG Leu 370	GTG Val	GTC Val	CTT Leu	GAC Asp	GCG Ala 375	GAT Asp	ACA Thr	ATT Ile	AAT Asn	CAT His 380	CCA Pro	GCT Ala	CAA Gln	CTC Leu	1152
AGT Ser 385	AAA Lys	ACC Thr	TCC Ser	TTG Leu	GCC Ala 390	CCT Pro	ATT Ile	ATA Ile	GTA Val	TAT Tyr 395	GTA Val	AAG Lys	ATT Ile	TCT Ser	TCT Ser 400	1200
CCT Pro	AAG Lys	GTT Val	TTA Leu	CAA Gln 405	AGG Arg	TTA Leu	ATA Ile	AAA Lys	TCT Ser 410	CGA Arg	GGG Gly	AAA Lys	TCT Ser	CAA Gln 415	GCT Ala	1248
AAA Lys	CAC His	CTC Leu	AAC Asn 420	GTC Val	CAG Gln	ATG Met	GTA Val	GCA Ala 425	GCT Ala	GAT Asp	AAA Lys	CTG Leu	GCT Ala 430	CAG Gln	TGT Cys	1296
CCT (-10	GAG Glu 435	CTG Leu	TTC Phe	GAT Asp	vaı	ATC Ile 440	TTG Leu	GAT Asp	GAG . Glu .	AAC Asn	CAG Gln 445	CTT Leu	GAG Glu	GAT Asp	1344
GCC :	TGT Cys	GAG Glu	CAC His	CTT Leu	GCC Ala	GAC Asp	TAT Tyr	CTG Leu	GAG Glu	GCC '	TAC Tyr	TGG Trp	AAG Lys	GCC Ala	ACC Thr	1392

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	450					455					460					•	
CAT His 465	CCT Pro	CCC Pro	AGC Ser	AGT Ser	AGC Ser 470	CTC Leu	CCC Pro	AAC Asn	CCT Pro	CTC Leu 475	CTT Leu	AGC Ser	CGT Arg	ACA Thr	TTA Leu 480		1440
GCC Ala	ACT Thr	TCA Ser	AGT Ser	CTG Leu 485	CCT Pro	CTT Leu	AGC Ser	CCC Pro	ACC Thr 490	CTA Leu	GCC Ala	TCT Ser	AAT Asn	TCA Ser 495	CAG Gln		1488
GGT Gly	TCT Ser	CAA Gln	GGT Gly 500	GAT Asp	CAG Gln	AGG Arg	ACT Thr	GAT Asp 505	CGC Arg	TCC Ser	GCT Ala	CCT Pro	ATC Ile 510	CGT Arg	TCT Ser		1536
GCT Ala	TCC Ser	CAA Gln 515	GCT Ala	GAA Glu	GAA Glu	GAA Glu	CCT Pro 520	AGT Ser	GTG Val	GAA Glu	CCA Pro	GTC Val 525	AAG Lys	AAA Lys	TCC Ser		1584
CAG Gln	CAC His 530	CGC Arg	TCT Ser	TCC Ser	TCC Ser	TCA Ser 535	GCC Ala	CCA Pro	CAC His	CAC His	AAC Asn 540	CAT His	CGC Arg	AGT Ser	GGG Gly	,	1632
ACA Thr 545	AGT Ser	CGC Arg	GGC Gly	CTC Leu	TCC Ser 550	AGG Arg	CAA Gln	GAG Glu	ACA Thr	TTT Phe 555	GAC Asp	TCG Ser	GAA Glu	ACC Thr	CAG Gln 560		1680
GAG Glu	AGT Ser	CGA Arg	GAC Asp	TCT Ser 565	GCC Ala	TAC Tyr	GTA Val	GAG Glu	CCA Pro 570	AAG Lys	GAA Glu	GAT Asp	TAT Tyr	TCC Ser 575	CAT His		1728
GAC Asp	CAC His	GTG Val	GAC Asp 580	CAC His	TAT Tyr	GCC Ala	TCA Ser	CAC His 585	CGT Arg	GAC Asp	CAC His	AAC Asn	CAC His 590	AGA Arg	GAC Asp	,	1776
GAG Glu	ACC Thr	CAC His 595	GGG Gly	AGC Ser	AGT Ser	GAC Asp	CAC His 600	AGA Arg	CAC His	AGG Arg	GAG Glu	TCC Ser 605	CGG Arg	CAC His	CGT Arg		1824
TCC Ser	CGG Arg 610	GAC Asp	GTG Val	GAT Asp	CGA Arg	GAG Glu 615	CAG Gln	GAC Asp	CAC His	AAC Asn	GAG Glu 620	TGC Cys	AAC Asn	AAG Lys	CAG Gln		1872
CGC Arg 625	Ser	CGT Arg	CAT His	AAA Lys	TCC Ser 630	AAG Lys	GAT Asp	CGC Arg	TAC Tyr	TGT Cys 635	GAA Glu	AAG Lys	GAT Asp	GGA Gly	GAA Glu 640		1920
GTG Val	ATA Ile	TCA Ser	AAA Lys	AAA Lys 645	Arg	AAT Asn	GAG Glu	GCT Ala	GGG Gly 650	Glu	TGG Trp	AAC Asn	AGG Arg	GAT Asp 655	GTT Val		1968
	ATC Ile				GTTT	TGC	CCTT	TTGT	GT T	TTTT	TTTT	T TT	TTTT	TTGA			2020
AGT	CTTG	TAT	AACT	AACA	GC A	TCCC	CAAA	A CA	AAAA	GTCT	TTG	GGGI	CTA	CACT	GCAAT	C	2080

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	•	
ATATGTGATC TGTCTTGTAA TATTTTGT	FAT TATTGCTGTT GCTTGAATAG CAATAGCATG	2140
	GT AAGTGCTACA TAAATTGGCC TGGTATGGCT	2200
	TC AAAAACTGTT TTGGGTAGCT GCCACTTGAA	2260
	'AG TGTTTTAAGA AATGTAGTTG ATGTATCCAA	2320
CAAGCCAGAA TCAGCACAGA TAAAAAGT	GG AATTTCTTGT TTCTCCAGAT TTTTAATACG	2380
	TC ATTCATGGAC CACTGTTTCT TGCTTGTACC	2440
	CA GTCTTGCCTT ACACAAAGGG GATCATAAAG	2500
	TG TGTACTGTAT AGACAGTTTG TAAATGTTAT	2560
	TA TAATATATA ATATATATCA GTTTGATCAC	2620
ACTATTTAG AGTC		2634
(2) INFORMATION FOR SEQ ID NO:2	27:	
(i) CD0777750		

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 1823 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (ix) FEATURE:

 - (A) NAME/KEY: CDS
 (B) LOCATION: 69..1631
 (D) OTHER INFORMATION: /standard_name= "Beta-4"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:27:

AGC	CCAG	CCT	CGGG	GGCC	AG C	cccc	TCCG	c cc	ACCG	CACA	CGG	GCTG	GCC	ATGC	GGCGGC	•	60
TCT	GAAC	G AT	G TC t Se 1	C TC r Se	C TC r Se	C TC r Se	C TA r Ty: 5	C GC r Al	C AA a Ly	G AA s As	C GG n Gl	y Th	C GC r Al	G GA a As	C GGG p Gly		110
CCG Pro 15	CAC His	TCC Ser	CCC Pro	ACC Thr	TCG Ser 20	CAG Gln	GTG Val	GCC Ala	CGA Arg	GGC Gly 25	ACC Thr	ACA Thr	ACC Thr	CGG Arg	AGG Arg 30		158
AGC Ser	AGG Arg	TTG Leu	AAA Lys	AGA Arg 35	TCC Ser	GAT Asp	GGC Gly	AGC Ser	ACC Thr 40	ACT Thr	TCG Ser	ACC Thr	AGC Ser	TTC Phe 45	ATC Ile		206
CTC Leu	AGA Arg	CAG Gln	GGT Gly 50	TCA Ser	GCG Ala	GAT Asp	TCC Ser	TAC Tyr 55	ACA Thr	AGC Ser	AGG Arg	CCG Pro	TCT Ser 60	GAC Asp	TCC Ser	;	254

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GAT Asp	GTC Val	TCT Ser 65	TTG Leu	GAA Glu	GAG Glu	GAC Asp	CGG Arg 70	GAA Glu	GCA Ala	ATT Ile	CGA Arg	CAG Gln 75	GAG Glu	AGA Arg	GAA Glu		302
CAG Gln	CAA Gln 80	GCA Ala	GCT Ala	ATC Ile	CAG Gln	CTT Leu 85	GAG Glu	AGA Arg	GCA Ala	AAG Lys	TCC Ser 90	AAA Lys	CCT Pro	GTA Val	GCA Ala		350
TTT Phe 95	GCC Ala	GTG Val	AAG Lys	ACA Thr	AAT Asn 100	GTG Val	AGC Ser	TAC Tyr	TGC Cys	GGC Gly 105	GCC Ala	CTG Leu	GAC Asp	GAG Glu	GAT Asp 110		398
GTG Val	CCT Pro	GTT Val	CCA Pro	AGC Ser 115	ACA Thr	GCT Ala	ATC Ile	TCC Ser	TTT Phe 120	GAT Asp	GCT Ala	AAA Lys	GAC Asp	TTT Phe 125	CTA Leu		446
CAT His	ATT Ile	AAA Lys	GAG Glu 130	AAA Lys	TAT Tyr	AAC Asn	AAT Asn	GAT Asp 135	TGG Trp	TGG Trp	ATA Ile	GGA Gly	AGG Arg 140	CTG Leu	GTG Val		494
AAA Lys	GAG Glu	GGC Gly 145	TGT Cys	GAA Glu	ATT Ile	GGC Gly	TTC Phe 150	ATT Ile	CCA Pro	AGT Ser	CCA Pro	CTC Leu 155	AGA Arg	TTG Leu	GAG Glu		542
AAC Asn	ATA Ile 160	Arg	ATC Ile	CAG Gln	CAA Gln	GAA Glu 165	CAA Gln	AAA Lys	AGA Arg	GGA Gly	CGT Arg 170	FIIC	CAC His	GGA Gly	GGG Gly		590
AAA Lys 175	Ser	AGT Ser	GGA Gly	AAT Asn	TCT Ser 180	Ser	TCA Ser	AGT Ser	CTT Leu	GGA Gly 185	GAA Glu	ATG Met	GTA Val	TCT Ser	GGG Gly 190		638
Thr	Phe	Arg	Ala	Thr 195	Pro	Inr	Ser	THE	200	БуS	G11.			205			686
ACG Thr	GAG Glu	CAC His	ATT Ile 210	Pro	CCT Pro	TAC Tyr	GAT Asp	GTT Val 215	val	CCG Pro	TCA Ser	ATG Met	CGT Arg 220		GTG Val		734
Va]	. Lev	225	Gly	Pro	ser	Leu	230)	/ IYL	. GIU	· vu	235	5		ATG Met		782
CAC Gl:	AAA 1 Lys 240	s Ala	CTC Lev	TTI Phe	GAT Asp	TCC Ser 245	Tel	AAC 1 Lys	CAC His	AGG Arg	TT: Phe 25		GGG Gly	AGG Arg	ATT J Ile		830
Se: 25:	r Ile 5	e Thi	. Arg	y val	260) L HIS	ı Ası	b 110	- 50.	265	5	,		-	r GTC r Val 270		878
CT. Le	A AA' u As:	T AA' n Asi	r cco	275 275	г гу:	G AG	A GC	A AT	A AT' e Ile 28		A CG u Ar	T TC	G AA(r Asi	n Th	C CGG r Arg 5	1	926

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	C Ac		2	90					29	5	re (∍1U	Arg	3 II	.е Р З	he 00	Glu	Leu		974
GC Al	A AG a Ar	A TO G Se 30	T T	rg cz eu g:	AA C	TG G eu V		TT Val	CT:	r ga ı As	AT G	CA la	GA(Asr	C AC Th	rI	TC le	AAT Asn	CAC His		1022
CC. Pr	A GC O Al. 32	A CA a Gl 0	A CI	TT AT	TA Ai le Ly		CT I hr S 25	CC er	TTA	A GC	A C	CA ro	ATI Ile 330	II	T G' e Va	TT (CAT His	GTA Val		1070
AA: Ly: 33:	A GTO S Val	C TC l Se	A TO	T CC	A A2 O L ₃		TT T	TA eu	CAG Gln	CG	a n	TG eu 45	ATT Ile	AA: Ly:	A TO	er 1	AGA Arg	GGA Gly 350		1118
AAC Lys	TCI S Sei	A CA	A AG n Se	T AA r Ly 35	A CA s Hi 5	C T	rg A eu A	AT (GTT Val	CA Gli 36	II TIE	rg (eu '	GTG Val	GCA Ala	A GC	a A	AT Asp			1166
CTI Leu	GCA Ala	CA Gli	A TG a Cy 37	C CC s Pr	C CC o Pr	A GA O Gl	A A. u Me	:	TTT Phe 375	GAT Asp	r Gr	rr 2 al 1	ATA Ile	TTG Leu	GA As 38	рG	AA lu	AAT Asn		1214
CAG Gln	CTT Leu	GAC Glu 385	GA! Asp	r GC	A TG	r ga s gl	A CA u Hi 39		CTA Leu	GGG Gly	G GA Gl	G 7	rac ryr	CTG Leu 395	GA Gl	G G u A	CG ' la '	TAC Tyr		1262
	CGT Arg 400					40	5		CI.	1111	PI	0 M	10	Thr	Pro	o Le	eu 1	Leu		1310
GGA Gly 415	AGG Arg	AAT Asn	Leu	GGC Gly	TCC Ser 420		G GC r Al	A C a L	TC eu	TCA Ser	CC Pr 42	0 1	AT 'yr	CCC Pro	ACZ Thi	A GO	la 1	TT le		1358
TCT Ser	GGG Gly	TTA Leu	CAG Gln	AGT Ser 435	CAG Gln	CGA	A AT	G A	ъ9 .	CAC His 440	AG(C A	AC (CAC His	TCC	Th:	ır G	AG lu		1406
AAC Asn	TCT Ser	CCA Pro	ATT Ile 450	GAA Glu	AGA Arg	CGA Arg	AG:	r C:	au i	ATG Met	ACC Thr	TO Se	CT (er /	GAT Asp	GAA Glu 460	AA As	T T n T	AT yr		1454
CAC His	AAT Asn	GAA Glu 465	AGG Arg	GCT Ala	CGG Arg	AAG Lys	AGT Ser 470		ig 1	AAC Asn	CGC Arg	TI Le	eu S	CT Ser	TCC Ser	AG Se	T To	CT er	;	1502
CAG Gln	CAT . His 480	AGC Ser	CGA Arg	GAT Asp	CAT His	TAC Tyr 485	CCI	CI Le	T G	TG Val	GAA Glu	GA Gl 49	u A	AT '	TAC Tyr	CC'	T GA	AC Sp	:	1550
TCA Ser 1	TAC (CAG Gln	GAC Asp	ACT Thr	TAC Tyr 500	AAA Lys	CCC	CA Hi	TA sA	æg.	AAC Asn 505	CG Ar	A G	GA ?	TCA Ser	CC: Pro	G G G S 1	У	· 3	1598

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GGA Gly	TAT Tyr	AGC Ser	His	GAC Asp 515	TCC Ser	CGA Arg	CAT His	AGG Arg	CTT Leu 520	TGAG	TCTA	AT G	AAAC	AAAA	ı A	1648
ATAT	TCAT	CT G	TTGA	CAAT	T TG	CCAT	AGCA	GTG	CTAG	GAT	AAAC	CAAT	CA T	CTTA	ACTTG	1708
GCTA	ACAT	AG C	ACAG	TATT	T AC	TGTG	CTAA	TGG	GCTG	CTG	TCAT	TTTA	TG C	TAAG	TAAGG	1768
GGCA	AAAA	AA A	TAAA	TACA	T TA	TGCC	CTTG	AGI	CTAG	ATG	GATA	TTAG	AT G	CCCG	;	1823
(2)	INFO	RMAT	NOI	FOR	SEQ	ID N	10:28	:								
	(i) S	(B)	LEN TYP	IGTH: E: a	520 minc	RIST ami aci inea	.no a .d		ŀ						
	(i	i) M	OLEC	ULE	TYPE	: pr	otei	.n								
	(ж	i) S	EQUE	NCE	DESC	RIPI	: NOI	SEC) ID	NO: 2	8:					
Met 1	Ser	Ser	Ser	Ser , 5	Tyr	Ala	Lys	Asn	Gly 10	Thr	Ala	Asp	Gly	Pro 15	His	
Ser	Pro	Thr	Ser 20	Gln	Val	Ala	Arg	Gly 25	Thr	Thr	Thr	Arg	Arg 30	Ser	Arg	
Leu	Lys	Arg 35	Ser	Asp	Gly	Ser	Thr 40	Thr	Ser	Thr	Ser	Phe 45	Ile	Leu	Arg	
Gln	Gly 50	Ser	Ala	Asp	Ser	Tyr 55	Thr	Ser	Arg	Pro	Ser 60	Asp	Ser	Asp	Val	
Ser 65	Leu	Glu	Glu	Asp	Arg 70	Glu	Ala	Ile	Arg	Gln 75	Glu	Arg	Glu	Gln	Gln 80	
Ala	Ala	Ile	Gln	Leu 85	Glu	Arg	Ala	Lys	Ser 90	Lys	Pro	Val	Ala	Phe 95	Ala	
Val	Lys	Thr	Asn 100	Val	Ser	Tyr	Cys	Gly 105	Ala	Leu	Asp	Glu	Asp 110	Val	Pro	
Val	Pro	Ser 115	Thr	Ala	Ile	Ser	Phe 120	Asp	Ala	Lys	Asp	Phe 125	Leu	His	Ile	
Lys	Glu 130	Lys	Tyr	Asn	Asn	Asp 135	Trp	Trp	Ile	Gly	Arg 140	Leu	Val	Lys	Glu	
Gly 145		Glu	Ile	Gly	Phe 150	Ile	Pro	Ser	Pro	Leu 155	Arg	Leu	Glu	Asn	Ile 160	
Arg	Ile	Gln	Gln	Glu 165	Gln	Lys	Arg	Gly	Arg 170	Phe	His	Gly	Gly	Lys 175	Ser	
Ser	Gly	Asn	Ser 180		Ser	Ser	Leu	Gly 185	Glu	Met	Val	Ser	Gly 190	Thr	Phe	

			_				20	U				20	15		ır Glu
Hi	s Il 21	e Pr O	0 P1	со Ту	r As	p Va 21	1 Va 5	l Pr	O Se	r Me	t Ar 22	g Pr 0	o Va	ıl Va	l Leu
Va 22	1 G1 5	y Pr	O Se	r Le	u Ly:	s Gl	у Ту	r Gl	u Va	l Th	r As	p Me	t Me	t Gl	n Lys 240
Ala	a Le	u Ph	e As	p Se 24	r Lei 5	ı Lys	s Hi	s Ar	g Ph 25	e As O	p Gl	y Ar	g Il	e Se. 25.	r Ile
Th:	r Ar	g Va	1 Th 26	r Ala	a Asp	o Ile	e Se:	r Le	u Al 5	a Ly	s Arg	g Se	r Va 27	l Le	ı Asn
Ası	n Pro	27	r Ly 5	s Arg	J Ala	Ile	280	e Gla	u Ar	g Se	r Ası	Th:	r Arg	g Sei	Ser
						2,00	•				300)			Arg
Ser 305	Leu	ı Glı	n Le	u Val	. Val 310	Leu	Asp	Ala	a Ası	Th:	r Ile	Ası	His	e Pro	Ala 320
				325					330	,				335	
								345	•				350	ı	
			-	Leu			360					365			
				Glu		3/5					380				
				Glu	220					395					400
Ala	Thr	His	Thr	Thr 405	Ser	Ser	Thr	Pro	Met 410	Thr	Pro	Leu	Leu	Gly 415	Arg
Asn	Leu	Gly	Ser 420	Thr	Ala	Leu	Ser	Pro 425	Tyr	Pro	Thr	Ala	Ile 430	Ser	Gly
Leu	Gln	Ser 435	Gln	Arg	Met	Arg	His 440	Ser	Asn	His	Ser	Thr 445	Glu	Asn	Ser
Pro	Ile 450	Glu	Arg	Arg	Ser	Leu 455	Met	Thr	Ser	Asp	Glu 460	Asn	Tyr	His	Asn
Glu 465	Arg	Ala	Arg	Lys	Ser 470	Arg	Asn	Arg	Leu	Ser 475	Ser	Ser	Ser	Gln	His 480
Ser	Arg	Asp	His	Tyr 485	Pro	Leu '	Val	Glu	Glu 490	Asp	Tyr	Pro	Asp	Ser 495	Tyr

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Gln	Asp	Thr	Tyr 500	Lys	Pro	His	Arg	Asn 505	Arg	Gly	Ser	Pro	Gly 510	Gly	Tyr	
Ser	His	Asp 515	Ser	Arg	His	Arg	Leu 520									•
(2)	INFO	RMAI	MOI	FOR	SEQ	ID N	10:29	€:								
	(i)	(E	OUENC (A) LE (B) TY (C) ST (C) TC	NGTH PE: RAND	: 36 nucl EDNE	36 b eic SS:	ase acid doub	pair 1	s							
	(ii)	MOI	ECUI	E TY	PE:	DNA	(ger	nomic	:)							
	(ix)	(F	A) NA	ME/F	ON:	35	3346 TION	5 : /st	anda	.rd_n	ame=	"Al	pha-	2a"		
	(ix)	FE# (#	ATURE A) NA B) LO	ME/F	ŒY:	5'UI	rr 34									
	(ix)) FE2 (2 (1	ATURI A) NA B) LO	ME/I	CEY:	3'UI 3347	rr 73	636								,
	(xi)) SE(QUENC	CE DI	ESCRI	(PTIC	ON:	SEQ I	D NO	:29:	:					
GCG	GGGG;	AGG (GGC?	attg/	AT CT	rtcg <i>i</i>	ATCG:	C GAZ	AG AT	rg go et Al	CT GC la Al	T GC .a Gl	C TO	SC CI 's Le 5	rG eu	52
CTG Leu	GCC Ala	TTG Leu	ACT Thr 10	CTG Leu	ACA Thr	CTT Leu	TTC Phe	CAA Gln 15	TCT Ser	TTG Leu	CTC Leu	ATC Ile	GGC Gly 20	CCC Pro	TCG Ser	10
TCG Ser	GAG Glu	GAG Glu 25	Pro	TTC Phe	CCT Pro	TCG Ser	GCC Ala 30	GTC Val	ACT Thr	ATC Ile	AAA Lys	TCA Ser 35	TGG Trp	GTG Val	GAT Asp	14
AAG Lys	ATG Met 40	Gln	GAA Glu	GAC Asp	CTT Leu	GTC Val 45	ACA Thr	CTG Leu	GCA Ala	AAA Lys	ACA Thr 50	GCA Ala	AGT Ser	GGA Gly	GTC Val	19
AAT Asn 55	Gln	CTT Leu	GTT Val	GAT Asp	ATT Ile 60	Tyr	GAG Glu	AAA Lys	TAT Tyr	CAA Gln 65	GAT Asp	TTG Leu	TAT Tyr	ACT Thr	GTG Val 70	24
GAA Glu	CCA Pro	AAT Asn	AAT Asn	GCA Ala 75	Arg	CAG Gln	CTG Lev	GTA Val	GAA Glu 80	TTE	GCA Ala	GCC Ala	AGG Arg	GAT Asp 85		29

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GA(AA/ Lys	CT:	r CTC Let 90	7 261	AAC Asn	AGA Arg	TCI Ser	AAA Lys 95	: Ala	CTC Leu	G GT(G AGO L Ser	CTC Lev	ı Ala	A TTG a Leu	340
GA/ Glu	A GCG	GAC Glu	тъ	A GTT Val	CAA Gln	GCA Ala	GCT Ala 110	His	CAG Gln	TGG Trp	AGA Arg	A GAA J Glu 115	Asp	TT:	r GCA e Ala	386
AGC	AAT Asn 120	GIL	A GTT 1 Val	GTC Val	TAC	TAC Tyr 125	Asn	GCA Ala	AAG Lys	GAT Asp	GAT Asp 130	Leu	GAT Asp	CCT Pro	GAG Glu	436
135	ASI	ASL) Sei	GIU	140	GIY	Ser	Gln	Arg	11e 145	Lys	Pro	Val	Phe	ATT Ile 150	484
	тор	AT G	ASI	155	GIY	Arg	Gin	IIe	Ser 160	Tyr	Gln	His	Ala	Ala 165		532
		110	170	Asp	TIE	Tyr	GIU	175	Ser	Thr	Ile	Val	Leu 180	Asn	GAA Glu	580
CTC Leu	AAC Asn	TGG Trp 185	TILL	AGT Ser	GCC Ala	TTA Leu	GAT Asp 190	GAA Glu	GTT Val	TTC Phe	AAA Lys	AAG Lys 195	AAT Asn	CGC Arg	GAG Glu	628
GAA Glu	GAC Asp 200	CCT Pro	TCA Ser	TTA Leu	TTG Leu	TGG Trp 205	CAG Gln	GTT Val	TTT Phe	GGC Gly	AGT Ser 210	GCC Ala	ACT Thr	GGC Gly	CTA Leu	676
GCT Ala 215	CGA Arg	TAT Tyr	TAT Tyr	CCA Pro	GCT Ala 220	TCA Ser	CCA Pro	TGG Trp	GTT Val	GAT Asp 225	AAT Asn	AGT Ser	AGA Arg	ACT Thr	CCA Pro 230	724
TAA Asn	AAG Lys	ATT Ile	GAC Asp	CTT Leu 235	TAT Tyr	GAT Asp	GTA Val	CGC Arg	AGA Arg 240	AGA Arg	CCA Pro	TGG Trp	TAC Tyr	ATC Ile 245	CAA Gln	772
GGA Gly	GCT Ala	GCA Ala	TCT Ser 250	CCT Pro	AAA Lys	GAC Asp	Met	CTT Leu 255	ATT Ile	CTG Leu	GTG Val	GAT Asp	GTG Val 260	AGT Ser	GGA Gly	820
AGT Ser	GTT Val	AGT Ser 265	GGA Gly	TTG Leu	ACA Thr	Leu :	AAA Lys 270	CTG Leu	ATC Ile	CGA Arg	ACA Thr	TCT Ser 275	GTC Val	TCC Ser	GAA Glu	868
iec	TTA Leu 280	GAA Glu	ACC Thr	CTC Leu	ser .	GAT (Asp 2 285	GAT (Asp .	GAT Asp	TTC Phe	Val	AAT Asn 290	GTA Val	GCT Ala	TCA Ser	TTT Phe	916
AC sn 95	AGC Ser	AAT Asn	GCT Ala	GIn .	GAT Asp 300	GTA A	AGC ' Ser (TGT Cys	Phe	CAG Gln : 305	CAC His	CTT Leu	GTC Val	CAA Gln	GCA Ala 310	964

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AAT Asn	GTA Val	AGA Arg	AAT Asn	AAA Lys 315	AAA Lys	GTG Val	TTG Leu	AAA Lys	GAC Asp 320	GCG Ala	GTG Val	AAT Asn	AAT Asn	ATC Ile 325	ACA Thr	1012
GCC Ala	AAA Lys	GGA Gly	ATT Ile 330	ACA Thr	GAT Asp	TAT Tyr	AAG Lys	AAG Lys 335	GGC Gly	TTT Phe	AGT Ser	TTT Phe	GCT Ala 340	TTT Phe	GAA Glu	1060
CAG Gln	CTG Leu	CTT Leu 345	AAT Asn	TAT Tyr	AAT Asn	GTT Val	TCC Ser 350	AGA Arg	GCA Ala	AAC Asn	TGC Cys	AAT Asn 355	AAG Lys	ATT Ile	ATT Ile	1108
ATG Met	CTA Leu 360	TTC Phe	ACG Thr	GAT Asp	GGA Gly	GGA Gly 365	GAA Glu	GAG Glu	AGA Arg	GCC Ala	CAG Gln 370	GAG Glu	ATA Ile	TTT Phe	AAC Asn	1156
AAA Lys 375	TAC Tyr	AAT Asn	AAA Lys	GAT Asp	AAA Lys 380	AAA Lys	GTA Val	CGT Arg	GTA Val	TTC Phe 385	AGG Arg	TTT Phe	TCA Ser	GTT Val	GGT Gly 390	1204
CAA Gln	CAC His	AAT Asn	TAT Tyr	GAG Glu 395	AGA Arg	GGA Gly	CCT Pro	ATT Ile	CAG Gln 400	TGG Trp	ATG Met	GCC Ala	TGT Cys	GAA Glu 405	AAC Asn	1252
AAA Lys	GGT Gly	TAT Tyr	TAT Tyr 410	TAT Tyr	GAA Glu	ATT Ile	CCT Pro	TCC Ser 415	ATT Ile	GGT Gly	GCA Ala	ATA Ile	AGA Arg 420	ATC Ile	AAT Asn	1300
ACT Thr	CAG Gln	GAA Glu 425	Tyr	TTG Leu	GAT Asp	GTT Val	TTG Leu 430	GGA Gly	AGA Arg	CCA Pro	ATG Met	GTT Val 435	TTA Leu	GCA Ala	GGA Gly	1348
GAC Asp	AAA Lys 440	Ala	AAG Lys	CAA Gln	GTC Val	CAA Gln 445	TGG Trp	ACA Thr	AAT Asn	GTG Val	TAC Tyr 450	CTG Leu	GAT Asp	GCA Ala	TTG Leu	1396
GAA Glu 455	Leu	GGA Gly	CTT Leu	GTC Val	ATT Ile 460	ACT Thr	GGA Gly	ACT	CTT Leu	CCG Pro 465	GTC Val	TTC Phe	AAC Asn	ATA Ile	ACC Thr 470	1444
GGC Gly	CAA Gln	TTT Phe	GAA Glu	AAT Asn 475	AAG Lys	ACA Thr	AAC Asn	TTA Leu	AAG Lys 480	Asn	CAG Gln	CTG Leu	ATT	CTT Leu 485	GGT Gly	1492
GTG Val	ATG Met	GGA Gly	GTA Val 490	Asp	GTG Val	TCT Ser	TTG	GAA Glu 495	Asp	ATT	AAA Lys	AGA Arg	Leu 500		Pro	1540
CGT Arg	TTI Phe	ACA Thr	Leu	TGC Cys	CCC Pro	AAT Asn	GGG Gly 510	Tyr	TAC	TTT Phe	GCA Ala	ATC Ile 515	ASP	CCT Pro	TAA '	1588
GGI Gly	TA1	· Val	TTA Lev	TTA Lev	A CAT	CCA Pro	Asr	CTI Lev	CAG Gln	CCA Pro	AAG Lys 530	PIC	ATI Ile	GGI Gly	GTA Val	1636

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GGT Gly 535	Ile	CCA Pro	ACA Thr	ATT Ile	AAT Asn 540	TTA Leu	AGA Arg	AAA Lys	AGG Arg	AGA Arg 545	CCC	AAT Asn	ATC Ile	CAG Gln	AAC Asn 550	1684
CCC	AAA Lys	TCT	CAG Gln	GAG Glu 555	CCA Pro	GTA Val	ACA Thr	TTG Leu	GAT Asp 560	TTC Phe	CTT Leu	GAT Asp	GCA Ala	GAG Glu 565	TTA Leu	1732
GAG Glu	TAA Taa	GAT Asp	ATT Ile 570	AAA Lys	GTG Val	GAG Glu	ATT Ile	CGA Arg 575	AAT Asn	AAG Lys	ATG Met	ATT Ile	GAT Asp 580	GGG Gly	GAA Glu	1780
AGT Ser	GGA Gly	GAA Glu 585	AAA Lys	ACA Thr	TTC Phe	AGA Arg	ACT Thr 590	CTG Leu	GTT Val	AAA Lys	TCT	CAA Gln 595	GAT Asp	GAG Glu	AGA Arg	1828
Tyr	Ile 600	Asp	Lys	Gly	Asn	Arg 605	Thr	Tyr	Thr	Trp	Thr 610	CCT Pro	Val	Asn	Gly	1876
ACA Thr 615	GAT Asp	TAC Tyr	AGT Ser	TTG Leu	GCC Ala 620	TTG Leu	GTA Val	TTA Leu	CCA Pro	ACC Thr 625	TAC Tyr	AGT Ser	TTT Phe	TAC Tyr	TAT Tyr 630	1924
Ile	Lys	Ala	Lys	Leu 635	Glu	Glu	Thr	Ile	Thr 640	Gln	Ala	AGA Arg	Tyr	Ser 645	Glu	1972
Thr	Leu	Lys	Pro 650	Asp	Asn	Phe	Glu	Glu 655	Ser	Gly	Tyr	ACA Thr	Phe 660	Ile	Ala	2020
CCA Pro	AGA Arg	GAT Asp 665	TAC Tyr	TGC Cys	AAT Asn	GAC Asp	CTG Leu 670	AAA Lys	ATA Ile	TCG Ser	GAT Asp	AAT Asn 675	AAC Asn	ACT Thr	GAA Glu	2068
TTT Phe	CTT Leu 680	TTA Leu	AAT Asn	TTC Phe	AAC Asn	GAG Glu 685	TTT Phe	ATT Ile	GAT Asp	AGA Arg	AAA Lys 690	ACT Thr	CCA Pro	AAC Asn	AAC Asn	2116
CCA Pro 695	TCA Ser	TGT Cys	AAC Asn	GCG Ala	GAT Asp 700	TTG Leu	ATT Ile	AAT Asn	AGA Arg	GTC Val 705	TTG Leu	CTT Leu	GAT Asp	GCA Ala	GGC Gly 710	2164
TTT Phe	ACA Thr	AAT Asn	GAA Glu	CTT Leu 715	GTC Val	CAA Gln	AAT Asn	TAC Tyr	TGG Trp 720	AGT Ser	AAG Lys	CAG Gln	AAA Lys	AAT Asn 725	ATC Ile	2212
												GGG Gly				2260
												AAC Asn 755				2308

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TAT Tyr	GAG Glu 760	GAC Asp	AGC Ser	TTC Phe	TAT	AAA Lys 765	AGG Arg	AGC Ser	CTA Leu	GAT Asp	AAT Asn 770	GAT Asp	AAC Asn	TAT Tyr	GTT Val	2356
TTC Phe 775	ACT Thr	GCT Ala	CCC Pro	TAC Tyr	TTT Phe 780	AAC Asn	AAA Lys	AGT Ser	GGA Gly	CCT Pro 785	GGT Gly	GCC Ala	TAT Tyr	GAA Glu	TCG Ser 790	2404
GGC Gly	ATT Ile	ATG Met	GTA Val	AGC Ser 795	AAA Lys	GCT Ala	GTA Val	GAA Glu	ATA Ile 800	TAT Tyr	ATT Ile	CAA Gln	GGG Gly	AAA Lys 805	CTT Leu	2452
CTT Leu	AAA Lys	CCT Pro	GCA Ala 810	GTT Val	GTT Val	GGA Gly	ATT Ile	AAA Lys 815	ATT Ile	GAT Asp	GTA Val	AAT Asn	TCC Ser 820	TGG Trp	ATA Ile	2500
GAG Glu	AAT Asn	TTC Phe 825	ACC Thr	AAA Lys	ACC Thr	TCA Ser	ATC Ile 830	AGA Arg	GAT Asp	CCG Pro	TGT Cys	GCT Ala 835	GGT Gly	CCA Pro	GTT Val	2548
TGT Cys	GAC Asp 840	TGC Cys	AAA Lys	AGA Arg	AAC Asn	AGT Ser 845	GAC Asp	GTA Val	ATG Met	GAT Asp	TGT Cys 850	GTG Val	ATT Ile	CTG Leu	GAT Asp	2596
GAT Asp .855	GGT Gly	GGG Gly	TTT Phe	CTT Leu	CTG Leu 860	ATG Met	GCA Ala	AAT Asn	CAT His	GAT Asp 865	GAT Asp	TAT Tyr	ACT Thr	AAT Asn	CAG Gln 870	2644
ATT Ile	GGA Gly	AGA Arg	TTT Phe	TTT Phe 875	GGA Gly	GAG Glu	ATT Ile	GAT Asp	CCC Pro 880	AGC Ser	TTG Leu	ATG Met	AGA Arg	CAC His 885	CTG Leu	2692
GTT Val	AAT Asn	ATA Ile	TCA Ser 890	GTT Val	TAT Tyr	GCT Ala	TTT Phe	AAC Asn 895	AAA Lys	TCT Ser	TAT Tyr	GAT Asp	TAT Tyr 900	CAG Gln	TCA Ser	2740
GTA Val	TGT Cys	GAG Glu 905	CCC Pro	GGT Gly	GCT Ala	GCA Ala	CCA Pro 910	AAA Lys	CAA Gln	GGA Gly	GCA Ala	GGA Gly 915	CAT His	CGC Arg	TCA Ser	2788
GCA Ala	TAT Tyr 920	Val	CCA Pro	TCA Ser	GTA Val	GCA Ala 925	GAC Asp	ATA Ile	TTA Leu	CAA Gln	ATT Ile 930	GGC Gly	TGG Trp	TGG Trp	GCC Ala	2836
ACT Thr 935	Ala	GCT Ala	GCC Ala	TGG Trp	TCT Ser 940	ATT Ile	CTA Leu	CAG Gln	CAG Gln	TTT Phe 945	CTC Leu	TTG Leu	AGT Ser	TTG Leu	ACC Thr 950	2884
TTT Phe	CCA Pro	CGA Arg	CTC Leu	CTT Leu 955	GAG Glu	GCA Ala	GTT Val	GAG Glu	ATG Met 960	Glu	GAT Asp	GAT Asp	GAC Asp	TTC Phe 965	ACG Thr	2932
GCC Ala	TCC	CTG Leu	TCC Ser 970	Lys	CAG Gln	AGC Ser	TGC Cys	ATT Ile 975	Thr	GAA Glu	CAA Gln	ACC Thr	CAG Gln 980	. I y I	TTC Phe	2980

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TTC (GAT Asp	AAC Asn 985	GAC Asp	AGT Ser	AAA Lys	TCA Ser	TTC Phe 990	AGT Ser	GGT Gly	GTA Val	TTA Leu	GAC Asp 995	TGT Cys	GGA Gly	AAC Asn	30	28
TGT 1 Cys S	rcc Ser L000	Arg	ATC Ile	TTT Phe	CAT His	GGA Gly 1005	Glu	AAG Lys	CTT Leu	ATG Met	AAC Asn 1010	Thr	AAC Asn	TTA Leu	ATA Ile	30	76
TTC A Phe I 1015	ATA []e	ATG Met	GTT Val	GAG Glu	AGC Ser 1020	Lys	GGG Gly	ACA Thr	TGT Cys	CCA Pro 1025	Cys	GAC Asp	ACA Thr	CGA Arg	CTG Leu 1030	31	24
CTC A	ATA [le	CAA Gln	Ala	GAG Glu 1035	GIN	ACT Thr	TCT Ser	GAC Asp	GGT Gly 1040	Pro	AAT Asn	CCT Pro	TGT Cys	GAC Asp 1045	Met	31	72
GTT A Val L	AG ys	GIN	CCT Pro 1050	Arg	TAC Tyr	CGA Arg	AAA Lys	GGG Gly 1055	Pro	GAT Asp	GTC Val	TGC Cys	TTT Phe 1060	Asp	AAC Asn	32:	20
AAT G Asn V	al.	TTG Leu 1065	GTI	GAT Asp	TAT Tyr	ACT Thr	GAC Asp 1070	Cys	GGT Gly	GGT Gly	GTT Val	TCT Ser 1075	Gly	TTA Leu	AAT Asn	32	68
CCC T Pro S 1	CC er 080	CTG Leu	TGG Trp	TAT Tyr	Ile	ATT Ile 1085	Gly	ATC Ile	CAG Gln	Phe	CTA Leu 1090	Leu	CTT Leu	TGG Trp	CTG Leu	33:	16
STA T Val S 1095	CT (GGC :	AGC . Ser	Ihr	CAC His 1100	Arg	CTG Leu	TTA Leu	TGAC	CTTC	TA A	AAAC	CAAA	T		336	53
CTGCA	TAG:	TT A	AACT	CCAG	A CC	CTGC	CAAA	ACA	TGAG	CCC	TGCC	CTCA	AT T	ACAG	TAACG	342	23
raggg'	TCA	SC T	ATAA	AATC.	A GA	CAAA	CATT	AGC	TGGG	CCT	GTTC	CATG	GC A	TAAC	ACTAA	348	33
GCGC	AGA	CT C	CTAA	GGCA	c cc	ACTG	GCTG	CAT	GTCA	GGG	TGTC	AGAT	CC T	TAAA	CGTGT	354	13
TGAA'	TGC	rg cz	ATCA'	rcta'	T GT	GTAA	CATC	AAA	GCAA	AAT	CCTA	TACG	TG T	CCTC'	TATTG	360)3
'AAAA	TTTC	G G	CGTT	rgtt	G TT	GCAT'	IGTT	GGT								363	16

(2) INFORMATION FOR SEQ ID NO:30:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 3585 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (ix) FEATURE:

 - (A) NAME/KEY: CDS
 (B) LOCATION: 35..3295

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		(D) OTI	ier .	LNFOI	RIVIAT.	LON:	/500	anda.	L C4						
	(ix)	A)	TURE) NAI) LO	νΕ/KI						٠						
	(ix)	(A	TURE) NAI) LO	ME/K	EY:	3′UT 3296	R 35	85								
	(xi)	SEQ	UENC:	E DE	SCRI	PTIO	N: S	EQ I	D NO	:30:						
GCGG	GGGA	.GG G	GGCA	TTGA	T CT	TCGA	TCGC	GAA	Me	G GC t Al 1	T GC a Al	T GG a Gl	C TG y Cy	C CT s Le 5	G u	52
CTG Leu	GCC Ala	TTG Leu	ACT Thr 10	CTG Leu	ACA Thr	CTT Leu	TTC Phe	CAA Gln 15	TCT Ser	TTG Leu	CTC . Leu	ATC (GGC Gly 20	CCC Pro	TCG Ser	100
TCG Ser	GAG Glu	GAG Glu 25	CCG Pro	TTC Phe	CCT Pro	TCG Ser	GCC Ala 30	GTC Val	ACT Thr	ATC Ile	AAA Lys	TCA Ser 35	TGG Trp	GTG Val	GAT Asp	148
AAG Lys	ATG Met 40	CAA Gln	GAA Glu	GAC Asp	CTT Leu	GTC Val 45	ACA Thr	CTG Leu	GCA Ala	AAA Lys	ACA Thr 50	GCA Ala	AGT Ser	GGA Gly	GTC Val	196
AAT Asn 55	CAG Gln	CTT Leu	GTT Val	GAT Asp	ATT Ile 60	TAT Tyr	GAG Glu	AAA Lys	TAT Tyr	CAA Gln 65	GAT Asp	TTG Leu	TAT Tyr	ACT Thr	GTG Val 70	244
GAA Glu	CCA Pro	AAT Asn	AAT Asn	GCA Ala 75	CGC Arg	CAG Gln	CTG Leu	GTA Val	GAA Glu 80	ATT Ile	GCA Ala	GCC Ala	AGG Arg	GAT Asp 85	ATT Ile	292
GAG Glu	AAA Lys	CTT Leu	CTG Leu 90	AGC Ser	AAC Asn	AGA Arg	TCT Ser	AAA Lys 95	GCC Ala	CTG Leu	GTG Val	AGC Ser	CTG Leu 100	GCA Ala	TTG Leu	340
GAA Glu	GCG Ala	GAG Glu 105	Lys	GTT Val	CAA Gln	GCA Ala	GCT Ala 110	CAC His	CAG Gln	TGG Trp	AGA Arg	GAA Glu 115	GAT Asp	TTT Phe	GCA Ala	388
Ser	AAT Asn 120	Glu	GTT Val	GTC Val	TAC Tyr	TAC Tyr 125	ASI	GCA Ala	AAG Lys	GAT Asp	GAT Asp 130		GAT Asp	CCT Pro	GAG Glu	436
Lys 135	Asn	Asp	Ser	Glu	140	GIY	Ser	GIII	AIG	145	נב				Ile 150	484
GAA Glu	GAT ASP	GCT Ala	AAT Asn	TTT Phe 155	GIA	CGA Arg	CAA Gln	ATA 11e	TCI Ser 160		CAG Gln	CAC His	GCA Ala	GCA Ala 165	GTC Val	532

CAT His	T ATT	CCI Pro	T ACT	ASI	T ATO	TAI	GAC Glu	G GG(1 Gl) 175	/ Sei	A ACA	A AT:	r gro	TTA Leu 180	ı Ası	r GAA n Glu	580
CTC	AAC Asn	TGG Trp 185	1111	A AG1	GCC Ala	TTA Leu	GAT Asp 190	GIU	A GTT 1 Val	TTC Phe	AAA Lys	A AAG Lys 195	Asn	CGC Arg	GAG Glu	628
GAA Glu	GAC Asp 200	PIC	TCA Ser	TTA Leu	TTG Leu	TGG Trp 205	GIN	GTI Val	TTI Phe	GGC Gly	Ser 210	Ala	ACT Thr	GGC	CTA Leu	676
GCT Ala 215	3	TA1	TAT	CCA Pro	GCT Ala 220	ser	CCA Pro	TGG	GTT Val	GAT Asp 225	Asn	AGT Ser	AGA Arg	ACT	CCA Pro 230	724
AAT Asn	AAG Lys	ATT	GAC Asp	Leu 235	TAL	GAT Asp	GTA Val	CGC Arg	AGA Arg 240	Arg	CCA Pro	TGG	TAC Tyr	ATC Ile 245	CAA Gln	772
GGA Gly	GCT Ala	GCA Ala	TCT Ser 250	PIO	AAA Lys	GAC Asp	ATG Met	CTT Leu 255	ATT Ile	CTG Leu	GTG Val	GAT Asp	GTG Val 260	AGT Ser	GGA Gly	820
AGT Ser	GTT Val	AGT Ser 265	GGA Gly	TTG Leu	ACA Thr	CTT Leu	AAA Lys 270	CTG Leu	ATC Ile	CGA Arg	ACA Thr	TCT Ser 275	GTC Val	TCC Ser	GAA Glu	868
ATG Met	TTA Leu 280	GAA Glu	ACC Thr	CTC Leu	TCA Ser	GAT Asp 285	GAT Asp	GAT Asp	TTC Phe	GTG Val	AAT Asn 290	GTA Val	GCT Ala	TCA Ser	TTT Phe	916
AAC Asn 295	AGC Ser	AAT Asn	GCT Ala	CAG Gln	GAT Asp 300	GTA Val	AGC Ser	TGT Cys	TTT Phe	CAG Gln 305	CAC His	CTT Leu	GTC Val	CAA Gln	GCA Ala 310	964
AAT Asn	GTA Val	AGA Arg	AAT Asn	AAA Lys 315	AAA Lys	GTG Val	TTG Leu	AAA Lys	GAC Asp 320	GCG Ala	GTG Val	AAT Asn	AAT Asn	ATC Ile 325	ACA Thr	1012
GCC Ala	AAA Lys	GGA Gly	ATT Ile 330	ACA Thr	GAT Asp	TAT Tyr	AAG Lys	AAG Lys 335	GGC Gly	TTT Phe	AGT Ser	TTT Phe	GCT Ala 340	TTT Phe	GAA Glu	1060
CAG Gln	ren	CTT Leu 345	AAT Asn	TAT Tyr	AAT Asn	Val	TCC Ser 350	AGA Arg	GCA Ala	AAC Asn	TGC Cys	AAT Asn 355	AAG Lys	ATT Ile	ATT Ile	1108
ATG Met	CTA Leu 360	TTC Phe	ACG Thr	GAT Asp	GIÀ	GGA Gly 365	GAA Glu	GAG Glu	AGA Arg	Ala	CAG Gln 370	GAG Glu	ATA Ile	TTT Phe	AAC Asn	1156
AAA Lys 375	TAC Tyr	AAT Asn	AAA Lys	Asp	AAA Lys 380	AAA Lys	GTA Val	CGT Arg	Val	TTC Phe 385	AGG Arg	TTT Phe	TCA Ser	Val	GGT Gly 390	1204

CAA Gln	CAC His	AAT Asn	TAT Tyr	Glu	AGA Arg	GGA Gly	CCT Pro	ATT Ile	CAG Gln 400	TGG Trp	ATG Met	GCC Ala	Cys	GAA Glu 405	AAC Asn	1252
AAA Lys	GGT Gly	TAT Tyr	TAT Tyr 410	395 TAT Tyr	GAA Glu	ATT Ile	CCT Pro	TCC Ser 415	ATT	GGT Gly	GCA Ala	ATA Ile	AGA Arg 420	ATC Ile	AAT Asn	1300
ACT Thr	CAG Gln	GAA Glu 425	TAT Tyr	TTG Leu	GAT Asp	GTT Val	TTG Leu 430	GGA Gly	AGA Arg	CCA Pro	ATG Met	GTT Val 435	TTA Leu	GCA Ala	GGA Gly	1348
GAC Asp	AAA Lys 440	GCT Ala	AAG Lys	CAA Gln	GTC Val	CAA Gln 445	TGG Trp	ACA Thr	AAT Asn	GTG Val	TAC Tyr 450	CTG Leu	GAT Asp	GCA Ala	TTG Leu	1396
GAA Glu 455	CTG Leu	GGA Gly	CTT Leu	GTC Val	ATT Ile 460	ACT Thr	GGA Gly	ACT Thr	CTT Leu	CCG Pro 465	GTC Val	TTC Phe	AAC Asn	ATA Ile	ACC Thr 470	1444
GGC Gly	CAA Gln	TTT Phe	GAA Glu	AAT Asn 475	AAG Lys	ACA Thr	AAC Asn	TTA Leu	AAG Lys 480	AAC Asn	CAG Gln	CTG Leu	ATT Ile	CTT Leu 485	GGT Gly	1492
GTG Val	ATG Met	GGA Gly	GTA Val 490	GAT Asp	GTG Val	TCT Ser	TTG Leu	GAA Glu 495	Asp	ATT Ile	AAA Lys	AGA Arg	CTG Leu 500	ACA Thr	CCA Pro	1540
CGT Arg	TTT Phe	ACA Thr 505	CTG Leu	TGC Cys	CCC Pro	AAT Asn	GGG Gly 510	Tyr	TAC Tyr	TTT Phe	GCA Ala	ATC Ile 515	GAT Asp	CCT Pro	AAT Asn	1588
GGT Gly	TAT Tyr	GTT Val	TTA Leu	TTA Leu	CAT His	CCA Pro	AAT Asn	CTT Leu	CAG Gln	CCA Pro	ъys	GIU	CCA Pro	GTA Val	ACA Thr	1636
	520					525					530					
TTG Leu 535	Asp	TTC	CTT Leu	GAT Asp	GCA Ala 540	GIU	Leu	GAG Glu	AAT Asn	GAT Asp 545		AAA Lys	GTG Val	GAG Glu	ATT Ile 550	1684
CGA Arg	LAA I	AAG	ATG Met	ATT	: Asp	GGG Gly	GAA Glu	AGI Ser	GGA Gly 560	GIU	AAA Lys	ACA Thr	TTC	AGA Arg 565	ACT	1732
CT0 Lev	GTT 1 Val	AA! Lys	A TCT S Ser 570	GLI	A GAT	GAG Glu	AGA 1 Arg	TAT TY1 575		GAC Asp	AAA Lys	GGA Gly	AAC Asn 580	AGG Arg	ACA Thr	1780
TAC Ty:	C ACA	TG(Tr]	o Thi	A CC	r GTC o Val	AAT L Asi	GG(1 Gl; 59	y 1111	A GAT	TAC Tyl	AG! Set	r TTC Lev 599	GCC Ala	TTC Lev	GTA Val	1828
TT: Let	A CCI u Pro 600	A AC		C AG'	r TT'	TAC E Ty: 60!	C TA	r AT	A AAI e Ly:	A GCC s Ala	C AA a Lya 61		A GAA	A GAC	ACA Thr	1876

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615	5	. 611	I AL	a Arç	620) L Lys	: Lys	e GT	Lys	625	Ly:	s Ası	Sex	r Glı	A ACC Thr 630	1924
Dec	. Dys	PIC	ASI	635	; Pne	e GIV	i GIt	ı Ser	640	Tyr	Thi	r Phe	: Ile	Ala 645		1972
AGA Arg	GAI Asp	TAC Tyr	C TGC Cys 650	ASI	GAC Asp	CTG Leu	AAA Lys	ATA Ile 655	Ser	GAT Asp	AA1 Asr	AAC Asn	ACT Thr	Glu	TTT Phe	2020
CTT Leu	TTA Leu	AAT Asn 665	PHE	AAC Asn	GAG Glu	TTT Phe	ATI Ile 670	: Asp	AGA Arg	AAA Lys	ACI Thr	CCA Pro 675	Asn	AAC Asn	CCA Pro	2068
TCA Ser	TGT Cys 680	WPII	GCG Ala	GAT Asp	TTG Leu	ATT Ile 685	AAT Asn	AGA Arg	GTC Val	TTG Leu	CTI Leu 690	Asp	GCA Ala	GGC	TTT Phe	2116
695	7.511	Giu	Leu	val	700	Asn	Tyr	Trp	Ser	Lys 705	Gln	Lys	Asn	Ile	710	2164
GGA Gly	GTG Val	AAA Lys	GCA Ala	CGA Arg 715	TTT Phe	GTT Val	GTG Val	ACT Thr	GAT Asp 720	GGT Gly	GGG Gly	ATT Ile	ACC Thr	AGA Arg 725	GTT Val	2212
- y -	110	Lys	730	GCT Ala	GIY	GIU	Asn	735	Gln	Glu	Asn	Pro	Glu 740	Thr	Tyr	2260
GIU	лэр	745	Pne	TAT Tyr	туѕ	Arg	750	Leu	Asp	Asn	Asp	Asn 755	Tyr	Val	Phe	2308
ACT Thr	GCT Ala 760	CCC Pro	TAC Tyr	TTT Phe	AAC Asn	AAA Lys 765	AGT Ser	GGA Gly	CCT Pro	GGT Gly	GCC Ala 770	TAT Tyr	GAA Glu	TCG Ser	GGC Gly	2356
ATT Ile 775	ATG Met	GTA Val	AGC Ser	AAA Lys	GCT Ala 780	GTA Val	GAA Glu	ATA Ile	TAT Tyr	ATT Ile 785	CAA Gln	GGG Gly	AAA Lys	CTT Leu	CTT Leu 790	2404
AAA Lys	CCT Pro	GCA Ala	GTT Val	GTT Val 795	GGA Gly	ATT Ile	AAA Lys	ATT Ile	GAT Asp 800	GTA Val	AAT Asn	TCC Ser	TGG Trp	ATA Ile 805	GAG Glu	2452
TAA naA	TTC Phe	ACC Thr	AAA Lys 810	ACC Thr	TCA Ser	ATC Ile	AGA Arg	GAT Asp 815	CCG Pro	TGT Cys	GCT Ala	GGT Gly	CCA Pro 820	GTT Val	TGT Cys	2500
GAC Asp	Cys	AAA Lys 825	AGA Arg	AAC Asn	AGT Ser	Asp	GTA Val 830	ATG Met	GAT Asp	TGT Cys	GTG Val	ATT Ile 835	CTG Leu	GAT Asp	GAT Asp	2548

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GGT Gly	GGG Gly 840	TTT Phe	CTT Leu	CTG Leu	ATG Met	GCA Ala 845	AAT Asn	CAT His	GAT Asp	GAT Asp	TAT Tyr 850	ACT Thr	TAA Asn	CAG Gln	ATT Ile	2596
GGA Gly 855	AGA Arg	TTT Phe	TTT Phe	GGA Gly	GAG Glu 860	ATT Ile	GAT Asp	CCC Pro	AGC Ser	TTG Leu 865	ATG Met	AGA Arg	CAC His	CTG Leu	GTT Val 870	2644
AAT Asn	ATA Ile	TCA Ser	GTT Val	TAT Tyr 875	GCT Ala	TTT Phe	AAC Asn	AAA Lys	TCT Ser 880	TAT Tyr	GAT Asp	TAT Tyr	CAG Gln	TCA Ser 885	GTA Val	2692
TGT Cys	GAG Glu	CCC Pro	GGT Gly 890	GCT	GCA Ala	CCA Pro	AAA Lys	CAA Gln 895	GGA	GCA Ala	GGA Gly	CAT His	CGC Arg 900	TCA Ser	GCA Ala	2740
TAT Tyr	GTG Val	CCA Pro 905	TCA Ser	GTA Val	GCA Ala	GAC Asp	ATA Ile 910	TTA Leu	CAA Gln	ATT Ile	GGC Gly	TGG Trp 915	TGG Trp	GCC Ala	ACT Thr	2788
GCT Ala	GCT Ala 920	GCC Ala	TGG Trp	TCT Ser	ATT Ile	CTA Leu 925	CAG Gln	CAG Gln	TTT Phe	CTC Leu	TTG Leu 930	AGT Ser	TTG Leu	ACC Thr	TTT Phe	2836
CCA Pro 935	CGA Arg	CTC Leu	CTT Leu	GAG Glu	GCA Ala 940	GTT Val	GAG Glu	ATG Met	GAG Glu	GAT Asp 945	GAT Asp	GAC Asp	TTC Phe	ACG Thr	GCC Ala 950	2884
TCC Ser	CTG Leu	TCC Ser	AAG Lys	CAG Gln 955	AGC Ser	TGC Cys	ATT Ile	ACT Thr	GAA Glu 960	CAA Gln	ACC Thr	CAG Gln	TAT Tyr	TTC Phe 965	TTC Phe	2932
GAT Asp	AAC Asn	GAC Asp	AGT Ser 970	AAA Lys	TCA Ser	TTC Phe	AGT Ser	GGT Gly 975	GTA Val	TTA Leu	GAC Asp	TGT Cys	GGA Gly 980	AAC Asn	TGT Cys	2980
TCC Ser	AGA Arg	ATC Ile 985	Phe	CAT His	GGA Gly	GAA Glu	AAG Lys 990	CTT	ATG Met	AAC Asn	ACC Thr	AAC Asn 995	TTA Leu	ATA Ile	TTC Phe	3028
ATA Ile	ATG Met 100	Val	GAG Glu	AGC Ser	AAA Lys	GGG Gly 100	Thr	TGT Cys	CCA Pro	TGT Cys	GAC Asp 101	Thr	CGA Arg	CTG Leu	CTC Leu	3076
ATA Ile 101	Gln	GCG Ala	GAG Glu	CAG Gln	ACT Thr 102	Ser	GAC Asp	GGT Gly	CCA Pro	AAT Asn 102	Pro	TGT Cys	GAC Asp	ATG Met	GTT Val 1030	3124
AAG Lys	CAA Gln	CCT Pro	AGA Arg	TAC	Arg	AAA Lys	GGG Gly	CCT	GAT Asp 104	val	TGC Cys	TTT Phe	GAT Asp	AAC Asn 104	AAT Asn 5	3172
GTC Val	TTG Leu	GAG	GAT Asp 105	тух	ACT Thr	GAC Asp	TGT Cys	GGT Gly	, ст	GTI Val	TCI Ser	GGA Gly	TTA Leu 106	. Au	CCC Pro	3220
TCC	CTG	TGC	TAT	TATO	TTA :	GGA	TA	CAC	TTI	CTA	CT.	A CTI	TGG	CTO	GTA	3268

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Ser Leu Trp Tyr Ile Ile Gly Ile Gln Phe Leu Leu Leu Trp Leu Val 1065 1070 1075	
TCT GGC AGC ACA CAC CGG CTG TTA TGACCTTCTA AAAACCAAAT CTGCATAGTT Ser Gly Ser Thr His Arg Leu Leu 1080 1085	3322
AAACTCCAGA CCCTGCCAAA ACATGAGCCC TGCCCTCAAT TACAGTAACG TAGGGTCAGC	3382
TATAAAATCA GACAAACATT AGCTGGGCCT GTTCCATGGC ATAACACTAA GGCGCAGACT	3442
CCTAAGGCAC CCACTGGCTG CATGTCAGGG TGTCAGATCC TTAAACGTGT GTGAATGCTG	3505
CATCATCTAT GTGTAACATC AAAGCAAAAT CCTATACGTG TCCTCTATTG GAAAATTTGG	3562
GCGTTTGTTG TTGCATTGTT GGT	3585
(2) INFORMATION FOR SEQ ID NO:31:	5505
(i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 3564 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: DNA (genomic)	
(ix) FEATURE:	
 (A) NAME/KEY: CDS (B) LOCATION: 353374 (Δ1625 to 1639 & Δ1908 to 1928) (D) OTHER INFORMATION: /standard_name= "Alpha-2d" 	
(ix) FEATURE: (A) NAME/KEY: 5'UTR (B) LOCATION: 134	
(ix) FEATURE:	
(A) NAME/KEY: 3'UTR (B) LOCATION: 33753565	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:31:	
GCGGGGGAGG GGGCATTGAT CTTCGATCGC GAAG ATG GCT GCT GGC TGC CTG	52
Met Ala Ala Gly Cys Leu 1 5	52
CTG GCC TTG ACT CTG ACA CTT TTC CAA TCT TTG CTC ATC GGC CCC TCG Leu Ala Leu Thr Leu Thr Leu Phe Gln Ser Leu Leu Ile Gly Pro Ser 10 15 20	100
TCG GAG GAG CCG TTC CCT TCG GCC GTC ACT ATC AAA TCA TGG GTG GAT Ser Glu Glu Pro Phe Pro Ser Ala Val Thr Ile Lys Ser Trp Val Asp 25 30 35	148
AAG ATG CAA GAA GAC CTT GTC ACA CTG GCA AAA ACA GCA AGT GGA GTC	

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	40					45					50						
AAT Asn 55	CAG Gln	CTT Leu	GTT Val	GAT Asp	ATT Ile 60	TAT Tyr	GAG Glu	AAA Lys	TAT Tyr	CAA Gln 65	GAT Asp	TTG Leu	TAT Tyr	ACT Thr	GTG Val 70	-	244
GAA Glu	CCA Pro	AAT Asn	AAT Asn	GCA Ala 75	CGC Arg	CAG Gln	CTG Leu	GTA Val	GAA Glu 80	ATT Ile	GCA Ala	GCC Ala	AGG Arg	GAT Asp 85	ATT Ile		292
GAG Glu	AAA Lys	CTT Leu	CTG Leu 90	AGC Ser	AAC Asn	AGA Arg	TCT Ser	AAA Lys 95	GCC Ala	CTG Leu	GTG Val	AGC Ser	CTG Leu 100	GCA Ala	TTG Leu		340
GAA Glu	GCG Ala	GAG Glu 105	AAA Lys	GTT Val	CAA Gln	GCA Ala	GCT Ala 110	CAC His	CAG Gln	TGG Trp	AGA Arg	GAA Glu 115	GAT Asp	TTT Phe	GCA Ala		388
AGC Ser	AAT Asn 120	GAA Glu	GTT Val	GTC Val	TAC Tyr	TAC Tyr 125	AAT Asn	GCA Ala	AAG Lys	GAT Asp	GAT Asp 130	CTC Leu	GAT Asp	CCT Pro	GAG Glu		436
AAA Lys 135	AAT Asn	GAC Asp	AGT Ser	GAG Glu	CCA Pro 140	GGC Gly	AGC Ser	CAG Gln	AGG Arg	ATA Ile 145	AAA Lys	CCT Pro	GTT Val	TTC Phe	ATT Ile 150		484
GAA Glu	GAT Asp	GCT Ala	AAT Asn	TTT Phe 155	GGA Gly	CGA Arg	CAA Gln	ATA Ile	TCT Ser 160	TAT Tyr	CAG Gln	CAC His	GCA Ala	GCA Ala 165	GTC Val		532
CAT His	ATT Ile	CCT Pro	ACT Thr 170	Asp	ATC Ile	TAT Tyr	GAG Glu	GGC Gly 175	Ser	ACA Thr	ATT Ile	GTG Val	TTA Leu 180	AAT Asn	GAA Glu		580
CTC Leu	AAC Asn	TGG Trp 185	Thr	AGT Ser	GCC Ala	TTA Leu	GAT Asp 190	Gru	GTT Val	TTC Phe	AAA Lys	AAG Lys 195		CGC Arg	GAG Glu		628
GAA Glu	GAC Asp	Pro	TCA Ser	TTA Leu	TTG Leu	TGG Trp 205	GIII	GTT Val	TTT Phe	GGC Gly	AGT Ser 210		ACT	GGC	CTA Leu		676
GCT Ala 215	Arg	TAT	TAT Tyr	CCA Pro	GCT Ala 220	Ser	CCA Pro	TGG Trp	GTI Val	GAT Asp 225	, mor	AGT Ser	AGA Arg	ACT Thr	CCA Pro 230		724
AA Ası	AAC Lys	ATT	GAC Asj	CTT Lev 235	ı Tyr	GAT Asp	GTA Val	CGC Arg	AGA Arg 240	, ALC	A CCI	TGC Tr	TAC Tyr	ATC 116 245	CAA Gln		772
GG/ Gl	A GCT	r GCI a Ala	A TC: a Se: 25	r Pro	Lys	A GAC S Asp	ATO Met	CT:	, <u>, , , , , , , , , , , , , , , , , , </u>	CTC Lev	GT(G GA'	r GTC o Val 260		GGA Gly		820
AG' Se:	r GT' r Va	r AG			3 ACA 1 Thi	A CT?	r AAI 1 Ly:	A CTO	G ATO	c CG	A AC	A TC' r Se	r GTO	C TCC	GAA r Glu		868

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		26	5				270)				27	5			
AT(TTI Let 280	a GT	A ACC	CTC Lev	TCA Ser	GAT Asp 285	Asp	GA:	r TT(Phe	GT(AA: L Asi 290	ı Va	A GC	T TC	A TTT	916
AAG Asi 295		C AA'	r GCT n Ala	CAG Glr	GAT Asp 300	val	AGC Ser	TG1 Cys	TTT Phe	CAG Glr 305	ı His	CT:	r GTG u Val	C CAM	A GCA Ala 310	964
AA7 Asr	r GTA	A AGA L Arg	LAA A ISA U	AAA Lys 315	Lys	GTG Val	TTG Leu	AAA Lys	GAC Asp 320	Ala	GTG Val	AA7 Ası	CAA 1 LEA 1	T ATC 1 11e 325	C ACA Thr	1012
GCC Ala	AAA Lys	GGZ Gly	ATI Ile 330	THE	GAT Asp	TAT	AAG Lys	AAG Lys 335	Gly	TTT Phe	AGT Ser	TTI Phe	GCT Ala 340	Phe	GAA Glu	1060
CAG Gln	CTG Leu	CTI Leu 345	MSII	TAT	AAT Asn	GTT Val	TCC Ser 350	AGA Arg	GCA Ala	AAC Asn	TGC Cys	AAT Asn 355	Lys	ATI	ATT Ile	1108
ATG Met	CTA Leu 360	FILE	ACG Thr	GAT Asp	GGA Gly	GGA Gly 365	GAA Glu	GAG Glu	AGA Arg	GCC Ala	CAG Gln 370	GAG Glu	ATA Ile	TTT	AAC Asn	1156
AAA Lys 375	TAC Tyr	AAT Asn	AAA Lys	GAT Asp	AAA Lys 380	AAA Lys	GTA Val	CGT Arg	GTA Val	TTC Phe 385	AGG Arg	TTT Phe	TCA Ser	GTT Val	GGT Gly 390	1204
CAA Gln	CAC His	AAT Asn	TAT Tyr	GAG Glu 395	AGA Arg	GGA Gly	CCT Pro	ATT Ile	Gln	TGG Trp	ATG Met	GCC Ala	TGT Cys	Glu	AAC Asn	1252
AAA Lys	GGT Gly	TAT Tyr	TAT Tyr 410	TAT	GAA Glu	ATT Ile	CCT Pro	TCC Ser 415	400 ATT Ile	GGT Gly	GCA Ala	ATA Ile	AGA Arg 420	405 ATC Ile	AAT Asn	1300
ACT Thr	CAG Gln	GAA Glu 425	TAT Tyr	TTG Leu	GAT Asp	GTT Val	TTG Leu 430	GGA Gly	AGA Arg	CCA Pro	ATG Met	GTT Val 435	TTA Leu	GCA Ala	GGA Gly	1348
GAC Asp	AAA Lys 440	GCT Ala	AAG Lys	CAA Gln	Val	CAA Gln 445	TGG Trp	ACA Thr	AAT Asn	GTG Val	TAC Tyr 450	CTG Leu	GAT Asp	GCA Ala	TTG Leu	1396
GAA Glu 455	CTG Leu	GGA Gly	CTT Leu	Vai	ATT Ile 460	ACT Thr	GGA Gly	ACT Thr	Leu	CCG Pro 465	GTC Val	TTC Phe	AAC Asn	ATA Ile	ACC Thr 470	1444
GGC Gly	CAA Gln	TTT Phe	GAA Glu	AAT Asn 475	AAG . Lys '	ACA . Thr .	AAC Asn	Leu	AAG Lys 480	AAC Asn	CAG Gln	CTG Leu	ATT Ile	CTT Leu 485	GGT Gly	1492
GTG Val	ATG Met	GGA Gly	GTA Val 490	GAT Asp	GTG :	TCT ' Ser :	Leu (GAA Glu 495	GAT . Asp	ATT Ile	AAA Lys	AGA Arg	CTG Leu 500	ACA Thr	CCA Pro	1540

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CGT Arg	TTT Phe	Thr	CTG Leu	TGC Cys	CCC Pro	AAT Asn	GGG Gly 510	TAT Tyr	TAC Tyr	TTT Phe	GCA Ala	ATC Ile 515	GAT Asp	CCT Pro	AAT Asn	1588
GGT Gly	TAT Tyr	505 GTT Val	TTA Leu	TTA Leu	CAT His	CCA Pro	יים <i>ב</i>	CTT Leu	CAG Gln	CCA Pro	AAG Lys	GAG Glu	CCA Pro	GTA Val	ACA Thr	1636
	520					525					530					
TTG Leu 535	GAT Asp	TTC Phe	CTT Leu	GAT Asp	GCA Ala 540	GAG Glu	TTA Leu	GAG Glu	AAT Asn	GAT Asp 545	ATT Ile	AAA Lys	GTG Val	GAG Glu	ATT Ile 550	1684
CGA Arg	AAT Asn	AAG Lys	ATG Met	ATT Ile 555	GAT Asp	GGG Gly	GAA Glu	AGT Ser	GGA Gly 560	GAA Glu	AAA Lys	ACA Thr	TTC Phe	AGA Arg 565	ACT Thr	1732
CTG Leu	GTT Val	AAA Lys	TCT Ser 570	CAA Gln	GAT Asp	GAG Glu	AGA Arg	TAT Tyr 575	ATT Ile	GAC Asp	AAA Lys	GGA Gly	AAC Asn 580	AGG Arg	ACA Thr	1780
TAC Tyr	ACA Thr	TGG Trp 585	ACA Thr	CCT Pro	GTC Val	AAT Asn	GGC Gly 590	Thr	GAT Asp	TAC Tyr	AGT Ser	TTG Leu 595	GCC Ala	TTG Leu	GTA Val	1828
TTA Leu	CCA Pro 600	Thr	TAC Tyr	AGT Ser	TTT Phe	TAC Tyr 605	TAT Tyr	ATA Ile	AAA Lys	GCC Ala	AAA Lys 610	Leu	GAA Glu	GAG Glu	ACA Thr	1876
ATA Ile 615	Thr	CAG Gln	GCC Ala	AGA Arg	TAT Tyr 620	TCG Ser	GAA Glu	ACC Thr	CTG Leu	AAG Lys 625	CCA Pro	GAT Asp	AAT Asn	TTT Phe	GAA Glu 630	1924
GAA Glu	TCT Ser	GGC	TAT	ACA Thr 635	TTC Phe	ATA Ile	GCA Ala	CCA Pro	AGA Arg 640	Asp	TAC	TGC Cys	AAT Asn	GAC Asp 645	CTG Leu	1972
AAA Lys	ATA Ile	TCG Ser	GAT Asp 650	Asn	AAC Asn	ACT Thr	GAA Glu	TTT Phe 655	Leu	TTA Leu	AAT Asn	TTC Phe	AAC Asn 660		TTT Phe	2020
ATT Ile	GAI Asp	AGA Arg	, Lys	ACT Thr	CCA Pro	AAC	AAC Asr 670	PIC	TCA Ser	TGT Cys	AAC Asi	GCG Ala 675		TTG Leu	ATT	2068
AAT Asn	AGA Arg	y Val	L Leu	Lev	GAT Asp	Ala	GT.	Pne	ini	ASI	GAZ Glu 690	, Dec	GTC Val	CAA Gln	AAT Asn	2116
TAC Tyr 695	Tr	AG: Sei	r AAG c Lys	CAG Glr	AAA Lys	ASI	T ATO	C AAG e Lys	G GG/ G Gly	A GTO 7 Val 70:	· Ly.	A GCA s Ala	A CGA A Arg	TTI Phe	GTT Val 710	2164
GT(Va	AC'	r GA' r Asj	r GG7 p Gly	GGG Gly 715	/ 11e	ACC Thi	C AG	A GT' g Vai	TA:		AA D Ly	A GAC s Glu	G GCT	GG7 a Gly 729	A GAA / Glu	2212

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AA :eA	T TG	p G	AA ln	GAA Glu 730		C CC	A GA o Gl	G AC u Th	A TA r Ty 73	r GI	G GA u As	C AG p Se	C TT	C TA e Ty 74	r Ly	A AGG 's Arg	2260
AG Se:	C CT r Le		AT sp 45	AA1 Asn	GAT Asp	AA T	C TA	T GT r Va 75	T bu	C AC	T GC r Al	T CC a Pr	C TA O Ty: 75:	r Ph	T AA e As	C AAA n Lys	2308
AG' Se:	r GG r Gl 76	y -	CT ro	GGT Gly	GCC Ala	TA:	GAI Gli 76	ı Se:	G GG(r Gl ₃	C AT	r at	G GT t Va: 770	l Se	C AA	A GC s Al	T GTA a Val	2356
GAI Glu 775	A AT	A T	AT yr	ATT Ile	CAA Gln	GG(Gl ₃ 78(, rai	A CT	r cri	T AAI 1 Lys	785	o Ala	A GTT a Val	r GTT L Val	r GG	A ATT Y Ile 790	2404
AA7 Lys	AT'	r g/ ≥ As	AT Sp	GTA Val	AAT Asn 795	261	Tr	ATA Ile	A GAG	AAT Asr 800	Phe	C ACC	AAA Lys	ACC Thr	TCZ Sei	A ATC	2452
AGA Arg	GAT Asp	C CC		TGT Cys 810	GCT Ala	GG1 Gly	Pro	GTI Val	TGI Cys 815	Asp	TGC Cys	AAA Lys	A AGA Arg	AAC Asn 820	Sex	GAC Asp	2500
GTA Val	Met	GA As	F,	TGT Cys	GTG Val	ATT	CTG	GAT Asp 830	Asp	GGT Gly	GGG	TTT Phe	CTT Leu 835	Leu	ATC Met	GCA Ala	2548
AAT Asn	CAT His 840		T (SAT Asp	TAT Tyr	ACT Thr	AAT Asn 845	CAG Gln	ATT	GGA Gly	AGA Arg	TTT Phe 850	Phe	GGA Gly	GAG Glu	ATT	2596
GAT Asp 855	CCC	AG Se	C 1 r I	rTG Leu	ATG Met	AGA Arg 860	CAC His	CTG Leu	GTT Val	AAT Asn	ATA Ile 865	TCA Ser	GTT Val	TAT Tyr	GCT Ala	TTT Phe 870	2644
	Lys	36.		Ϋ́	875	Tyr	Gin	Ser	Val	Cys	Glu	Pro	GGT Gly	Ala	Ala	Pro	2692
Lys	Gln	Gl	Y A	la 90	GGA Gly	CAT His	CGC Arg	TCA Ser	GCA Ala 895	TAT Tyr	GTG Val	CCA Pro	TCA Ser	GTA Val 900	GCA Ala	GAC Asp	274 0
ATA Ile	TTA Leu	Gl: 905	т т	TT (GGC Gly	TGG Trp	TGG Trp	GCC Ala 910	ACT Thr	GCT Ala	GCT Ala	GCC Ala	TGG Trp 915	TCT Ser	ATT Ile	CTA Leu	2788
CAG Gln	CAG Gln 920	TTT Phe	C C	TC !	TTG . Leu	AGT Ser	TTG Leu 925	ACC Thr	TTT Phe	CCA Pro	CGA Arg	CTC Leu 930	CTT Leu	GAG Glu	GCA Ala	GTT Val	2836
GAG Glu 935	ATG Met	GAG Glu	G A	AT (sp /	usp.	GAC Asp 940	TTC Phe	ACG Thr	GCC Ala	TCC Ser	CTG Leu 945	TCC Ser	AAG Lys	CAG Gln	AGC Ser	TGC Cys 950	2884
TTA	ACT	GAA	C.	AA A	ACC (CAG	TAT	TTC	TTC	GAT	AAC	GAC	AGT	AAA	TCA	TTC	2932

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				955		•		••••	960		•		-	965	Phe	
AGT Ser	GGT Gly	GTA Val	TTA Leu 970	GAC Asp	TGT Cys	GGA Gly	AAC Asn	TGT Cys 975	TCC Ser	AGA Arg	ATC Ile	TTT Phe	CAT His 980	GGA Gly	GAA Glu	2980
AAG Lys	CTT Leu	ATG Met 985	AAC Asn	ACC Thr	AAC Asn	TTA Leu	ATA Ile 990	TTC Phe	ATA Ile	ATG Met	GTT Val	GAG Glu 995	AGC Ser	AAA Lys	GGG Gly	3028
ACA Thr	TGT Cys 1000	Pro	TGT Cys	GAC Asp	ACA Thr	CGA Arg 100	Leu	CTC Leu	ATA Ile	CAA Gln	GCG Ala 1010	Glu	CAG Gln	ACT Thr	TCT Ser	3076
GAC Asp 1015	Gly	CCA Pro	AAT Asn	CCT Pro	TGT Cys 1020	Asp	ATG Met	GTT Val	AAG Lys	CAA Gln 1025	Pro	AGA Arg	TAC Tyr	CGA Arg		3124
GGG Gly	CCT Pro	GAT Asp	GTC Val	TGC Cys 103!	Phe	GAT Asp	AAC Asn	AAT Asn	GTC Val 1040	Leu	GAG Glu	GAT Asp	TAT Tyr	ACT Thr 104	Asp	3172
TGT Cys	GGT Gly	GGT Gly	GTT Val 105	Ser	GGA Gly	TTA Leu	AAT Asn	CCC Pro 105	Ser	CTG Leu	TGG Trp	TAT Tyr	ATC Ile 106	Ile	GGA Gly	3220
ATC Ile	CAG Gln	TTT Phe 106	Leu	CTA Leu	CTT Leu	TGG Trp	CTG Leu 107	Val	TCT Ser	GGC Gly	AGC Ser	ACA Thr 107	His	CGG Arg	CTG Leu	3268
TTA Leu		CCTT	CTA :	AAAA	CCAA	AT C	rgca'	TAGT"	T AA	ACTC	CAGA	CCC'	TGCC	AAA		3321
ACA'	TGAG	ccc	TGCC	CTCA	AT T	ACAG	TAAC	G TA	GGGT	CAGC	TAT.	AAAA	TCA (GACA	AACATT	3381
AGC	TGGG	CCT	GTTC	CATG	GC A	TAAC	ACTA	A GG	CGCA	GACT	CCT	AAGG	CAC	CCAC	TGGCTG	3441
CAT	GTCA	GGG	TGTC	agat	сс т	AAAT	CGTG	T GT	GAAT	GCTG	CAT	CATC	TAT	GTGT.	AACATC	3501
AAA	GCAA	AAT	CCTA	TACG	TG T	CCTC	TATT	g ga	AAAT	TTGG	GCG	TTTG	TTG	TTGC	ATTGTT	3561
GGT																3564

(2) INFORMATION FOR SEQ ID NO:32:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 3579 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (ix) FEATURE:

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(A)	NAME	/KEY:	CDS

(B) LOCATION: 35..3289
(D) OTHER INFORMATION: /standard_name= "Alpha-2e"

(ix) FEATURE:

(A) NAME/KEY: 5'UTR (B) LOCATION: 1..34

(ix) FEATURE:

(A) NAME/KEY: 3'UTR
(B) LOCATION: 3289..3579

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:15:

GCG	GGGG	AGG	GGGC	ATTG	AT C	TTCG	ATCG	C GA	AG A M	TG G et A 1	CT G la A	CT G la G	GC T ly C	GC C ys L 5	TG eu	52
CTG Leu	GCC Ala	TTG Leu	ACT Thr 10	Leu	ACA Thr	CTT Leu	TTC Phe	CAA Gln 15	TCT Ser	TTG Leu	CTC Leu	ATC Ile	GGC Gly 20	CCC Pro	TCG Ser	100
TCG Ser	GAG Glu	GAG Glu 25	CCG Pro	TTC Phe	CCT Pro	TCG Ser	GCC Ala 30	GTC Val	ACT Thr	ATC Ile	AAA Lys	TCA Ser 35	TGG Trp	GTG Val	GAT Asp	148
AAG Lys	ATG Met 40	CAA Gln	GAA Glu	GAC Asp	CTT Leu	GTC Val 45	ACA Thr	CTG Leu	GCA Ala	AAA Lys	ACA Thr 50	GCA Ala	AGT Ser	GGA Gly	GTC Val	196
AAT Asn 55	CAG Gln	CTT Leu	GTT Val	GAT Asp	ATT Ile 60	TAT Tyr	GAG Glu	AAA Lys	TAT Tyr	CAA Gln 65	GAT Asp	TTG Leu	TAT Tyr	ACT Thr	GTG Val 70	244
GAA Glu	CCA Pro	AAT Asn	AAT Asn	GCA Ala 75	CGC Arg	CAG Gln	CTG Leu	GTA Val	GAA Glu 80	ATT Ile	GCA Ala	GCC Ala	AGG Arg	GAT Asp 85	ATT	.292
GAG Glu	AAA Lys	CTT Leu	CTG Leu 90	AGC Ser	AAC Asn	AGA Arg	TCT Ser	AAA Lys 95	GCC Ala	CTG Leu	GTG Val	AGC Ser	CTG Leu 100	GCA Ala	TTG Leu	340
GAA Glu	GCG Ala	GAG Glu 105	AAA Lys	GTT Val	CAA Gln	GCA Ala	GCT Ala 110	CAC His	CAG Gln	TGG Trp	AGA Arg	GAA Glu 115	GAT Asp	TTT Phe	GCA Ala	388
AGC Ser	AAT Asn 120	GAA Glu	GTT Val	GTC Val	TAC Tyr	TAC Tyr 125	AAT Asn	GCA Ala	AAG Lys	GAT Asp	GAT Asp 130	CTC Leu	GAT Asp	CCT Pro	GAG Glu	436
AAA Lys 135	AAT Asn	GAC Asp	AGT Ser	GAG Glu	CCA Pro 140	GGC Gly	AGC Ser	CAG Gln	AGG Arg	ATA Ile 145	AAA Lys	CCT Pro	GTT Val	TTC Phe	ATT Ile 150	484
GAA	GAT	GCT	TAA	TTT	GGA	CGA	CAA	ATA	TCT	TAT	CAG	CAC	GCA	GCA	GTC	532

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Glu	Asp	Ala	Asn	Phe 155	Gly	Arg	Gln	Ile	Ser 160	Tyr	Gln	His	Ala	Ala 165	Val	
CAT His	ATT Ile	CCT Pro	ACT Thr 170	GAC Asp	ATC Ile	TAT Tyr	GAG Glu	GGC Gly 175	TCA Ser	ACA Thr	ATT Ile	GTG Val	TTA Leu 180	AAT Asn	GAA Glu	580
CTC Leu	AAC Asn	TGG Trp 185	ACA Thr	AGT Ser	GCC Ala	TTA Leu	GAT Asp 190	GAA Glu	GTT Val	TTC Phe	AAA Lys	AAG Lys 195	AAT Asn	CGC Arg	GAG Glu	628
					TTG Leu											676
					GCT Ala 220											. 724
					TAT Tyr											772
GGA Gly	GCT Ala	GCA Ala	TCT Ser 250	CCT Pro	AAA Lys	GAC Asp	ATG Met	CTT Leu 255	ATT Ile	CTG Leu	GTG Val	GAT Asp	GTG Val 260	AGT Ser	GGA Gly	820
AGT Ser	GTT Val	AGT Ser 265	GGA Gly	TTG Leu	ACA Thr	CTT Leu	AAA Lys 270	CTG Leu	ATC Ile	CGA Arg	ACA Thr	TCT Ser 275	GTC Val	TCC Ser	GAA Glu	868
ATG Met	TTA Leu 280	GAA Glu	ACC Thr	CTC Leu	TCA Ser	GAT Asp 285	GAT Asp	GAT Asp	TTC Phe	GTG Val	AAT Asn 290	GTA Val	GCT Ala	TCA Ser	TTT Phe	916
AAC Asn 295	AGC Ser	AAT Asn	GCT Ala	CAG Gln	GAT Asp 300	GTA Val	AGC Ser	TGT Cys	TTT Phe	CAG Gln 305	CAC His	CTT Leu	GTC Val	CAA Gln	GCA Ala 310	964
AAT Asn	GTA Val	AGA Arg	AAT Asn	AAA Lys 315	AAA Lys	GTG Val	TTG Leu	AAA Lys	GAC Asp 320	GCG Ala	GTG Val	AAT Asn	AAT Asn	ATC Ile 325	ACA Thr	1012
GCC Ala	AAA Lys	GGA Gly	ATT Ile 330	ACA Thr	GAT Asp	TAT Tyr	AAG Lys	AAG Lys 335	GGC Gly	TTT Phe	AGT Ser	TTT Phe	GCT Ala 340	TTT Phe	GAA Glu	1060
CAG Gln	CTG Leu	CTT Leu 345	AAT Asn	TAT Tyr	AAT Asn	GTT Val	TCC Ser 350	AGA Arg	GCA Ala	AAC Asn	TGC Cys	AAT Asn 355	AAG Lys	ATT Ile	ATT Ile	1108
ATG Met	CTA Leu 360	TTC Phe	ACG Thr	GAT Asp	GGA Gly	GGA Gly 365	GAA Glu	GAG Glu	AGA Arg	GCC Ala	CAG Gln 370	GAG Glu	ATA Ile	TTT	AAC Asn	1156
AAA	TAC	AAT	AAA	GAT	AAA	AAA	GTA	CGT	GTA	TTC	AGG	TTT	TCA	GTT	GGT	1204

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Lys 375	Tyr	Asn	Lys	Asp	Lys 380		Val	Arg	Val	Phe 385		Phe	Ser	Val	Gly 390	
CAA Gln	CAC	AAT Asn	TAT	GAG Glu 395	AGA Arg	GGA Gly	CCT Pro	ATT Ile	CAG Gln 400	TGG Trp	ATG Met	GCC Ala	TGT Cys	GAA Glu 405	AAC Asn	1252
AAA Lys	GGT Gly	TAT	TAT Tyr 410	Tyr	GAA Glu	ATT Ile	CCT Pro	TCC Ser 415	ATT Ile	GGT Gly	GCA Ala	ATA Ile	AGA Arg 420	ATC Ile	AAT Asn	1300
ACT Thr	CAG Gln	GAA Glu 425	Tyr	TTG Leu	GAT Asp	GTT Val	TTG Leu 430	GGA Gly	AGA Arg	CCA Pro	ATG Met	GTT Val 435	TTA Leu	GCA Ala	GGA Gly	1348
GAC Asp	AAA Lys 440	GCT Ala	AAG Lys	CAA Gln	GTC Val	CAA Gln 445	TGG Trp	ACA Thr	AAT Asn	GTG Val	TAC Tyr 450	CTG Leu	GAT Asp	GCA Ala	TTG Leu	1396
GAA Glu 455	CTG Leu	GGA Gly	CTT Leu	GTC Val	ATT Ile 460	ACT Thr	GGA Gly	ACT Thr	CTT Leu	CCG Pro 465	GTC Val	TTC Phe	AAC Asn	ATA Ile	ACC Thr 470	1444
GGC Gly	CAA Gln	TTT Phe	GAA Glu	AAT Asn 475	AAG Lys	ACA Thr	AAC Asn	TTA Leu	AAG Lys 480	AAC Asn	CAG Gln	CTG Leu	ATT Ile	CTT Leu 485	GGT Gly	1492
GTG Val	ATG Met	GGA Gly	GTA Val 490	GAT Asp	GTG Val	TCT Ser	TTG Leu	GAA Glu 495	GAT Asp	ATT Ile	AAA Lys	AGA Arg	CTG Leu 500	ACA Thr	CCA Pro	1540
CGT Arg	TTT Phe	ACA Thr 505	CTG Leu	TGC Cys	CCC Pro	AAT Asn	GGG Gly 510	TAT Tyr	TAC Tyr	TTT Phe	GCA Ala	ATC Ile 515	GAT Asp	CCT Pro	AAT Asn	1588
GGT Gly	TAT Tyr 520	GTT Val	TTA Leu	TTA Leu	CAT His	CCA Pro 525	AAT Asn	CTT Leu	CAG Gln	CCA Pro	AAG Lys 530	AAC Asn	CCC Pro	AAA Lys	TCT Ser	1636
CAG Gln 535	GAG Glu	CCA Pro	GTA Val	ACA Thr	TTG Leu 540	GAT Asp	TTC Phe	CTT Leu	GAT Asp	GCA Ala 545	GAG Glu	TTA Leu	GAG Glu	AAT Asn	GAT Asp 550	1684
ATT Ile	AAA Lys	GTG Val	GAG Glu	ATT Ile 555	CGA Arg	AAT Asn	AAG Lys	ATG Met	ATT Ile 560	GAT Asp	GGG Gly	GAA Glu	AGT Ser	GGA Gly 565	GAA Glu	1732
AAA Lys	ACA Thr	TTC Phe	AGA Arg 570	ACT Thr	CTG Leu	GTT Val	AAA Lys	TCT Ser 575	CAA Gln	GAT Asp	GAG Glu	AGA Arg	TAT Tyr 580	ATT Ile	GAC Asp	1780
AAA Lys	GGA Gly	AAC Asn 585	AGG Arg	ACA Thr	TAC Tyr	ACA Thr	TGG Trp 590	ACA Thr	CCT Pro	GTC Val	AAT Asn	GGC Gly 595	ACA Thr	GAT Asp	TAC Tyr	1828
AGT	TTG	GCC	TTG	GTA	ATT	CCA	ACC	TAC	AGT	TTT	TAC	TAT	ATA	AAA	GCC	1876

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Ser	Leu 600	Ala	Leu	Val	Leu	Pro 605	Thr	Tyr	Ser	Phe	Tyr 610	Tyr	Ile	Lys	Ala		
AAA Lys 615	CTA Leu	GAA Glu	GAG Glu	ACA Thr	ATA Ile 620	ACT Thr	CAG Gln	GCC Ala	AGA Arg	TAT Tyr 625	TCG Ser	GAA Glu	ACC Thr	CTG Leu	AAG Lys 630		1924
CCA Pro	GAT Asp	AAT Asn	TTT Phe	GAA Glu 635	GAA Glu	TCT Ser	GGC Gly	TAT Tyr	ACA Thr 640	TTC Phe	ATA Ile	GCA Ala	CCA Pro	AGA Arg 645	GAT Asp	:	1972
TAC Tyr	TGC Cys	AAT Asn	GAC Asp 650	CTG Leu	AAA Lys	ATA Ile	TCG Ser	GAT Asp 655	AAT Asn	AAC Asn	ACT Thr	GAA Glu	TTT Phe 660	CTT Leu	TTA Leu	:	2020
AAT Asn	TTC Phe	AAC Asn 665	GAG Glu	TTT Phe	ATT Ile	GAT Asp	AGA Arg 670	AAA Lys	ACT Thr	CCA Pro	AAC Asn	AAC Asn 675	CCA Pro	TCA Ser	TGT Cys	:	2068
AAC Asn	GCG Ala 680	GAT Asp	TTG Leu	ATT Ile	AAT Asn	AGA Arg 685	GTC Val	TTG Leu	CTT Leu	GAT Asp	GCA Ala 690	GGC Gly	TTT Phe	ACA Thr	AAT Asn	:	2116
GAA Glu 695	CTT Leu	GTC Val	CAA Gln	AAT Asn	TAC Tyr 700	TGG Trp	AGT Ser	AAG Lys	CAG Gln	AAA Lys 705	AAT Asn	ATC Ile	AAG Lys	GGA Gly	GTG Val 710	;	2164
AAA Lys	GCA Ala	CGA Arg	TTT Phe	GTT Val 715	GTG Val	ACT Thr	GAT Asp	GGT Gly	GGG Gly 720	ATT Ile	ACC Thr	AGA Arg	GTT Val	TAT Tyr 725	CCC Pro		2212
AAA Lys	GAG Glu	GCT Ala	GGA Gly 730	GAA Glu	AAT Asn	TGG Trp	CAA Gln	GAA Glu 735	AAC Asn	CCA Pro	GAG Glu	ACA Thr	TAT Tyr 740	GAG Glu	GAC Asp		2260
AGC Ser	TTC Phe	TAT Tyr 745	AAA Lys	AGG Arg	AGC Ser	CTA Leu	GAT Asp 750	AAT Asn	GAT Asp	AAC Asn	TAT Tyr	GTT Val 755	TTC Phe	ACT Thr	GCT Ala		2308
CCC Pro	TAC Tyr 760	Phe	AAC Asn	AAA Lys	AGT Ser	GGA Gly 765	CCT Pro	GGT Gly	GCC Ala	TAT Tyr	GAA Glu 770	TCG Ser	GGC Gly	ATT Ile	ATG Met		2356
GTA Val 775	Ser	AAA Lys	GCT Ala	GTA Val	GAA Glu 780	ATA Ile	TAT Tyr	ATT Ile	CAA Gln	GGG Gly 785	AAA Lys	CTT Leu	CTT Leu	AAA Lys	CCT Pro 790	• 0	2404
GCA Ala	GTT Val	GTT Val	GGA Gly	ATT Ile 795	Lys	ATT Ile	GAT Asp	GTA Val	AAT Asn 800	Ser	TGG Trp	ATA Ile	GAG Glu	AAT Asn 805	FILE		2452
ACC	AAA Lys	ACC Thr	TCA Ser 810	Ile	AGA Arg	GAT Asp	CCG Pro	TGT Cys 815	Ala	GGT Gly	CCA Pro	GTT Val	TGT Cys 820	Asp	TGC Cys		2500
AAA	AGA	AAC	AGT	GAC	GTA	ATG	GAT	TGI	GTG	TTA	CTG	GAT	GAT	GGT	GGG		2548

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Lys	Arg	825	Ser	Asp	Va]	. Met	Asp 830		Val	. Ile	e Lev	Asp 835		Gly	gly	
TT1 Phe	CTT Leu 840	ren	ATC Met	GCA Ala	AAT Asn	CAT His 845	Asp	GAI Asp	TAI Tyr	ACI Thr	AAT Asn 850	Gln	ATT	GGA Gly	AGA Arg	2596
TTI Phe 855	Pile	GGA Gly	GAG Glu	ATI	GAT Asp 860	Pro	AGC Ser	TTG Leu	ATG Met	AGA Arg 865	His	CTG Leu	GTT Val	AAT Asn	ATA Ile 870	2644
TCA Ser	GTT Val	TAT	GCT Ala	TTT Phe 875	Asn	AAA Lys	TCT Ser	TAT	GAT Asp 880	Tyr	CAG Gln	TCA Ser	GTA Val	TGT Cys 885	GAG Glu	2692
CCC	GGT Gly	GCT Ala	GCA Ala 890	Pro	AAA Lys	CAA Gln	GGA Gly	GCA Ala 895	GGA Gly	CAT	CGC Arg	TCA Ser	GCA Ala 900	TAT	GTG Val	2740
CCA Pro	TCA Ser	GTA Val 905	GCA Ala	GAC Asp	ATA Ile	TTA Leu	CAA Gln 910	ATT Ile	GGC Gly	TGG Trp	TGG Trp	GCC Ala 915	ACT Thr	GCT Ala	GCT Ala	2788
GCC Ala	TGG Trp 920	TCT Ser	ATT Ile	CTA Leu	CAG Gln	CAG Gln 925	TTT Phe	CTC Leu	TTG Leu	AGT Ser	TTG Leu 930	ACC Thr	TTT Phe	CCA Pro	CGA Arg	2836
CTC Leu 935	CTT Leu	GAG Glu	GCA Ala	GTT Val	GAG Glu 940	ATG Met	GAG Glu	GAT Asp	GAT Asp	GAC Asp 945	TTC Phe	ACG Thr	GCC Ala	TCC Ser	CTG Leu 950	2884
TCC Ser	AAG Lys	CAG Gln	AGC Ser	TGC Cys 955	ATT Ile	ACT Thr	GAA Glu	CAA Gln	ACC Thr 960	CAG Gln	TAT Tyr	TTC Phe	TTC Phe	GAT Asp 965	AAC Asn	2932
GAC Asp	AGT Ser	AAA Lys	TCA Ser 970	TTC Phe	AGT Ser	GGT Gly	GTA Val	TTA Leu 975	GAC Asp	TGT Cys	GGA Gly	AAC Asn	TGT Cys 980	TCC Ser	AGA Arg	2980
ATC Ile	TTT Phe	CAT His 985	GGA Gly	GAA Glu	AAG Lys	CTT Leu	ATG Met 990	AAC Asn	ACC Thr	AAC Asn	TTA Leu	ATA Ile 995	TTC Phe	ATA Ile	ATG Met	3028
GTT Val	GAG Glu 1000	Ser	AAA Lys	GGG Gly	ACA Thr	TGT Cys 1005	Pro	TGT Cys	GAC Asp	ACA Thr	CGA Arg 1010	Leu	CTC Leu	ATA Ile	CAA Gln	3076
GCG Ala 1015	GAG Glu	CAG Gln	ACT Thr	TCT Ser	GAC Asp 1020	Gly	CCA Pro	AAT Asn	CCT Pro	TGT Cys 1025	Asp	ATG Met	GTT Val	AAG Lys	CAA Gln 1030	3124
CCT Pro	AGA Arg	TAC Tyr	CGA Arg	AAA Lys 1035	Gly	CCT Pro	GAT Asp	Val	TGC Cys 1040	Phe	GAT Asp	AAC Asn	Asn	GTC Val 1045	Leu	3172
GAG	GAT	TAT	ACT	GAC	TGT	GGT	GGT	GTT	TCT	GGA	TTA	AAT	ccc	TCC	CTG	3220

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Glu Asp Tyr Thr Asp Cys Gly Gly Val Ser Gly Leu Asn Pro Ser Leu 1050 1055 1060	
TGG TAT ATC ATT GGA ATC CAG TTT CTA CTA CTT TGG CTG GTA TCT GGC Trp Tyr Ile Ile Gly Ile Gln Phe Leu Leu Trp Leu Val Ser Gly 1065 1070 1075	3268
AGC ACA CAC CGG CTG TTA TGACCTTCTA AAAACCAAAT CTGCATAGTT Ser Thr His Arg Leu Leu 1080 108	3316
AAACTCCAGA CCCTGCCAAA ACATGAGCCC TGCCCTCAAT TACAGTAACG TAGGGTCAGC	3376
TATAAAATCA GACAAACATT AGCTGGGCCT GTTCCATGGC ATAACACTAA GGCGCAGACT	3436
CCTAAGGCAC CCACTGGCTG CATGTCAGGG TGTCAGATCC TTAAACGTGT GTGAATGCTG	3496
CATCATCTAT GTGTAACATC AAAGCAAAAT CCTATACGTG TCCTCTATTG GAAAATTTGG	3556
GCGTTTGTTG TTGCATTGTT GGT	3579
(2) INFORMATION FOR SEQ ID NO:33:	
(A) LENGTH: 1681 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear (ii) MOLECULE TYPE: DNA (genomic) (ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 11437 (D) OTHER INFORMATION: /standard_name= "Beta-1-1" (ix) FEATURE: (A) NAME/KEY: 3'UTR (B) LOCATION: 14351681	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:33:	
ATG GTC CAG AAG ACC AGC ATG TCC CGG GGC CCT TAC CCA CCC TCC CAG Met Val Gln Lys Thr Ser Met Ser Arg Gly Pro Tyr Pro Pro Ser Gln 1 5	4.6
GAG ATC CCC ATG GAG GTC TTC GAC CCC AGC CCG CAG GGC AAA TAC AGC Glu Ile Pro Met Glu Val Phe Asp Pro Ser Pro Gln Gly Lys Tyr Ser 20 25 30	96
AAG AGG AAA GGG CGA TTC AAA CGG TCA GAT GGG AGC ACG TCC TCG GAT Lys Arg Lys Gly Arg Phe Lys Arg Ser Asp Gly Ser Thr Ser Ser Asp	144

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ACC	ACA Thr	sei	AAC Asr	AGC Ser	TTI Phe	GTC Val	Arg	CAG Glr	GGC Gly	TCA Ser	GCG Ala	ı Glı	TC(TAC Ty	C ACC	192
AGC Ser 65	MLG	Pro	A TCA Ser	GAC Asp	TCT Ser 70	Asp	GTA Val	TCI Ser	CTG Leu	GAG Glu 75	Glu	GAC Asp	CGC Arg	GAZ J Glu	A GCC Ala 80	240
TTA Leu	AGG Arg	AAG Lys	GAA Glu	GCA Ala 85	GIU	CGC Arg	CAG Gln	GCA Ala	TTA Leu 90	Ala	CAG Gln	CTC Leu	GAG Glu	AAC Lys	GCC Ala	288
AAG Lys	ACC Thr	AAG Lys	CCA Pro 100	val	GCA Ala	TTT Phe	GCT Ala	GTG Val 105	Arg	ACA Thr	AAT Asn	GTT Val	GGC Gly 110	Tyr	AAT Asn	336
CCG Pro	TCT Ser	CCA Pro 115	GIA	GAT Asp	GAG Glu	GTG Val	CCT Pro 120	GTG Val	CAG Gln	GGA Gly	GTG Val	GCC Ala 125	ATC	ACC	TTC Phe	384
GIU	130	ьуs	Asp	Pne	Leu	H1S	Ile	Lys	Glu	Lys	Tyr 140	Asn	Asn	Asp		432
TGG Trp 145	ATC Ile	GGG	CGG Arg	CTG Leu	GTG Val 150	AAG Lys	GAG Glu	GGC Gly	TGT Cys	GAG Glu 155	GTT Val	GGC Gly	TTC Phe	ATT Ile	CCC Pro 160	480
AGC Ser	CCC Pro	GTC Val	AAA Lys	CTG Leu 165	GAC Asp	AGC Ser	CTT Leu	CGC Arg	CTG Leu 170	CTG Leu	CAG Gln	GAA Glu	CAG Gln	AAG Lys 175	CTG Leu	528
Arg	GIII	ASI	CGC Arg 180	ren	GIY	Ser	ser	Lys 185	Ser	Gly	Asp	Asn	Ser 190	Ser	Ser	576
ser	ren	195	GAT Asp	Val	Val	Thr	Gly 200	Thr	Arg	Arg	Pro	Thr 205	Pro	Pro	Ala	624
Ser	GGT Gly 210	AAT Asn	GAA Glu	ATG Met	ACT Thr	AAC Asn 215	TTA Leu	GCC Ala	TTT Phe	Glu	CTA Leu 220	GAC Asp	CCC Pro	CTA Leu	GAG Glu	672
TTA Leu 225	GAG Glu	GAG Glu	GAA Glu	GAG Glu	GCT Ala 230	GAG Glu	CTT Leu	GGT Gly	Glu	CAG Gln 235	AGT Ser	GGC Gly	TCT Ser	GCC Ala	AAG Lys 240	720
ACT Thr	AGT Ser	GTT Val	AGC Ser	AGT Ser 245	GTC Val	ACC . Thr	ACC Thr	Pro	CCA Pro 250	CCC Pro	CAT His	GGC Gly	Lys	CGC Arg 255	ATC Ile	768
CCC Pro	TTC Phe	Phe	AAG Lys 260	AAG Lys	ACA Thr	GAG Glu	His '	GTG Val 265	CCC Pro	CCC '	TAT Tyr	Asp	GTG Val 270	GTG Val	CCT Pro	816

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TCC ATG A Ser Met A 2	GG CCC rg Pro	ATC A	ATC (CTG Leu	GTG Val 280	GGA Gly	CCG Pro	TCG Ser	CTC Leu	AAG Lys 285	GGC Gly	TAC Tyr	GAG Glu	864
GTT ACA G Val Thr A 290	AC ATG	ATG (3ln I	AAA Lys 295	GCT Ala	TTA Leu	TTT Phe	GAC Asp	TTC Phe 300	TTG Leu	AAG Lys	CAT His	CGG Arg	912
TTT GAT G Phe Asp G 305	GC AGG ly Arg	Ile S	rcc A Ser I 310	ATC Ile	ACT Thr	CGT Arg	GTG Val	ACG Thr 315	GCA Ala	GAT Asp	ATT Ile	TCC Ser	CTG Leu 320	960
GCT AAG C Ala Lys A	GC TCA rg Ser	GTT (Val I 325	CTC / Leu /	AAC Asn	AAC Asn	CCC Pro	AGC Ser 330	AAA Lys	CAC His	ATC Ile	ATC Ile	ATT Ile 335	GAG Glu	1008
CGC TCC A	AC ACA sn Thr 340	CGC T	rcc A Ser S	AGC Ser	CTG Leu	GCT Ala 345	GAG Glu	GTG Val	CAG Gln	AGT Ser	GAA Glu 350	ATC Ile	GAG Glu	1056
CGA ATC T Arg Ile P 3	TC GAG he Glu 55	CTG (GCC (Ala <i>l</i>	CGG Arg	ACC Thr 360	CTT Leu	CAG Gln	TTG Leu	GTC Val	GCT Ala 365	CTG Leu	GAT Asp	GCT Ala	1104
GAC ACC A Asp Thr I 370	TC AAT le Asn	CAC (Pro 1	GCC Ala 375	CAG Gln	CTG Leu	TCC Ser	AAG Lys	ACC Thr 380	TCG Ser	CTG Leu	GCC Ala	CCC Pro	1152
ATC ATT G Ile Ile V 385	TT TAC	Ile I	AAG 1 Lys : 390	ATC Ile	ACC Thr	TCT Ser	CCC Pro	AAG Lys 395	GTA Val	CTT Leu	CAA Gln	AGG Arg	CTC Leu 400	1200
ATC AAG T Ile Lys S	CC CGA Ser Arg	GGA 1 Gly 1 405	AAG : Lys :	TCT Ser	CAG Gln	TCC Ser	AAA Lys 410	CAC His	CTC Leu	AAT Asn	GTC Val	CAA Gln 415	ATA Ile	1248
GCG GCC T Ala Ala S	CG GAA Ser Glu 420	AAG (Lys 1	CTG (Leu <i>i</i>	GCA Ala	CAG Gln	TGC Cys 425	CCC Pro	CCT Pro	GAA Glu	ATG Met	TTT Phe 430	GAC Asp	ATC Ile	1296
ATC CTG G Ile Leu A 4	AT GAG Sp Glu	AAC (Asn (CAA '	TTG Leu	GAG Glu 440	GAT Asp	GCC Ala	TGC Cys	GAG Glu	CAT His 445	CTG Leu	GCG Ala	GAG Glu	1344
TAC TTG G Tyr Leu G 450	SAA GCC Slu Ala	TAT Tyr	Trp :	AAG Lys 455	GCC Ala	ACA Thr	CAC His	CCG Pro	CCC Pro 460	AGC Ser	AGC Ser	ACG Thr	CCA Pro	1392
CCC AAT C Pro Asn F 465	CCG CTG Pro Leu	Leu :	AAC Asn 470	CGC Arg	ACC Thr	ATG Met	GCT Ala	ACC Thr 475	GCA Ala	GCC Ala	CTG Leu	GCT Ala		1437
GCCAGCCCT	rg cccc	TGTCT	C CA	ACC:	rcca(G GT	ACAG	GTGC	TCA	CCTC	GCT	CAGG.	AGAAAC	1497
CTCGGCTTC	CT GGGG	CGGGC	T GG	AGT	CCTC	A CA	GCGG	GGCA	GTG	TGGT	GCC	CCAG	GAGCAG	1557

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GAA	CATO	CCA	TGT	GTG	GC (CCCI	rgcco	G T	CTTCC	CTC	TGC	CTCTC	GGG	TCG	AACT	GG 161
AGI	GCAG	GGA	ACAT	GGAG	GA C	GAAG	GGAZ	G AG	CTT	TTTAT	TGT	AAA?	AAA	TAAC	ATGA	GC 167
GGC	'A															168
(2)	INF	'ORMA	TION	FOR	SEC	ID	NO:3	4:								
		(A) I B) T C) S D) T	ENGT YPE : TRAN OPOL	H: 1 nuc DEDN OGY:	CTER 526 leic ESS: lin	base aci dou ear	pai d ble								
	(ii) MO	LECU	LE I	YPE:	DNA	. (ge	nomi	.c)							
	(ix	(A) N B) L	AME/	: NOI	CDS 1 ORMA	651	: /s	tand	ard_	name	= "B	eta-	1-4"		
	(xi) SE	QUEN	CE D	ESCR	IPTI	ON:	SEQ	ID N	0:34	:					
ATG Met 1	GTC Val	CAG Gln	AAG Lys	ACC Thr 5	AGC Ser	ATG Met	TCC Ser	CGG Arg	GGC Gly 10	Pro	TAC Tyr	CCA Pro	CCC Pro	TCC Ser 15	CAG Gln	4.8
GAG Glu	ATC Ile	CCC Pro	ATG Met 20	GAG Glu	GTC Val	TTC Phe	GAC Asp	CCC Pro 25	AGC Ser	CCG Pro	CAG Gln	GGC Gly	AAA Lys 30	TAC Tyr	AGC Ser	96
AAG Lys	AGG Arg	AAA Lys 35	GGG Gly	CGA Arg	TTC Phe	AAA Lys	CGG Arg 40	TCA Ser	GAT Asp	GGG Gly	AGC Ser	ACG Thr 45	TCC Ser	TCG Ser	GAT Asp	144
ACC Thr	ACA Thr 50	TCC Ser	AAC Asn	AGC Ser	TTT Phe	GTC Val 55	CGC Arg	CAG Gln	GGC Gly	TCA Ser	GCG Ala 60	GAG Glu	TCC Ser	TAC Tyr	ACC Thr	192
AGC Ser 65	CGT Arg	CCA Pro	TCA Ser	GAC Asp	TCT Ser 70	GAT Asp	GTA Val	TCT Ser	CTG Leu	GAG Glu 75	GAG Glu	GAC Asp	CGG Arg	GAA Glu	GCC Ala 80	240
TTA Leu	AGG Arg	Lys	Glu	Ala	Glu	CGC Arg	Gln	Ala	Leu	Ala	Gln	Leu	Glu	Lys	Ala	288
AAG Lys	ACC Thr	AAG Lys	CCA Pro 100	GTG Val	GCA Ala	TTT Phe	GCT Ala	GTG Val 105	CGG Arg	ACA Thr	AAT Asn	GTT Val	GGC Gly 110	TAC Tyr	AAT Asn	336
CCG Pro	TCT Ser	CCA Pro 115	GGG Gly	GAT Asp	GAG Glu	GTG Val	CCT Pro 120	GTG Val	CAG Gln	GGA Gly	GTG Val	GCC Ala 125	ATC Ile	ACC Thr	TTC Phe	. 384

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GAG (CCC Pro 130	AAA Lys	GAC Asp	TTC Phe	CTG Leu	CAC His 135	ATC Ile	AAG Lys	GAG Glu	AAA Lys	TAC Tyr 140	AAT Asn	TAA neA	GAC Asp	TGG Trp	432
TGG 7 Trp 145	ATC Ile	GGG Gly	CGG Arg	CTG Leu	GTG Val 150	AAG Lys	GAG Glu	GGC Gly	TGT Cys	GAG Glu 155	GTT Val	GGC Gly	TTC Phe	ATT Ile	CCC Pro 160	480
AGC Ser	CCC Pro	GTC Val	AAA Lys	CTG Leu 165	GAC Asp	AGC Ser	CTT Leu	CGC Arg	CTG Leu 170	CTG Leu	CAG Gln	GAA Glu	CAG Gln	AAG Lys 175	CTG Leu	528
CGC Arg	CAG Gln	AAC Asn	CGC Arg 180	CTC Leu	GGC Gly	TCC Ser	AGC Ser	AAA Lys 185	TCA Ser	GGC Gly	GAT Asp	AAC Asn	TCC Ser 190	AGT Ser	TCC Ser	576
AGT Ser	CTG Leu	GGA Gly 195	GAT Asp	GTG Val	GTG Val	ACT Thr	GGC Gly 200	ACC Thr	CGC Arg	CGC Arg	CCC Pro	ACA Thr 205	CCC Pro	CCT Pro	GCC Ala	624
Ser .	GAC Asp 210	AGA Arg	GCA Ala	TGT Cys	GCC Ala	CCC Pro 215	CTA Leu	TGA	CGTG	STG (CCTT	CCAT	GA G	GCCC	ATCAT	678
CCTG	GTG	GA (CCGT	CGCT	CA AC	GGC:	racg:	A GG	TTAC	AGAC	ATG	ATGC	AGA :	AAGC'	TTATT	738
TGAC	TTC	TG I	AAGC	ATCG	ST T	rgato	GCA	G GA	rctc	CATC	ACT	CGTG'	TGA	CGGC	AGATAT	798
TTCC	CTG	CT Z	AAGÇ	GCTC	AG T	CTC	AACA	A CC	CCAG	CAAA	CAC	ATCA'	TCA '	TTGA	GCGCTC	858
CAAC	ACA	CGC :	TCCA	GCCT	GG C	rgago	STGC	A GA	GTGA.	AATC	GAG	CGAA'	TCT	TCGA	GCTGGC	918
CCGG	ACC	TT (CAGT'	rggr	CG C	rctg	GATG	C TG	ACAC	CATC	AAT	CACC	CAG	CCCA	GCTGTC	978
CAAG	ACC:	rcg (CTGG	cccc	CA TO	CATT	STTT	A CA	rcaa(GATC	ACC'	rctc	CCA .	AGGT	ACTTCA	1038
AAGG	CTC	ATC :	AAGT	CCCG	AG G	AAAG'	rctc	A GT	CCAA	ACAC	CTC	AATG'	TCC .	TAAA	AGCGGC	1098
CTCG	GAA	AAG (CTGG	CACA	GT G	cccc	CCTG	AA A	TGTT'	TGAC	ATC	ATCC	TGG .	ATGA	GAACCA	1158
ATTG	GAG	GAT (GCCT	GCGA	GC A	rctg	GCGG	A GT	ACTT	GGAA	GCC'	TATT	GGA .	AGGC	CACACA	1218
CCCG	ccc	AGC .	AGCA	CGCC	AC C	CAAT	CCGC'	T GC	TGAA	CCGC	ACC.	ATGG	CTA	CCGC.	AGCCCT	1278
GGCT	GCC	AGC	CCTG	CCCC	TG T	CTCC	AACC'	T CC.	AGGT.	ACAG	GTG	CTCA	CCT	CGCT	CAGGAG	1338
AAAC	CTC	GGC '	TTCT	GGGG	CG G	GCTG	GAGT	C CT	CACA	GCGG	GGC	AGTG	TGG	TGCC	CCAGGA	1398
GCAG	GAA	CAT	GCCA	TGTA	GT G	GGCG	CCCT	G CC	CGTC	TTCC	CTC	CTGC	TCT	GGGG	TCGGAA	1458
CTGG	AGT	GCA	GGGA	ACAT	GG A	GGAG	GAAG	G GA	AGAG	CTTT	ATT	TTGT	AAA	AAAA	TAAGAT	1518
GAGO	GGC	A														1526

(2) INFORMATION FOR SEQ ID NO:35:

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	(i) s	(C)	LENG TYPE STRA	CHAR TH: : nu NDED LOGY	1393 clei NESS	bas c ac : do	e pa id uble	irs							
	(i	i) M	OLEC	ULE	TYPE	: DN	A (g	enom	ic)							
	(i	x) F	(B)	NAME LOCA	/KEY TION R IN	: 1.	.660	N: /:	stan	dard	_nam	e= "]	Beta	-1-5	11	
			EQUE										•			
ATC Met	GT(Val	C CAC	G AAG n Lys	G ACC	- 261	ATC Met	TC(C CG(G GGG G Gly 10	Pro	TAC Ty:	C CCI	A CCC	TCC Ser	CAG Gln	48
GA0 Glu	ATO	C CCC	ATO Met		GTC Val	TTC Phe	GAC Asp	C CCC Pro 25	Ser	CCC Pro	G CAC	G GGC	AAA Lys	Туз	AGC Ser	96
AAG Lys	AGG Arg	AAA Lys 35	, Gry	CGA Arg	TTC Phe	AAA Lys	CGG Arg	Ser	GAT Asp	GGG Gly	AGO Ser	ACG Thr	Ser	TCG Ser	GAT Asp	144
ACC	ACA Thr 50		AAC Asn	AGC Ser	TTT Phe	GTC Val 55	Arg	CAG Gln	GGC	TCA Ser	GCG Ala	Glu	TCC Ser	TAC	ACC	192
AGC Ser 65	CGT Arg	CCA Pro	TCA Ser	GAC Asp	TCT Ser 70	GAT Asp	GTA Val	TCT	CTG Leu	GAG Glu 75	GAG Glu	GAC Asp	CGG Arg	GAA Glu	GCC Ala 80	240
TTA Leu	AGG Arg	AAG Lys	GAA Glu	GCA Ala 85	GAG Glu	CGC Arg	CAG Gln	GCA Ala	TTA Leu 90	GCG Ala	CAG Gln	CTC Leu	GAG Glu	AAG Lys 95	GCC Ala	288
AAG Lys	ACC Thr	AAG Lys	CCA Pro 100	GTG Val	GCA Ala	TTT Phe	GCT Ala	GTG Val 105	CGG Arg	ACA Thr	AAT Asn	GTT Val	GGC Gly 110	TAC Tyr	AAT Asn	336
CCG Pro	TCT Ser	CCA Pro 115	GGG Gly	GAT Asp	GAG Glu	GTG Val	CCT Pro 120	GTG Val	Gln	Gly	Val	GCC Ala 125	Ile	ACC Thr	TTC Phe	384
GAG Glu	CCC Pro 130	AAA Lys	GAC Asp	TTC Phe	CTG Leu	CAC His 135	ATC Ile	AAG Lys	GAG Glu	AAA Lys	TAC Tyr 140	AAT Asn	AAT Asn	GAC Asp	TGG Trp	432
TGG Trp 145	ATC Ile	GGG Gly	CGG Arg	CTG Leu	GTG Val 150	AAG Lys	GAG Glu	GGC Gly	Cys	GAG Glu 155	GTT Val	GGC Gly	TTC Phe	ATT Ile	CCC Pro 160	480

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AGC Ser	CCC Pro	GTC Val	AAA Lys	CTG Leu 165	GAC Asp	AGC Ser	CTT Leu	CGC Arg	CTG Leu 170	CTG Leu	CAG Gln	GAA Glu	CAG Gln	AAG Lys 175	CTG Leu		528
CGC Arg	CAG Gln	AAC Asn	CGC Arg 180	CTC Leu	GGC Gly	TCC	AGC Ser	AAA Lys 185	TCA Ser	GGC Gly	GAT Asp	AAC Asn	TCC Ser 190	AGT Ser	TCC Ser	-	576
AGT Ser	CTG Leu	GGA Gly 195	GAT Asp	GTG Val	GTG Val	ACT Thr	GGC Gly 200	ACC Thr	CGC Arg	CGC Arg	CCC Pro	ACA Thr 205	CCC Pro	CCT Pro	GCC Ala		624
Ser	GGT Gly 210	TAC Tyr	AGA Arg	CAT His	GAT Asp	GCA Ala 215	GAA Glu	AGC Ser	TTT Phe	ATT Ile	TGA0	CTTC:	rtg :	AAGC	ATCGGT	•	677
TTGA	TGG	CAG	GATC'	rcca!	rċ A	CTCG'	rgtg:	A CG	GCAG:	TATA	TTC	CCTG	GCT .	AAGC	GCTCAG	}	737
TTCI	CAAC	CAA	cccc	AGCA	AA C	ACAT	CATC	A TT	GAGC	GCTC	CAA	CACA	CGC	TCCA	GCCTGG	;	797
CTGA	GGT	ЭCА	GAGT	GAAA'	TC G	AGCG.	AATC'	T TC	GAGC'	TGGC	CCG	GACC	CTT	CAGT	TGGTCG	}	857
CTCI	GGA:	rgc	TGAC	ACCA'	TC A	ATCA	CCCA	G CC	CAGC'	TGTC	CAA	GACC'	TCG	CTGG	CCCCCA	١	917
															CCCGAG		977
															CACAGI		1037
															GCGAGC		1097
															CGCCAC		1157
															CCCCT		1217
															GGGGC		1277
															TGTAG		1337
														GGGA			1393
333	٠٠٠٠																

(2) INFORMATION FOR SEQ ID NO:36:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 6725 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (ix) FEATURE:

 - (A) NAME/KEY: CDS
 (B) LOCATION: 226..6642
 - (D) OTHER INFORMATION: /standard_name= "Alpha-1C-2"

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	(x	i) S	EQUE	NCE :	DESC	RIPT	ON:	SEQ	ID	NO:3	6:						
CT	CGAG	GAGG	CAG	TAGT	GGA .	AAGG	AGCA	GT T	TTTG	GGGT	T TG	ATGC	CATA	ATG	GGAATC	A	60
GG	TAAT	CGTC	GGC	GGGG	AAG .	AAGA	AACG	CT G	CAGA	CCAC	G GC	TTCC	TCGA	ATC	TTGCGC	G 1	20
															CACATT		80
				GCTG													
												j	Met '	Val 1	Asn	2:	34
		5	. AL,	a we	- 1yı	10	Pro	GI	ı Gli	ı Ası	His	Glı	n Gly	y Sei	AAC Asn	28	32
20)			, ,,,,	25	AIG	HIE	Ale	ASI	30	Asr	1 Ala	a Asr	ı Ala	GCA Ala 35	33	30
GC0 Ala	GGG Gly	CTC Lev	GCC Ala	CCT Pro 40	GIU	CAC His	ATC Ile	CCC Pro	ACC Thr 45	Pro	GGG Gly	GCT Ala	GCC Ala	CTC Lev	TCG Ser	37	8
TGG	CAG Gln	GCG Ala	GCC Ala 55	116	GAC Asp	GCA Ala	GCC	CGG Arg 60	GIn	GCT Ala	Lys	CTG Leu	ATG Met	Gly	AGC Ser	42	6
GCT Ala	GGC	AAT Asn 70	ALA	ACC Thr	ATC Ile	TCC Ser	ACA Thr 75	val	AGC Ser	TCC Ser	ACG Thr	CAG Gln 80	Arg	AAG Lys	CGG Arg	47	4
CAG Gln	CAA Gln 85	- Y -	GGG Gly	AAA Lys	CCC Pro	AAG Lys 90	AAG Lys	CAG Gln	GGC Gly	AGC Ser	ACC Thr 95	ACG Thr	GCC Ala	ACA Thr	CGC Arg	52	2
CCG Pro 100	CCC Pro	CGA Arg	GCC Ala	CTG Leu	CTC Leu 105	TGC Cys	CTG Leu	ACC Thr	CTG Leu	AAG Lys 110	AAC Asn	CCC Pro	ATC Ile	CGG Arg	AGG Arg 115	57	0
GCC Ala	TGC Cys	ATC Ile	AGC Ser	ATT Ile 120	GTC Val	GAA Glu	TGG Trp	AAA Lys	CCA Pro 125	TTT Phe	GAA Glu	ATA Ile	ATT Ile	ATT Ile 130	TTA Leu	618	В
CTG Leu	ACT Thr	ATT Ile	TTT Phe 135	GCC Ala	AAT Asn	TGT Cys	GTG Val	GCC Ala 140	TTA Leu	GCG Ala	ATC Ile	TAT Tyr	ATT Ile 145	CCC Pro	TTT Phe	666	5
CCA Pro	GAA Glu	GAT Asp 150	GAT Asp	TCC Ser	AAC Asn	Ala	ACC Thr 155	AAT Asn	TCC Ser	AAC Asn	CTG Leu	GAA Glu 160	CGA Arg	GTG Val	GAA Glu	714	ļ
TYL	CTC Leu	Pne	CTC Leu	ATA Ile	TTE	TTT Phe	Thr	GTG Val	GAA Glu	GCG Ala	TTT Phe	TTA Leu	AAA Lys	GTA Val	ATC Ile	762	!

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GCC Ala 180	TAT Tyr	GGA Gly	CTC Leu	CTC Leu	TTT Phe 185	CAC His	CCC Pro	AAT Asn	GCC Ala	TAC Tyr 190	CTC Leu	CGC Arg	AAC Asn	GGC Gly	TGG Trp 195	810
AAC Asn	CTA Leu	CTA Leu	GAT Asp	TTT Phe 200	ATA Ile	ATT Ile	GTG Val	GTT Val	GTG Val 205	GGG Gly	CTT Leu	TTT Phe	AGT Ser	GCA Ala 210	ATT Ile	858
TTA Leu	GAA Glu	CAA Gln	GCA Ala 215	ACC Thr	AAA Lys	GCA Ala	GAT Asp	GGG Gly 220	GCA Ala	AAC Asn	GCT Ala	CTC Leu	GGA Gly 225	GGG Gly	AAA Lys	906
GGG Gly	GCC Ala	GGA Gly 230	TTT Phe	GAT Asp	GTG Val	AAG Lys	GCG Ala 235	CTG Leu	AGG Arg	GCC Ala	TTC Phe	CGC Arg 240	GTG Val	CTG Leu	CGC Arg	954
CCC Pro	CTG Leu 245	CGG Arg	CTG Leu	GTG Val	TCC Ser	GGA Gly 250	GTC Val	CCA Pro	AGT Ser	CTC Leu	CAG Gln 255	GTG Val	GTC Val	CTG Leu	AAT Asn	1002
TCC Ser 260	ATC Ile	ATC Ile	AAG Lys	GCC Ala	ATG Met 265	GTC Val	CCC Pro	CTG Leu	CTG Leu	CAC His 270	ATC Ile	GCC Ala	CTG Leu	CTT Leu	GTG Val 275	1050
CTG Leu	TTT Phe	GTC Val	ATC Ile	ATC Ile 280	ATC Ile	TAC Tyr	GCC Ala	ATC Ile	ATC Ile 285	GGC Gly	TTG Leu	GAG Glu	CTC Leu	TTC Phe 290	ATG Met	1098
GGG Gly	AAG Lys	ATG Met	CAC His 295	AAG Lys	ACC Thr	TGC Cys	TAC Tyr	AAC Asn 300	CAG Gln	GAG Glu	GGC Gly	ATA Ile	GCA Ala 305	GAT Asp	GTT Val	1146
CCA Pro	GCA Ala	GAA Glu 310	GAT Asp	GAC Asp	CCT Pro	TCC Ser	CCT Pro 315	TGT Cys	GCG Ala	CTG Leu	GAA Glu	ACG Thr 320	GGC Gly	CAC His	GGG	1194
CGG Arg	CAG Gln 325	Cys	CAG Gln	AAC Asn	GGC Gly	ACG Thr 330	GTG Val	TGC Cys	AAG Lys	CCC Pro	GGC Gly 335	Trp	GAT Asp	GGT Gly	CCC Pro	1242
AAG Lys 340	His	GGC Gly	ATC Ile	ACC Thr	AAC Asn 345	TTT Phe	GAC Asp	AAC Asn	TTT Phe	GCC Ala 350	Pne	GCC Ala	ATG Met	CTC Leu	ACG Thr 355	1290
GTG Val	TTC Phe	CAG Gln	TGC Cys	ATC Ile 360	Thr	ATG Met	GAG Glu	GGC Gly	TGG Trp 365	Thr	GAC Asp	GTG Val	CTG Leu	TAC Tyr 370	TGG Trp	1338
GTC Val	AAT Asn	GAT Asp	GCC Ala 375	. Val	GGA Gly	AGG Arg	GAC Asp	TGG Trp	PIC	TGG Trp	ATC Ile	TAT Tyr	TTT Phe 385		ACA Thr	1386
CTA Lev	ATC	ATC Ile	Ile	GGG Gly	TCA Ser	TTT Phe	TTT Phe	. val	CTI Lev	AAC Asn	TTC Lev	GTI Val 400	. Let	GGT Gly	GTG Val	1434

CT Le	T AG u Se: 40	C GG r Gl 5	A GA y Gl	G TT u Ph	T TC e Se	C AA r Ly: 41	SGI	G AG	G GA	G AA	G GC s Al 41	a Ly	G GC S Al	C CC	G GGA	1482
42	Ò		,	. <u>.</u>	42	5	тъ	s G11	n Gli	1 Let 430	o GI	u Gl	u As	p Le	C AAA u Lys 435	
	•			440	0	- 1111	. 611.	I Ale	445	a Asp) I16	≥ As]	p Pr	o Gl 45		
	•		45	5	- nor	GIL	GIU	460)	Arg	J AST	n Met	Se:	r Me 5	G CCC t Pro	1626
		470)		- 501	Val	475	1111	GIU	AST	vaı	480	i Gly	/ G1:	T GAC y Asp	1674
	485				. Cys	490	MIG	Arg	ren	Ala	495	Arg	, Ile	: Se:	C AAG	1722
500	•			9	505	110	Arg	Arg	тър	510	Arg	Phe	Cys	Arg	A AGG J Arg 515	1770
	-,-	• 9	7,14	520	val	ьуs	ser	Asn	Val 525	Phe	Tyr	Trp	Leu	Val 530		1818
			535	200	ASII	1111	neu	540	TTE	ATA	ser	Glu	His 545	Туг	AAC Asn	1866
		550	11p	Deu	Inr	GIU	555	GIn	Asp	Thr	Ala	Asn 560	Lys	Ala	CTG Leu	1914
	565	De u	FILE	1111	Ala	570	Met	Leu	Leu	Lys	Met 575	Tyr	Ser	Leu		1962
CTG Leu 580	CAG Gln	GCC Ala	TAC Tyr	TTC Phe	GTG Val 585	TCC Ser	CTC Leu	TTC Phe	AAC Asn	CGC Arg 590	TTT Phe	GAC Asp	TGC Cys	TTC Phe	GTC Val 595	2010
vui	Cys	GIY	GGC Gly	600	ren	GIU	Tnr	IIe	Leu 605	Val	Glu	Thr	Lys	Ile 610	Met	2058
TCC Ser	CCA Pro	CTG Leu	GGC Gly 615	ATC Ile	TCC Ser	GTG Val	Leu .	AGA Arg 620	TGC Cys	GTC Val	CGG Arg	Leu	CTG Leu 625	AGG Arg	ATT Ile	2106

					TAC Tyr												2154
TTG Leu	CTG Leu 645	AAC Asn	TCT Ser	GTG Val	CGC Arg	TCC Ser 650	ATC Ile	GCC Ala	TCC Ser	CTG Leu	CTC Leu 655	CTT Leu	CTC Leu	CTC Leu	TTC Phe	-	2202
					TTC Phe 665												2250
					GAG Glu												2298
TTC Phe	CCC Pro	CAG Gln	TCC Ser 695	CTC Leu	CTC Leu	ACT Thr	GTG Val	TTT Phe 700	CAG Gln	ATC Ile	CTG Leu	ACC Thr	GGG Gly 705	GAG Glu	GAC Asp		2346
TGG Trp	AAT Asn	TCG Ser 710	GTG Val	ATG Met	TAT Tyr	GAT Asp	GGG Gly 715	ATC Ile	ATG Met	GCT Ala	TAT Tyr	GGC Gly 720	GGC Gly	CCC Pro	TCT Ser		2394
Phe	Pro 725	Gly	Met	Leu	GTC Val	Cys 730	Ile	Tyr	Phe	Ile	Ile 735	Leu	Phe	Ile	Cys		2442
GGA Gly 740	AAC Asn	TAT Tyr	ATC Ile	CTA Leu	CTG Leu 745	AAT Asn	GTG Val	TTC Phe	TTG Leu	GCC Ala 750	ATT	GCT Ala	GTG Val	GAC Asp	AAC Asn 755		2490
Leu	Ala	Asp	Ala	Glu 760	AGC Ser	Leu	Thr	Ser	Ala 765	Gln	Lys	Glu	Glu	Glu 770	Glu		2538
Glu	Lys	Glu	Arg 775	Lys	AAG Lys	Leu	Ala	Arg 780	Thr	Ala	Ser	Pro	Glu 785	Lys	Lys		2586
Gln	Glu	Leu 790	Val	Glu	AAG Lys	Pro	Ala 795	Val	Gly	Glu	Ser	Lys 800	Glu	Glu	Lys		2634
ATT Ile	GAG Glu 805	CTG Leu	AAA Lys	TCC Ser	ATC Ile	ACG Thr 810	GCT Ala	GAC Asp	GGA Gly	GAG Glu	TCT Ser 815	CCA Pro	CCC Pro	GCC Ala	ACC Thr		2682
AAG Lys 820	Ile	AAC Asn	ATG Met	GAT Asp	GAC Asp 825	CTC Leu	CAG Gln	CCC Pro	AAT Asn	GAA Glu 830	AAT Asn	GAG Glu	GAT Asp	AAG Lys	AGC Ser 835		2730
CCC Pro	TAC Tyr	CCC Pro	AAC Asn	CCA Pro 840	GAA Glu	ACT Thr	ACA Thr	GGA Gly	GAA Glu 845	GAG Glu	GAT Asp	GAG Glu	GAG Glu	GAG Glu 850	CCA Pro		2778

GA0	ATG Met	CCI Pro	Val	. GIY	CCI Pro	CGC Arg	CCA Pro	CGA Arg 860	Pro	CTC Leu	TCT Ser	GAG Glu	CTI Leu 865	His	CTT Leu	2826
AAC Lys	GAA Glu	AAG Lys 870	Ala	GTG Val	Pro	ATG Met	CCA Pro 875	Glu	GCC	AGC Ser	GCG Ala	Phe	TTC Phe	ATC	TTC Phe	2874
AGC Ser	Ser 885	ASI	AAC Asn	AGG Arg	TTT Phe	CGC Arg 890	CTC	CAG Gln	TGC Cys	CAC	CGC Arg 895	ATT	GTC Val	AAT Asn	GAC Asp	2922
ACG Thr 900	116	TTC Phe	ACC Thr	AAC Asn	CTG Leu 905	ATC Ile	CTC Leu	TTC Phe	TTC Phe	ATT Ile 910	CTG Leu	CTC Leu	AGC Ser	AGC Ser	ATT Ile 915	2970
TCC Ser	CTG Leu	GCT Ala	GCT Ala	GAG Glu 920	Asp	CCG Pro	GTC Val	CAG Gln	CAC His 925	ACC Thr	TCC Ser	TTC Phe	AGG Arg	AAC Asn 930	CAT His	3018
ATT Ile	CTG Leu	TTT Phe	TAT Tyr 935	TTT Phe	GAT Asp	ATT Ile	GTT Val	TTT Phe 940	ACC Thr	ACC Thr	ATT Ile	TTC Phe	ACC Thr 945	ATT Ile	GAA Glu	3066
ATT Ile	GCT Ala	CTG Leu 950	AAG Lys	ATG Met	ACT Thr	GCT Ala	TAT Tyr 955	GGG Gly	GCT Ala	TTC Phe	TTG Leu	CAC His 960	AAG Lys	GGT Gly	TCT Ser	3114
TTC Phe	TGC Cys 965	CGG Arg	AAC Asn	TAC Tyr	TTC Phe	AAC Asn 970	ATC Ile	CTG Leu	GAC Asp	CTG Leu	CTG Leu 975	GTG Val	GTC Val	AGC Ser	GTG Val	3162
TCC Ser 980	CTC Leu	ATC Ile	TCC Ser	TTT Phe	GGC Gly 985	ATC Ile	CAG Gln	TCC Ser	AGT Ser	GCA Ala 990	ATC Ile	AAT Asn	GTC Val	GTG Val	AAG Lys 995	3210
ATC Ile	TTG Leu	CGA Arg	GTC Val	CTG Leu 1000	Arg	GTA Val	CTC Leu	AGG Arg	CCC Pro 1005	Leu	AGG Arg	GCC Ala	ATC Ile	AAC Asn 1010	Arg	3258
GCC Ala	AAG Lys	GGG Gly	CTA Leu 1015	Lys	CAT His	GTG Val	GTT Val	CAG Gln 1020	Cys	GTG Val	TTT Phe	GTC Val	GCC Ala 1025	Ile	CGG Arg	3306
ACC Thr	ATC Ile	GGG Gly 1030	Asn	ATC Ile	GTG Val	Ile	GTC Val 1035	Thr	ACC Thr	CTG Leu	CTG Leu	CAG Gln 1040	Phe	ATG Met	TTT Phe	3354
GCC Ala	TGC Cys 1045	Ile	GGG Gly	GTC Val	CAG Gln	CTC Leu 1050	Phe	AAG Lys	GGA Gly	Lys	CTG Leu 1055	TAC Tyr	ACC Thr	TGT Cys	TCA Ser	3402
GAC Asp 1060	Ser	TCC Ser	AAG Lys	Gln	ACA Thr 1065	Glu .	GCG Ala	GAA Glu	Cys	AAG Lys 1070	Gly	AAC Asn	TAC Tyr	Ile	ACG Thr 1075	3450

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					Val	GAC Asp				Ile					Trp	3498
_				Phe		TTT Phe			Val					Met		3546
			Val			TTC Phe		Gly					Leu			3594
		Asp				GAA Glu 1130	Asp					Tyr				3642
	Glu					TTC Phe					Ile					3690
_					Phe	GTG Val				Ile					Glu	3738
				Glu		AAG Lys			Glu					Gln		3786
			Glu			CTC Leu		Ala					Arg			3834
		Asn				TAC Tyr 1210	Lys					Val				3882
	Phe					TTC Phe					Leu					3930
CTG Leu	GCC Ala	ATG Met	CAG Gln	CAC His 124	Tyr	GGC Gly	CAG Gln	AGC Ser	TGC Cys 124	Leu	TTC Phe	AAA Lys	ATC Ile	GCC Ala 1250	Met	3978
AAC Asn	ATC Ile	CTC Leu	AAC Asn 125!	Met	CTC Leu	TTC Phe	ACT Thr	GGC Gly 1260	Leu	TTT Phe	ACC Thr	GTG Val	GAG Glu 126	Met	ATC Ile	4026
CTG Leu	AAG Lys	CTC Leu 1270	Ile	GCC Ala	TTC Phe	AAA Lys	CCC Pro 1275	Lys	CAC His	TAT Tyr	TTC Phe	TGT Cys 1280	Asp	GCA Ala	TGG Trp	4074
AAT Asn	ACA Thr 128	Phe	GAC Asp	GCC Ala	TTG Leu	ATT Ile 1290	Val	GTG Val	GGT Gly	AGC Ser	ATT Ile 129	Val	GAT Asp	ATA Ile	GCA Ala	4122

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ATC ACC Ile Thr 1300	GAG GTA AA Glu Val As	C CCA GCT n Pro Ala 1305	GAA CAT Glu His	ACC CAA Thr Gln 1310	Cys Ser	CCC TCT Pro Ser	ATG Met 1315	4170
AAC GCA Asn Ala	GAG GAA AA Glu Glu As 13	n Ser Arg	ATC TCC Ile Ser	ATC ACC Ile Thr 1325	TTC TTC	CGC CTG Arg Leu 133	Phe	4218
CGG GTC Arg Val	ATG CGT CT Met Arg Le 1335	G GTG AAG u Val Lys	CTG CTG Leu Leu 1340	Ser Arg	GGG GAG Gly Glu	GGC ATC Gly Ile 1345	CGG Arg	4266
ACG CTG Thr Leu	CTG TGG AC Leu Trp Th 1350	r Phe Ile	AAG TCC Lys Ser 1355	TTC CAG Phe Gln	GCC CTG Ala Leu 136	Pro Tyr	GTG Val	4314
GCC CTC Ala Leu 1365	CTG ATC GT Leu Ile Va	G ATG CTG l Met Leu 1370	Phe Phe	ATC TAC Ile Tyr	GCG GTG Ala Val 1375	ATC GGG Ile Gly	ATG Met	4362
CAG GTG Gln Val 1380	TTT GGG AA Phe Gly Ly	A ATT GCC s Ile Ala 1385	CTG AAT Leu Asn	GAT ACC Asp Thr 1390	Thr Glu	ATC AAC Ile Asn	CGG Arg 1395	4410
AAC AAC Asn Asn	AAC TTT CA Asn Phe Gl: 14	n Thr Phe	Pro Gln	GCC GTG Ala Val 1405	CTG CTC Leu Leu	CTC TTC Leu Phe 141	Arg	4458
TGT GCC Cys Ala	ACC GGG GAG Thr Gly Glo 1415	G GCC TGG	CAG GAC Gln Asp 1420	Ile Met	CTG GCC Leu Ala	TGC ATG Cys Met 1425	CCA Pro	4506
GGC AAG Gly Lys	AAG TGT GCC Lys Cys Ala 1430	a Pro Glu	TCC GAG Ser Glu 1435	CCC AGC Pro Ser	AAC AGC Asn Ser 144	Thr Glu	GGT Gly	4554
	CCC TGT GG Pro Cys Gly			Val Phe				4602
	CTC TGT GCC Leu Cys Ala				Phe Val			4650
	AAC TTT GAG Asn Phe Asp 148	Tyr Leu ?	Thr Arg .				Pro	4698
	CTG GAT GAG Leu Asp Glu 1495			Trp Ala				4746
Ala Lys	GGT CGT ATO Gly Arg Ile 1510	Lys His I				Leu Arg		4794

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	CTA GGT TTT GGG Leu Gly Phe Gly 1530			
TGC AAA CGC CTG Cys Lys Arg Leu 1540	GTC TCC ATG AAG Val Ser Met Ass 1545	ATG CCT CTG Met Pro Leu 155	Asn Ser Asp (GGG ACA 4890 Gly Thr 1555
	GCC ACC CTG TTT Ala Thr Leu Pho 1560		Arg Thr Ala I	
ATC AAA ACA GAA Ile Lys Thr Glu 157	GGG AAC CTA GAA Gly Asn Leu Glu 5	CAA GCC AAT Gln Ala Asn 1580	GAG GAG CTG C Glu Glu Leu A 1585	CGG GCG 4986 Arg Ala
	ATC TGG AAG CGG Ile Trp Lys Arg 159	Thr Ser Met		
	GCA GGT GAT GAT Ala Gly Asp Asp 1610			
	ATC CAG GAG TAG Ile Gln Glu Tyn 1625		Phe Lys Lys P	
GAG CAG GGC CTT Glu Gln Gly Leu	GTG GGC AAG CCC Val Gly Lys Pro 1640	TCC CAG AGG Ser Gln Arg 1645	Asn Ala Leu S	CCT CTG 5178 Ser Leu 1650
CAG GCT GGC TTG Gln Ala Gly Leu 165	CGC ACA CTG CAT Arg Thr Leu His 5	GAC ATC GGG Asp Ile Gly 1660	CCT GAG ATC (Pro Glu Ile A 1665	CGA CGG 5226 Arg Arg
GCC ATC TCT GGA Ala Ile Ser Gly 1670	GAT CTC ACC GCT Asp Leu Thr Ala 16	Glu Glu Glu	CTG GAC AAG C Leu Asp Lys A 1680	SCC ATG 5274 Ala Met
AAG GAG GCT GTG Lys Glu Ala Val 1685	TCC GCT GCT TC Ser Ala Ala Se 1690	GAA GAT GAC Glu Asp Asp	ATC TTC AGG A Ile Phe Arg A 1695	AGG GCC 5322 Arg Ala
GGT GGC CTG TTC Gly Gly Leu Phe 1700	GGC AAC CAC GTG Gly Asn His Va 1705	AGC TAC TAC Ser Tyr Tyr 171	Gln Ser Asp (GGC CGG 5370 Gly Arg 1715
AGC GCC TTC CCC Ser Ala Phe Pro	C CAG ACC TTC ACC Gln Thr Phe Th 1720	C ACT CAG CGC r Thr Gln Arg 1725	Pro Leu His	ATC AAC 5418 Ile Asn 1730
AAG GCG GGC AGC Lys Ala Gly Ser 173	: AGC CAG GGC GA : Ser Gln Gly As :5	C ACT GAG TCG Thr Glu Ser 1740	CCA TCC CAC (Pro Ser His (1745	GAG AAG 5466 Glu Lys

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CTG GTG GAC TO Leu Val Asp So 1750	CC ACC TTC ACer Thr Phe Th	C CCG AGC r Pro Ser 1755	AGC TAC TCG Ser Tyr Ser	TCC ACC GGC Ser Thr Gly 1760	TCC 5514 Ser
AAC GCC AAC A Asn Ala Asn I 1765	TC AAC AAC GC le Asn Asn Ala 17	a Asn Asn	ACC GCC CTG Thr Ala Leu 177!	Gly Arg Leu	CCT 5562 Pro
CGC CCC GCC GC Arg Pro Ala G 1780	GC TAC CCC AGG ly Tyr Pro Se 1785	ACG GTC Thr Val	AGC ACT GTG Ser Thr Val 1790	GAG GGC CAC Glu Gly His	GGG 5610 Gly 1795
CCC CCC TTG TO Pro Pro Leu Se	CC CCT GCC ATO Pro Ala Ile 1800	Arg Val	CAG GAG GTG Gln Glu Val 1805	GCG TGG AAG Ala Trp Lys 181	Leu
AGC TCC AAC AG Ser Ser Asn Al	GG TGC CAC TCC TG Cys His Sei 315	CGG GAG : Arg Glu 1820	Ser Gln Ala	GCC ATG GCG Ala Met Ala 1825	GGT 5706 Gly
CAG GAG GAG AC Gln Glu Glu Th 1830	CG TCT CAG GAT or Ser Gln Asp	GAG ACC S Glu Thr S 1835	TAT GAA GTG Tyr Glu Val	AAG ATG AAC Lys Met Asn 1840	CAT 5754 His
GAC ACG GAG GO Asp Thr Glu Al 1845	CC TGC AGT GAG a Cys Ser Glu 185	Pro Ser 1	CTG CTC TCC Leu Leu Ser 1855	Thr Glu Met	CTC 5802 Leu
TCC TAC CAG GA Ser Tyr Gln As 1860	T GAC GAA AAT p Asp Glu Asn 1865	CGG CAA (Arg Gln)	CTG ACG CTC Leu Thr Leu 1870	CCA GAG GAG Pro Glu Glu	GAC 5850 Asp 1875
AAG AGG GAC AT Lys Arg Asp Il	C CGG CAA TCT e Arg Gln Ser 1880	Pro Lys A	AGG GGT TTC Arg Gly Phe 1885	CTC CGC TCT Leu Arg Ser 1890	Ala
TCA CTA GGT CG Ser Leu Gly Ar 18	A AGG GCC TCC g Arg Ala Ser 95	TTC CAC (Phe His I 1900	CTG GAA TGT Leu Glu Cys	CTG AAG CGA Leu Lys Arg 1905	CAG 5946 Gln
AAG GAC CGA GG Lys Asp Arg Gl 1910	G GGA GAC ATC y Gly Asp Ile	TCT CAG A Ser Gln I 1915	Lys Thr Val	CTG CCC TTG Leu Pro Leu 1920	CAT 5994 His
CTG GTT CAT CA Leu Val His Hi 1925	T CAG GCA TTG s Gln Ala Leu 193	Ala Val A	GCA GGC CTG A Ala Gly Leu 1935	AGC CCC CTC Ser Pro Leu	CTC 6042 Leu
CAG AGA AGC CA Gln Arg Ser Hi 1940	T TCC CCT GCC s Ser Pro Ala 1945	TCA TTC C Ser Phe P	CCT AGG CCT 'Pro Arg Pro 1	Phe Ala Thr	CCA 6090 Pro 1955
CCA GCC ACA CC Pro Ala Thr Pro	T GGC AGC CGA o Gly Ser Arg 1960	Gly Trp P	CCC CCA CAG (Pro Pro Gln 1	CCC GTC CCC Pro Val Pro 1970	Thr

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	CGG Arg			Gly					Glu					Ser		6186
	TCC Ser		His					Ala					Gly			6234
	AGC Ser 2005	Ser					Val					Leu				6282
	CAG Gln					Gly					Gly					6330
	GTG Val				Leu					Leu					Gln	6378
	CCC Pro			Ile					Gln					Ala		6426
	ATG Met		Ile					Ser					Ile			6474
	GGC Gly 2085	Ala					Asn					Pro				6522
	AGG Arg					Asp					Glu					6570
	GTG Val				Gly					Glu					Ser	6618
	GTC Val			Ser			TAGT	regec	GC 1	rgcca	GATG	C GG	GCTI	rttt1	•	6669
TTAT	TTGI	TT C	AATO	TTCC	T AZ	TGGG	TTCC	TTI	CAG	AGT	GCCI	CACI	GT I	CTC	T	6725

(2) INFORMATION FOR SEQ ID NO:37:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2970 base pairs
 (B) TYPE: nucleic acid
 (C) STRANDEDNESS: double
 (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)

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(ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 502..2316
(D) OTHER INFORMATION: /standard_name= "Beta-2C"

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:37:

						•
CAGCAGCGTG	CTAAGAAGC	A GTCACATAA	A CAGCAGCAGG	AGTAGGCCTC	CTGCTTTTCA	60
AAAGCAGAGT	ACTGCAGGG	T CGCGAAATG	C AAGACACTCA	GATGTTTGAA	AATCTCCCGA	120
GTTGAGAATG	GCTACTGTA	A AAGCGTCAC	C AAGAAACTCT	GACGATCTGG	ACAGTCCTAA	180
CTCTGTGTTA	GCAATACTT	a CTTCCGGAA	A ATTAATGCTA	CTTCTTGTAG	ATTTTTGCAA	240
ATAGGAAACC	CCCTTGAAG	A AGATCTCAA	A TTACGCCCCC	CACCCCAAA	AAAAGACAAA	300
CAGGGGAGAA	CAAAGTTTT	G GCATGCCTG	C AGGAACGGTG	GCTTTTTTAG .	AAACTACCTA	360
GGAGGCAGAA	GCTAAGTGA	T TTGCTCATG	C CTCTTACCTG	GGAGTAGAAG	GTGGGAAGAA	420
ATGGACCGAG	GCTGTGACGA	A GAAGACAAG	G CACAGTGCAG	CTTGGTGAAG	CCACACGCTG	480
ACTGCGTTCT	GCCCCTCTT		TGC TGC GGG Cys Cys Gly 5			531
CGA GTA CG Arg Val Ar	G GTG TCC 1 g Val Ser 1 15	TAT GGT TCG Tyr Gly Ser	GCA GAC TCC Ala Asp Ser 20	TAC ACT AGC Tyr Thr Ser	CGT CCA Arg Pro 25	579
TCC GAT TC Ser Asp Se	C GAT GTA 1 r Asp Val 8 30	TCT CTG GAG Ser Leu Glu	GAG GAC CGG Glu Asp Arg 35	GAG GCA GTG Glu Ala Val 40	CGC AGA Arg Arg	627
GAA GCG GA Glu Ala Gl 4	u Arg Gln A	GCC CAG GCA Ala Gln Ala 50	CAG TTG GAA Gln Leu Glu	AAA GCA AAG Lys Ala Lys 55	ACA AAG Thr Lys	675
			AAT GTC AGC Asn Val Ser			723
			ATG GCC ATC Met Ala Ile 85			771
			TTT AAC AAT Phe Asn Asn 100			819
			ATC GGA TTC Ile Gly Phe 115			867

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AAA Lys	CTA Leu	GAA Glu 125	AAC Asn	ATG Met	AGG Arg	CTG Leu	CAG Gln 130	CAT His	GAA Glu	CAG Gln	AGA Arg	GCC Ala 135	AAG Lys	CAA Gln	GGG Gly		915
AAA Lys	TTC Phe 140	TAC Tyr	TCC Ser	AGT Ser	AAA Lys	TCA Ser 145	GGA Gly	GGA Gly	AAT Asn	TCA Ser	TCA Ser 150	TCC Ser	AGT Ser	TTG Leu	GGT Gly	-	963
GAC Asp 155	ATA Ile	GTA Val	CCT Pro	AGT Ser	TCC Ser 160	AGA Arg	AAA Lys	TCA Ser	ACA Thr	CCT Pro 165	CCA Pro	TCA Ser	TCT Ser	GCT Ala	ATA Ile 170		1011
GAC Asp	ATA Ile	GAT Asp	GCT Ala	ACT Thr 175	GGC Gly	TTA Leu	GAT Asp	GCA Ala	GAA Glu 180	GAA Glu	AAT Asn	GAT Asp	ATT Ile	CCA Pro 185	GCA Ala		1059
AAC Asn	CAC His	CGC Arg	TCC Ser 190	CCT Pro	AAA Lys	CCC Pro	AGT Ser	GCA Ala 195	AAC Asn	AGT Ser	GTA Val	ACG Thr	TCA Ser 200	CCC Pro	CAC His		1107
TCC Ser	AAA Lys	GAG Glu 205	AAA Lys	AGA Arg	ATG Met	CCC Pro	TTC Phe 210	TTT Phe	AAG Lys	AAG Lys	ACA Thr	GAG Glu 215	CAC His	ACT Thr	CCT Pro		1155
CCG Pro	TAT Tyr 220	GAT Asp	GTG Val	GTA Val	CCT Pro	TCC Ser 225	ATG Met	CGA Arg	CCA Pro	GTG Val	GTC Val 230	CTA Leu	GTG Val	GGC	CCT Pro		1203
TCT Ser 235	CTG Leu	AAG Lys	GGC Gly	TAC Tyr	GAG Glu 240	GTC Val	ACA Thr	GAT Asp	ATG Met	ATG Met 245	CAA Gln	AAA Lys	GCG Ala	CTG Leu	TTT Phe 250		1251
GAT Asp	TTT Phe	TTA Leu	AAA Lys	CAC His 255	AGA Arg	TTT Phe	GAA Glu	GGG Gly	CGG Arg 260	ATA Ile	TCC Ser	ATC Ile	ACA Thr	AGG Arg 265	GTC Val		1299
ACC Thr	GCT Ala	GAC Asp	ATC Ile 270	TCG Ser	CTT Leu	GCC Ala	AAA Lys	CGC Arg 275	TCG Ser	GTA Val	TTA Leu	AAC Asn	AAT Asn 280	CCC Pro	AGT Ser		1347
AAG Lys	CAC His	GCA Ala 285	ATA Ile	ATA Ile	GAA Glu	AGA Arg	TCC Ser 290	AAC Asn	ACA Thr	AGG Arg	TCA Ser	AGC Ser 295	TTA Leu	GCG Ala	GAA Glu		1395
GTT Val	CAG Gln 300	AGT Ser	GAA Glu	ATC Ile	GAA Glu	AGG Arg 305	ATT Ile	TTT Phe	GAA Glu	CTT Leu	GCA Ala 310	Arg	ACA Thr	TTG Leu	CAG Gln		1443
TTG Leu 315	Val	GTC Val	CTT Leu	GAC Asp	GCG Ala 320	GAT Asp	ACA Thr	ATT	AAT Asn	CAT His 325	Pro	GCT Ala	CAA Gln	CTC Leu	AGT Ser 330		1491
AAA Lys	ACC	TCC	TTG Leu	GCC Ala 335	Pro	ATT Ile	ATA Ile	GTA Val	TAT Tyr 340	Val	AAG Lys	ATT	TCT Ser	TCT Ser 345	CCT Pro		1539

AAG Lys	GTT Val	TTA Leu	CAA Gln 350	Arg	TTA Leu	ATA Ile	AAA Lys	TCT Ser 355	CGA Arg	GGG Gly	AAA Lys	TCT Ser	CAA Gln 360	GCT Ala	AAA Lys	1587
CAC His	CTC Leu	AAC Asn 365	GTC Val	CAG	ATG Met	GTA Val	GCA Ala 370	GCT Ala	GAT Asp	AAA Lys	CTG Leu	GCT Ala 375	CAG Gln	TGT Cys	CCT Pro	1635
CCA Pro	GAG Glu 380	CTG Leu	TTC Phe	GAT Asp	GTG Val	ATC Ile 385	TTG Leu	GAT Asp	GAG Glu	AAC Asn	CAG Gln 390	CTT Leu	GAG Glu	GAT Asp	GCC Ala	1683
TGT Cys 395	GAG Glu	CAC His	CTT Leu	GCC Ala	GAC Asp 400	TAT Tyr	CTG Leu	GAG Glu	GCC Ala	TAC Tyr 405	TGG Trp	AAG Lys	GCC Ala	ACC Thr	CAT His 410	1731
CCT Pro	CCC Pro	AGC Ser	AGT Ser	AGC Ser 415	CTC Leu	CCC Pro	AAC Asn	CCT Pro	CTC Leu 420	CTT Leu	AGC Ser	CGT Arg	ACA Thr	TTA Leu 425	GCC Ala	1779
ACT Thr	TCA Ser	AGT Ser	CTG Leu 430	CCT Pro	CTT Leu	AGC Ser	CCC Pro	ACC Thr 435	CTA Leu	GCC Ala	TCT Ser	AAT Asn	TCA Ser 440	CAG Gln	GGT Gly	1827
TCT Ser	CAA Gln	GGT Gly 445	GAT Asp	CAG Gln	AGG Arg	ACT Thr	GAT Asp 450	CGC Arg	TCC Ser	GCT Ala	CCT Pro	ATC Ile 455	CGT Arg	TCT Ser	GCT Ala	187 5
TCC Ser	CAA Gln 460	GCT Ala	GAA Glu	GAA Glu	GAA Glu	CCT Pro 465	AGT Ser	GTG Val	GAA Glu	CCA Pro	GTC Val 470	AAG Lys	AAA Lys	TCC Ser	CAG Gln	1923
CAC His 475	CGC Arg	TCT Ser	TCC Ser	TCC Ser	TCA Ser 480	GCC Ala	CCA Pro	CAC His	CAC His	AAC Asn 485	CAT His	CGC Arg	AGT Ser	GGG Gly	ACA Thr 490	1971
AGT Ser	CGC Arg	GGC Gly	CTC Leu	TCC Ser 495	AGG Arg	CAA Gln	GAG Glu	ACA Thr	TTT Phe 500	GAC Asp	TCG Ser	GAA Glu	ACC Thr	CAG Gln 505	GAG Glu	2019
AGT Ser	CGA Arg	GAC Asp	TCT Ser 510	GCC Ala	TAC Tyr	GTA Val	GAG Glu	CCA Pro 515	AAG Lys	GAA Glu	GAT Asp	TAT Tyr	TCC Ser 520	CAT His	GAC Asp	2067
CAC His	GTG Val	GAC Asp 525	CAC His	TAT Tyr	GCC Ala	TCA Ser	CAC His 530	CGT Arg	GAC Asp	CAC His	AAC Asn	CAC His 535	AGA Arg	GAC Asp	GAG Glu	2115
ACC Thr	CAC His 540	GGG Gly	AGC Ser	AGT Ser	GAC Asp	CAC His 545	AGA Arg	CAC His	AGG Arg	Glu	TCC Ser 550	CGG Arg	CAC His	CGT Arg	TCC Ser	2163
CGG Arg 555	GAC Asp	GTG Val	GAT Asp	CGA Arg	GAG Glu 560	CAG Gln	GAC Asp	CAC His	AAC Asn	GAG Glu 565	TGC Cys	AAC Asn	AAG Lys	CAG Gln	CGC Arg 570	2211

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														GAA Glu 585			2259
														GTT Val			2307
ATC (TGAG	TTTT	rgc (CTTI	TGTG	T TI	TTTT	TTTT	TTI	TTTT	TTGA				2356
AGTC'	TTGT	'AT	ACTA	ACAC	C A	rcccc	AAAA:	CAA	DAAA	TCT	TTGG	GGTC	TA (CACTO	CAA!	rc	2416
ATAT	GTGA	TC 1	rgtci	TGT	A T	ATTTI	GTAI	TAT	TGCI	GTT	GCTI	'GAA'I	TAG (CAATA	GCA!	r G	2476
GATA	GAGI	'AT I	rgaga	TACT	T T	TCTI	TTGI	AAG	TGCI	ACA	TAAT	TTGG	CC :	rggt <i>i</i>	TGG	CT	2536
GCAG'	TCCI	CC G	GTTG	CATA	C TO	GACI	CTTC	: AAA	AACI	GTT	TTGG	GTAG	CT (GCCAC	TTG	A.A.	2596
CAAA	ATCI	GT 1	rgcca	CCCI	AG GT	rgate	TTAG	TGI	TTTT	AGA	DTAA	TAGI	TG 2	ATGTA	TCC	AA	2656
CAAG	CCAG	C AA	CAGO	CACAC	A T	AAAA	GTGG	TAA S	TTCI	TGT	TTCT	CCAG	TA	TTTT?	ATA	CG	2716
TTAA!	TACG	CA C	GCAI	CTG	TT	rgcai	ATTO	ATI	CATO	GAC	CACI	GTTI	CT :	rgcti	GTA	CC	2776
TCTG	GCTG	SAC 1	LAAA 1	TTG	G G	ACAGA	TTCA	GTO	TTGC	CTT	ACAC	AAA:	GG (GATCA	AAT	AG	2836
TTAG	AATC	TA I	TTTC	TATO	A TE	CTAGI	ACTO	TG1	TACTO	TAT	AGAC	'AGT'I	TG :	raaat	GTT	TA	2896
TTCT	GCAA	AC A	AAACA	CCTC	C T	TTTAT	TATA	A TAA	XTAT!	TAT	ATA	CATA	CA (STTTC	ATC	AC	2956
ACTA'	TTTI	AG A	AGTC														2970

(2) INFORMATION FOR SEQ ID NO:38:

- (i) SEQUENCE CHARACTERISTICS:
 - (A) LENGTH: 2712 base pairs
 - (B) TYPE: nucleic acid
 - (C) STRANDEDNESS: double (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: DNA (genomic)
- (ix) FEATURE:
 - (A) NAME/KEY: CDS
 - (B) LOCATION: 223..2061
 - (D) OTHER INFORMATION: /standard_name= "Beta-2E"
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:38:

AGTGTGTGTT TTCAGCCCCT CCTGGAATGG GAAAATAAGA ATCTCCCTGG ATGGGAGTCC TCTGGGGCAG GGAGTGAAAG CCCCGGAGGC AGAAAGGGAC GGAGAACAGG GGCTTGCCCA

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GAG	CATG	GAT	AGGA	AAGG	AG C	TGGG	GTTC	T CC	GGGG	CTCA	GCG	CGC	CTG	AGAA	CCTGTG	;	18
CCC	GGGG	CTG	CAGC	TGCG	GA C	GATA	AAGG	C GC	TGTC	TGGC				GCC Ala		2	234
TGG Trp 5	ATC Ile	AGG Arg	CTT Leu	CTG Leu	AAA Lys 10	Arg	GCC Ala	AAG Lys	GGA Gly	GGA Gly 15	AGG Arg	CTG Leu	AAC Lys	AAT Asn	TCT Ser 20	2	282
GAT Asp	ATC Ile	TGT Cys	GGT Gly	TCG Ser 25	GCA Ala	GAC Asp	TCC Ser	TAC Tyr	ACT Thr 30	Ser	CGT Arg	CCA Pro	TCC Ser	GAT Asp 35	TCC	3	330
GAT Asp	GTA Val	TCT Ser	CTG Leu 40	Glu	GAG Glu	GAC Asp	CGG Arg	GAG Glu 45	GCA Ala	GTG Val	CGC Arg	AGA Arg	GAA Glu 50	Ala	GAG Glu	3	378
CGG Arg	CAG Gln	GCC Ala 55	CAG Gln	GCA Ala	CAG Gln	TTG Leu	GAA Glu 60	AAA Lys	GCA Ala	AAG Lys	ACA Thr	AAG Lys 65	Pro	GTT Val	GCA Ala	4	126
TTT Phe	GCG Ala 70	GTT Val	CGG Arg	ACA Thr	AAT Asn	GTC Val 75	AGC Ser	TAC Tyr	AGT Ser	GCG Ala	GCC Ala 80	His	GAA Glu	GAT Asp	GAT Asp	4	74
GTT Val 85	CCA Pro	GTG Val	CCT Pro	GGC Gly	ATG Met 90	GCC Ala	ATC Ile	TCA Ser	TTC Phe	GAA Glu 95	GCA Ala	AAA Lys	GAT Asp	TTT Phe	CTG Leu 100	5	22
CAT His	GTT Val	AAG Lys	GAA Glu	AAA Lys 105	TTT Phe	AAC Asn	AAT Asn	GAC Asp	TGG Trp 110	TGG Trp	ATA Ile	GGG Gly	CGA Arg	TTG Leu 115	GTA Val	5	70
AAA Lys	GAA Glu	GGC Gly	TGT Cys 120	GAA Glu	ATC Ile	GGA Gly	TTC Phe	ATT Ile 125	CCA Pro	AGC Ser	CCA Pro	GTC Val	AAA Lys 130	CTA Leu	GAA Glu	6	18
AAC Asn	ATG Met	AGG Arg 135	CTG Leu	CAG Gln	CAT His	GAA Glu	CAG Gln 140	AGA Arg	GCC Ala	AAG Lys	CAA Gln	GGG Gly 145	AAA Lys	TTC Phe	TAC Tyr	6	66
TCC Ser	AGT Ser 150	AAA Lys	TCA Ser	GGA Gly	GGA Gly	AAT Asn 155	TCA Ser	TCA Ser	TCC Ser	AGT Ser	TTG Leu 160	GGT Gly	GAC Asp	ATA Ile	GTA Val	7	14
				AAA Lys												7	62
GCT Ala	ACT Thr	GGC Gly	TTA Leu	GAT Asp 185	GCA Ala	GAA Glu	GAA Glu	AAT Asn	GAT Asp 190	ATT Ile	CCA Pro	GCA Ala	AAC Asn	CAC His 195	CGC Arg	8.	10
TCC	CCT	AAA	CCC	AGT	GCA	AAC	AGT	GTA	ACG	TCA	CCC	CAC	TCC	AAA	GAG	8	58

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			200					205					210				
AAA Lys	AGA Arg	ATG Met 215	CCC Pro	TTC Phe	TTT Phe	AAG Lys	AAG Lys 220	ACA Thr	GAG Glu	CAC His	ACT Thr	CCT Pro 225	CCG Pro	TAT Tyr	GAT Asp	-	906
GTG Val	GTA Val 230	CCT Pro	TCC Ser	ATG Met	CGA Arg	CCA Pro 235	GTG Val	GTC Val	CTA Leu	GTG Val	GGC Gly 240	CCT Pro	TCT Ser	CTG Leu	AAG Lys		954
GGC Gly 245	TAC Tyr	GAG Glu	GTC Val	ACA Thr	GAT Asp 250	ATG Met	ATG Met	CAA Gln	AAA Lys	GCG Ala 255	CTG Leu	TTT Phe	GAT Asp	TTT Phe	TTA Leu 260		1002
AAA Lys	CAC His	AGA Arg	TTT Phe	GAA Glu 265	GGG Gly	CGG Arg	ATA Ile	TCC Ser	ATC Ile 270	ACA Thr	AGG Arg	GTC Val	ACC Thr	GCT Ala 275	GAC Asp		1050
ATC Ile	TCG Ser	CTT Leu	GCC Ala 280	AAA Lys	CGC Arg	TCG Ser	GTA Val	TTA Leu 285	AAC Asn	AAT Asn	CCC Pro	AGT Ser	AAG Lys 290	CAC His	GCA Ala		1098
ATA Ile	ATA Ile	GAA Glu 295	AGA Arg	TCC Ser	AAC Asn	ACA Thr	AGG Arg 300	TCA Ser	AGC Ser	TTA Leu	GCG Ala	GAA Glu 305	GTT Val	CAG Gln	AGT Ser		1146
GAA Glu	ATC Ile 310	GAA Glu	AGG Arg	ATT Ile	TTT Phe	GAA Glu 315	CTT Leu	GCA Ala	AGA Arg	ACA Thr	TTG Leu 320	CAG Gln	TTG Leu	GTG Val	GTC Val		1194
CTT Leu 325	GAC Asp	GCG Ala	GAT Asp	ACA Thr	ATT Ile 330	AAT Asn	CAT His	CCA Pro	GCT Ala	CAA Gln 335	CTC Leu	AGT Ser	AAA Lys	ACC Thr	TCC Ser 340		1242
TTG Leu	GCC Ala	CCT Pro	ATT Ile	ATA Ile 345	GTA Val	TAT Tyr	GTA Val	AAG Lys	ATT Ile 350	TCT Ser	TCT Ser	CCT Pro	AAG Lys	GTT Val 355	TTA Leu		1290
CAA Gln	AGG Arg	TTA Leu	ATA Ile 360	AAA Lys	TCT Ser	CGA Arg	GGG Gly	AAA Lys 365	TCT Ser	CAA Gln	GCT Ala	AAA Lys	CAC His 370	CTC Leu	AAC Asn		1338
GTC Val	CAG Gln	ATG Met 375	GTA Val	GCA Ala	GCT Ala	GAT Asp	AAA Lys 380	Leu	GCT Ala	CAG Gln	TGT Cys	CCT Pro 385	PIO	GAG Glu	CTG Leu		1386
TTC Phe	GAT Asp 390	Val	ATC Ile	TTG Leu	GAT Asp	GAG Glu 395	Asn	CAG Gln	CTT Leu	GAG Glu	GAT Asp 400	Ala	TGT Cys	GAG Glu	CAC His		1434
CTT Leu 405	Ala	GAC Asp	TAT	CTG Leu	GAG Glu 410	Ala	TAC Tyr	TGG	AAG Lys	GCC Ala 415	Thi	CAT His	CCT Pro	CCC	AGC Ser 420		1482
AGT Ser	AGC Ser	CTC	CCC Pro	AAC Asn	CCT Pro	CTC	CTT Leu	AGC Ser	CGI	ACA Thr	TTA Lev	GCC	ACT Thr	TCA Ser	AGT Ser		1530

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				425					430					435		
CTG Leu	CCT	CTT Leu	AGC Ser 440	Pro	ACC Thr	CTA Leu	GCC Ala	TCT Ser 445	Asn	TCA Ser	CAG Gln	GGT Gly	TCT Ser 450	Gln	GGT Gly	1578
GAT Asp	CAG Gln	AGG Arg 455	ACT Thr	GAT Asp	CGC	TCC Ser	GCT Ala 460	Pro	ATC Ile	CGT Arg	TCT	GCT Ala 465	TCC Ser	CAA Gln	GCT Ala	1626
GAA Glu	GAA Glu 470	GIU	CCT	AGT Ser	GTG Val	GAA Glu 475	CCA Pro	GTC Val	AAG Lys	AAA Lys	TCC Ser 480	CAG Gln	CAC His	CGC Arg	TCT Ser	1674
485	ser	ser	ATA	Pro	H1S 490	CAC His	Asn	His	Arg	Ser 495	Gly	Thr	Ser	Arg	Gly 500	1722
Leu	ser	Arg	GIn	G1u 505	Thr	TTT Phe	Asp	Ser	Glu 510	Thr	Gln	Glu	Ser	Arg 515	Asp	1770
ser	ATA	Tyr	520	Glu	Pro	AAG Lys	Glu	Asp 525	Tyr	Ser	His	Asp	His 530	Val	Asp	1818
HIS	ıyr	535	Ser	His	Arg	GAC Asp	His 540	Asn	His	Arg	Asp	Glu 545	Thr	His	Gly	1866
ser	550	Asp	His	Arg	His	AGG Arg 555	Glu	Ser	Arg	His	Arg 560	Ser	Arg	Asp	Val	1914
565	Arg	Glu	GIN	qaA	His 570	AAC Asn	Glu	Cys	Asn	Lys 575	Gln	Arg	Ser "	Arg	His 580	1962
Lys	Ser	Lys	Asp	Arg 585	Tyr	TGT Cys	Glu	Lys	Asp 590	Gly	Glu	Val	Ile	Ser 595	Lys	2010
AAA Lys :	CGG Arg	Asn	GAG Glu 600	GCT Ala	GGG Gly	GAG Glu	TGG Trp	AAC Asn 605	AGG Arg	GAT Asp	GTT Val	Tyr	ATC Ile 610	CCC Pro	CAA Gln	2058
TGAG'	TTTT	GC C	CTTT	TGTG	T TT	TTTT	TTTT	TTT	TTTT	TGA	AGTC	TTGT.	A TA	ACTA	ACAGC	2118
ATCC	CCAA	AA C	AAAA	AGTC	T TT	GGGG	TCTA	CAC	TGCA	ATC	TATA	GTGA	TC T	GTCT	TGTAA	2178
TATT'	TTGT	AT T	ATTG	CTGT	T GC	TTGA	ATAG	CAA	TAGC	ATG	GATA	GAGT	AT T	GAGA	TACTT	2238
TTTC:	TTTT	GT A	agtg	CTAC	A TA	TTAA	GGCC	TGG	TATG	GCT	GCAG	TCCT	CC G	GTTG	CATAC	2298
TGGA	CTCT	TC A	AAAA	CTGT	т тт	GGGT:	AGCT	GCC	ACTT	GAA	CAAA	ATCT	GT T	GCCA	CCCAG	2358

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GTGATGTTAG	TGTTTTAAGA	AATGTAGTTG	ATGTATCCAA	CAAGCCAGAA	TCAGCACAGA	2418
TAAAAAGTGG	AATTTCTTGT	TTCTCCAGAT	TTTTAATACG	TTAATACGCA	GGCATCTGAT	2478
TTGCATATTC	ATTCATGGAC	CACTGTTTCT	TGCTTGTACC	TCTGGCTGAC	TAAATTTGGG	2538
GACAGATTCA	GTCTTGCCTT	ACACAAAGGG	GATCATAAAG	TTAGAATCTA	TTTTCTATGT	2598
ACTAGTACTG	TGTACTGTAT	AGACAGTTTG	TAAATGTTAT	TTCTGCAAAC	AAACACCTCC	2658
ТТАТТАТАТА	ТААТАТАТАТ	ATATATATCA	GTTTGATCAC	ACTATTTTAG	AGTC	2712

WHAT IS CLAIMED IS:

- 1. An isolated DNA fragment, comprising a sequence of nucleotides that encodes an α_1 subunit selected from the group consisting of $\alpha_{1\text{A-1}}$, $\alpha_{1\text{A-2}}$, $\alpha_{1\text{E-1}}$, $\alpha_{1\text{C-2}}$ and $\alpha_{1\text{E-3}}$.
- 5 2. The DNA fragment of claim 1, wherein the α_1 subunit is α_{1A-1} or α_{1A-2} .
 - 3. The DNA fragment of claim 1, wherein the α_1 subunit is $\alpha_{\text{1E-1}}$ or $\alpha_{\text{1E-3}}$
- 4. The DNA fragment of claim 1, wherein the α_1 subunit 10 is $\alpha_{1\text{C-}2\text{.}}$
 - 5. An isolated DNA fragment, comprising a sequence of nucleotides that encodes a β subunit selected from the group consisting of β_2 , β_3 and β_4 .
- 6. The DNA fragment of claim 5, wherein the subunit is a β_{2C} , β_{2D} or β_{2E} subunit.
 - 7. The DNA fragment of claim 5, wherein the subunit is a β_3 subunit.
 - 8. The DNA fragment of claim 7, wherein the subunit is a β_{3-1} subunit.
- 9. The DNA fragment of claim 5, wherein the subunit is a β_4 subunit.
 - 10. The DNA fragment of claim 9, wherein the subunit has an amino acid sequence set forth in SEQ ID No. 28.
- 11. A eukaryotic cell, comprising heterologous DNA that encodes an α_1 subunit selected from the group of subunits consisting of α_{1A-1} , α_{1A-2} , α_{1C-2} , α_{1E-1} , and α_{1E-3} .
 - 12. A eukaryotic cell, comprising heterologous DNA that encodes an α_1 subunit and heterologous DNA that encodes a β subunit, wherein at least one subunit is selected from the group of subunits consisting of α_{1A-1} , α_{1A-2} , α_{1C-2} , α_{1E-1} , α_{1E-3} , β_{2C} , β_{2D} , β_{2E} , β_{3-1} , a β_4 subunit.
 - 13. The eukaryotic cell of claim 12, wherein the β subunit is a β_2 subunit.
- 14. The eukaryotic cell of claim 12, wherein the β 35 subunit is a β_4 subunit.

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- 15. The eukaryotic cell of claim 11, selected from the group consisting of HEK 293 cells, Chinese hamster ovary cells, African green monkey cells, and mouse L cells.
- 16. The eukaryotic cell of claim 12 selected from the group consisting of HEK 293 cells, Chinese hamster ovary cells, African green monkey cells, and mouse L cells.
 - 17. A eukaryotic cell with a functional, heterologous calcium channel, produced by a process comprising:

introducing into the cell heterologous nucleic acid that 10 encodes an α_1 -subunit of a human calcium channel, wherein:

the α_1 subunit is selected from the group consisting of α_{1A-1} , α_{1A-2} , α_{1C-2} , α_{1E-1} and α_{1E-3} ;

the heterologous calcium channel contains at least one subunit encoded by the heterologous nucleic acid; and

the only heterologous ion channels are calcium channels.

18. A eukaryotic cell with a functional, heterologous calcium channel, produced by a process comprising:

introducing into the cell nucleic acid that encodes an α_1 subunit of a human calcium channel and introducing into the cell nucleic acid that encodes a β subunit of a human calcium channel, wherein:

at least one of the subunits is s elected from the group consisting of $\alpha_{\text{1A-1}}$, $\alpha_{\text{1A-2}}$, $\alpha_{\text{1E-1}}$, $\alpha_{\text{1E-3}}$, β_{2C} , β_{2D} , β_{2E} , a β_{3} and a β_{4} subunit;

the heterologous calcium channel contains at least one subunit encoded by the heterologous nucleic acid; and

the only heterologous ion channels are calcium channels.

- 19. The eukaryotic cell of claim 17 selected from the group consisting of HEK 293 cells, Chinese hamster ovary cells, African green monkey cells, mouse L cells and amphibian oöcytes.
- 20. The eukaryotic cell of claim 18 selected from the group consisting of HEK 293 cells, Chinese hamster ovary cells, African green monkey cells, mouse L cells and amphibian 35 oöcytes.

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- 21. The eukaryotic cell of claim 18, wherein the β subunit is a β_2 , β_3 or β_4 subunit of a human calcium channel.
- 22. The eukaryotic cell of claim 18, wherein the calcium channel includes an α_{2b} subunit of a human calcium channel, an α_{1B-1} subunit of a human calcium channel and a β_3 subunit of a human calcium channel.
- 23. The eukaryotic cell of claim 18, wherein the calcium channel includes an α_{1B-1} , α_{2b} , and a β_{1-2} subunit, or an α_{1B-1} , α_{2b} , and a β_{1-3} subunit, or an α_{1B-2} , α_{2b} , and a β_{1-3} subunit, or an α_{1B-2} , α_{2b} , and a β_{3-1} subunit, or a α_{1B-1} , α_{2b} , and an β_{3-1} subunit.
- 24. The eukaryotic cell of claim 18, wherein the calcium channel contains an α_{2b} subunit of a human calcium channel, an α_{1B} or an α_{1D} subunit of a human calcium channel and a β_{1-1} , β_{1-2} or β_{1-3} subunit of a human calcium channel.
- 25. A method for identifying a compound that modulates the activity of a calcium channel, comprising;

suspending a eukaryotic cell that has a functional, heterologous calcium channel, in a solution containing the compound and a calcium channel-selective ion:

depolarizing the cell membrane of the cell; and detecting the current flowing into the cell, wherein:

the heterologous calcium channel includes at least one 5 human calcium channel subunit encoded by DNA or RNA that is heterologous to the cell;

at least one subunit is selected from the group consisting of α_{1A-1} , α_{1A-2} , α_{1E-1} , α_{1E-3} , α_{1C-2} , β_{2C} , β_{2D} , β_{2E} , a β_3 subunit and a β_4 subunit;

the current that is detected is different from that produced by depolarizing the same or a substantially identical cell in the presence of the same calcium channel selective ion but in the absence of the compound.

26. The method of claim 25, wherein the heterologous DNA or RNA encodes a β_3 subunit.

- 27. The method of claim 26, wherein the heterologous DNA or RNA encodes a β_4 subunit.
- 28. A subunit-specific antibody selected from the group consisting of antibodies that bind to an α subunit type or α subunit subtype of a human calcium channels, wherein the subunit is an α_1 subunit.
- 29. The antibody of claim 28, wherein antibody is subtype specific and the α_1 subunit is α_{1A} , α_{1E} and α_{1B} .
- 30. An RNA or single-stranded DNA probe of at least 16 bases in length comprising at least 16 substantially contiguous bases from nucleic acids that encode a subunit of a human calcium channel selected from the group of subunits consisting of α_{1A-1} , α_{1A-2} , α_{1E-1} , α_{1C-2} , α_{1E-3} , β_{3-1} , β_{2C} , β_{2D} , β_{2E} and β_4 .
- 15 31. The probe of claim 30 that contains at least 30 bases that are from nucleic acids that encode a subunit of a human calcium channel selected from the group of subunits consisting of α_{1A-1} , α_{1A-2} , α_{1E-1} , α_{1C-2} , α_{1E-3} , β_{3-1} , β_{2C} , β_{2D} , β_{2E} and β_4 subunits.
- 20 32. A method for identifying nucleic acids that encode a human calcium channel subunit, comprising hybridizing under conditions of at least low stringency a probe of claim 30 to a library of nucleic acid fragments, and selecting hybridizing fragments.
- 25 33. A method for identifying cells or tissues that express a calcium channel subunit-encoding nucleic acid, comprising hybridizing under conditions of at least low stringency a probe of claim 30 with mRNA expressed in the cells or tissues or cDNA produced from the mRNA, and thereby identifying cells or tissue that express mRNA that encodes the subunit.
 - 34. A substantially pure human calcium channel subunit selected from the group consisting of α_{1A-1} , α_{1A-2} , α_{1E-1} , α_{1C-2} , α_{1E-3} , β_{3-1} , β_{2C} , β_{2D} , β_{2E} and β_{4} .

INTERNATIONAL SEARCH REPORT

Intern al Application No

PCT/US 94/09230 A. CLASSIFICATION OF SUBJECT MATTER IPC 6 C12N15/12 C12N5/ C07K16/28 G01N33/50 C12Q1/68 C12N5/10 C07K14/705 According to International Patent Classification (IPC) or to both national classification and IPC **B. FIELDS SEARCHED** Minimum documentation searched (classification system followed by classification symbols) C12Q CO7K C12N GO1N IPC 6 Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched Electronic data base consulted during the international search (name of data base and, where practical, search terms used) C. DOCUMENTS CONSIDERED TO BE RELEVANT Relevant to claim No. Citation of document, with indication, where appropriate, of the relevant passages Category * 1,4,5,7, WO,A,93 04083 (THE SALK INSTITUTE X 11,17, BIOTECHNOLOGY/INDUSTRIAL ASSOCIATES, INC.) 25,26, 4 March 1993 28-32,34 see page 8, line 12 - page 10, line 33 see page 11, line 10 - page 12, line 9 see page 12, line 15 - page 13, line 18 see page 23, line 13 - page 26, line 3 see page 29, line 31 - page 30, line 3 see page 31, line 28 - page 32, line 15 see page 33, line 13 - line 30 see page 35, line 4 - line 32 see page 37, line 3 - line 16 see page 28, line 25 - page 44, line 34 see page 59, line 11 - line 23 see SEQ ID NO. 3 see SEQ ID NO. 10 -/--Patent family members are listed in annex. Further documents are listed in the continuation of box C. X Special categories of cited documents: "T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the "A" document defining the general state of the art which is not considered to be of particular relevance invention "X" document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to "E" earlier document but published on or after the international filing date involve an inventive step when the document is taken alone "L" document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another "Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such document. citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or ments, such combination being obvious to a person skilled other means document published prior to the international filing date but later than the priority date claimed in the art. "&" document member of the same patent family Date of mailing of the international search report Date of the actual completion of the international search

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